

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
25 January 2001 (25.01.2001)

PCT

(10) International Publication Number
WO 01/05831 A1

- (51) International Patent Classification⁷: C07K 14/705, C12N 5/10, 15/12, 15/63
- (21) International Application Number: PCT/US00/19342
- (22) International Filing Date: 14 July 2000 (14.07.2000)
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data:
09/354,147 16 July 1999 (16.07.1999) US
- (71) Applicant (*for all designated States except US*): YALE UNIVERSITY [US/US]; Room 210, 155 Whitney Avenue, P.O. Box 208336, New Haven, CT 06520-8336 (US).
- (72) Inventors; and
- (75) Inventors/Applicants (*for US only*): DIB-HAJJ, Sulayman [US/US]; Room 210, 155 Whitney Avenue, P.O. Box 208336, New Haven, CT 06520-8336 (US). WAXMAN, Stephen, G. [US/US]; 155 Whitney Avenue, P.O. Box 208336, New Haven, CT 06520-8336 (US).
- (74) Agent: TUSCAN, Michael, S.; Morgan, Lewis & Bockius LLP, 1800 M. Street, N.W., Washington, DC 20036-5869 (US).
- (81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.
- (84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).
- Published:
— With international search report.
- For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.*



WO 01/05831 A1

(54) Title: MODULATION OF HUMAN SODIUM CHANNELS IN DORSAL ROOT GANGLIA

(57) Abstract: A novel human tetrodotoxin resistant sodium channel is described, along with isolated nucleic acid molecules that encode this channel. Methods for identifying agents that modulate the Na⁺ current through the channel are provided, as well as related therapeutic and diagnostic methods.

MODULATION OF HUMAN SODIUM CHANNELS IN DORSAL ROOT GANGLIA

FIELD OF THE INVENTION

The present invention relates to a novel human tetrodotoxin-resistant sodium channel and related nucleotides, as well as screening assays for identifying agents useful in treating
5 acute or chronic pain or other hyperexcitability states. This application is related to U.S. Provisional Application 60/072,990, filed January 29, 1998; U.S. Provisional Application 60/109,402 filed November 20, 1998; U.S. Provisional Application 60/109,666 filed on November 20, 1998 ; PCT International Application PCT/US99/02008 filed January 29, 1999 and U.S. Patent Application 09/354,147 filed July 16, 1999, all of which are herein
10 incorporated by reference in their entirety.

BACKGROUND

A. Sodium Channels

Voltage-gated sodium channels are a class of specialized protein molecules that act as molecular batteries permitting excitable cells (neurons and muscle fibers) to produce and
15 propagate electrical impulses. Voltage-gated Na⁺ channels from rat brain are composed of three subunits, the pore-forming α subunit (260 KDa) and two auxiliary subunits, β 1 (36 KDa) and β 2 (33 KDa) that may modulate the properties of the α -subunit; the α subunit is sufficient to form a functional channel that generates a Na current flow across the membrane (Catterall, (1993) Trends Neurosci. 16, 500-506; Isom *et al.*, (1994) Neuron 12, 1183-1194).
20 Nine distinct α subunits have been identified in vertebrates and are encoded by members of an expanding gene family (Goldin (1995) Handbook of receptors and channels (North, editor) CRC Press; Akopian *et al.*, (1996) Nature 379, 257-262; Akopian *et al.*, (1997) FEBS Lett. 400, 183-187; Sangameswaran *et al.*, (1996) J. Biol. Chem. 271, 5953-5956) and respective orthologues of a number of them have been cloned from various mammalian species including
25 humans. Specific α subunits are expressed in a tissue- and developmentally-specific manner (Beckh *et al.*, (1989) EMBO J. 8, 3611-3616; Mandel, (1992) J. Membr. Biol. 125, 193-205).

Aberrant expression patterns or mutations of voltage-gated sodium channel α -subunits underlie a number of human and animal disorders (Roden & George, (1997) *Am. J. Physiol.* 273, H511-H525; Ptacek, (1997) *Neuromuscul. Disord.* 7, 250-255; Cannon, (1997) *Neuromuscul. Disord.* 7; 241-249; Cannon, (1996) *Trends Neurosci.* 19, 3-10); Rizzo *et al.*,
5 (1996) *Eur. Neurol.* 36, 3-12).

Voltage-gated sodium channel α -subunits consist of four domains (D1-4) of varying internal homology but of similar predicted structure, connected by three intracellular loops (L1-3). The four domains fold to form a channel that opens to both the cytoplasm and the extracellular space via a pore. The pore opens and closes depending upon the physiological
10 state of the cell membrane.

Each domain consists of six transmembrane segments (S1-6) that allow the protein to weave through the membrane with intra- and extracellular linkers. The linkers of S5-S6 segments of the four domains contain sequences that line the pore of the channel, and a highly conserved subset of amino acids that acts as a filter to selectively allow sodium ions to
15 traverse the channel pore into the cytoplasm, thus generating an electric current. The amphipathic S4 segment, in each of the four domains, rich in basic residues repeated every third amino acid, acts as a voltage sensor and undergoes a conformational change as a result of the change in the voltage difference across the cell membrane. This in turn triggers the conformational change of the protein to open its pore to the extracellular Na^+ ion gradient.

20 In most of the known voltage-gated sodium channel α -subunits the channels close and change into an inoperable state quickly (inactivate) within a few milliseconds after opening of the pore (activation); SNS-type channels, on the other hand, inactivate slowly and require a greater voltage change to activate. L3, the loop that links domains D3 and D4, contains a tripeptide which acts as an intracellular plug that closes the pore after activation, thus inducing
25 the channel to enter the inactive state. After inactivation, these channels further undergo conformational change to restore their resting state and become available for activation. This period is referred to as recovery from inactivation (repriming). Different channels reprime at different rates, and repriming in SNS is relatively rapid.

Based on amino acid similarities, the voltage-gated sodium channel family has been further subdivided into two subfamilies (Felipe *et al.*, (1994) J. Biol. Chem. 269, 30125-30131). Eight of the nine cloned channels belong to subfamily 1. They share many structural features, particularly in their S4 transmembrane segments. However, some of them
5 have been shown to have distinct kinetic properties of inactivation and repriming. Only a single channel of subfamily 2, also referred to as atypical channels, has been identified in human, rat and mouse tissues. This subfamily is primarily characterized by reduced numbers of basic residues in its S4 segments, and thus is predicted to have different voltage-dependence compared to subfamily 1. The physiological function of subfamily 2
10 channels is currently unknown because its electrophysiological properties have not yet been elucidated.

The blocking of voltage-gated sodium channels by tetrodotoxin, a neurotoxin, has served to functionally classify these channels into sensitive (TTX-S) and resistant (TTX-R) phenotypes. Two mammalian TTX-R channels have so far been identified, one specific to the
15 cardiac muscle and to very limited areas of the central nervous system (CNS) and the second, SNS, is restricted to peripheral neurons (PNS) of the dorsal root ganglia (DRG) and trigeminal ganglia. Specific amino acid residues that confer resistance or sensitivity to TTX have been localized to the ion selectivity filter of the channel pore. The SNS channel is also described in International Patent Application WO 97/01577.

20 **B. Role of Sodium Channels in Disease States**

Because different Na⁺ channel α -subunit isotypes exhibit different kinetics and voltage-dependence, the firing properties of excitable cells depend on the precise mixture of channel types that they express. Mutants of the cardiac and skeletal muscle α -subunit have been shown to cause a number of muscle disorders. Some examples are as follows: A change
25 of a single basic amino acid residue in the S4 of the skeletal muscle channel is sufficient to change the kinetic properties of this channel and induce a disease state in many patients. A tripeptide deletion in L3 of the cardiac channel, proximal to the inactivation gate, induces a cardiac disorder called Long QT syndrome. A single amino acid change in the S5-S6 linker of

domain 1 of Scn8a, the region lining the pore of the channel, causes the mouse mutant “jolting”. The total loss of this channel by a different mutation causes motor end plate “med” disease in mice. This mutation is characterized by loss of motor neuron stimulation of the innervated muscle.

5 **C. Sodium Channels and Pain**

Axonal injury (injury to nerve fibers, also called axons) can produce chronic pain (termed neuropathic pain). A number of studies have demonstrated altered excitability of the neuronal cell body and dendrites after axonal injury (Eccles *et al.*, (1958) J. Physiol. 143: 11-40; Gallego *et al.*, (1987) J. Physiol. (Lond) 391, 39-56; Kuno & Llinas, (1970) J. Physiol. 10 (Lond.) 210, 807-821), and there is evidence for a change in Na⁺ channel density over the neuronal cell body and dendrites following axonal injury (Dodge & Cooley, (1973) IBM J. Res. Dev. 17, 219-229; Titmus & Faber (1986) J. Neurophysiol. 55, 1440-1454; Sernagor *et al.*, (1986) Proc. Natl. Acad. Sci. USA 83, 7966-7970). The expression of abnormal mixtures of different types of sodium channels in a neuronal cell can also lead to abnormal firing (Rizzo 15 *et al.*, (1996) Eur. Neurol. 36, 3-12), and can contribute to hyperexcitability, paresthesia or pain.

Recent studies on rat sensory DRG neurons have demonstrated a dramatic change in the expression profile of TTX-R and TTX-S currents and in a number of mRNA transcripts that could encode the channels responsible for these currents in DRG neurons following 20 various insults (Rizzo *et al.*, (1995) Neurobiol. Dis. 2: 87-96; Cummins *et al.*, (1997) J. Neurophysiol. 17, 3503-3514; Dib-Hajj *et al.*, (1996) Proc. Natl. Acad. Sci. USA 93, 14950-14954). For example, it has been shown an attenuation of the slowly inactivating, TTX-R current and simultaneous enhancement of the rapidly inactivating, TTX-S Na⁺ currents in identified sensory cutaneous afferent neurons following axotomy (Rizzo *et al.*, (1995) 25 Neurobiol. Dis. 2, 87-96). A loss of TTX-S, slowly repriming current and TTX-R current and a gain in TTX-S, rapidly repriming current in nociceptive (pain) neurons following axotomy (Cummins & Waxman (1997) J. Neurophysiol. 17, 3503-3514), down-regulation of SNS transcripts and a simultaneous up-regulation of α -III Transcripts has also been shown

(Dib-Hajj *et al.*, (1996) Proc. Natl. Acad. Sci. USA 93, 14950-14954). Also associated with axotomy is a moderate elevation in the levels of αI and αII mRNAs (Waxman *et al.*, (1994) J. Neurophysiol. 72, 466-470). These changes in the sodium channel profile appear to contribute to abnormal firing that underlies neuropathic pain that patients suffer following axonal injury.

5 Inflammation, which is also associated with pain (termed inflammatory pain), also causes alteration in the sodium current profile in nociceptive DRG neurons. Inflammatory modulators up-regulate TTX-R current in small C-type nociceptive DRG neurons in culture (Gold *et al.*, (1996) Proc. Natl. Acad. Sci. USA 93, 1108-1112; England *et al.*, (1996) J. Physiol. 495, 429-440). The rapid action of these modulators suggests that their action
10 include posttranslational modification of existing TTX-R channels. It has now been determined that inflammation also increases a TTX-R Na^+ current and up-regulates SNS transcripts in C-type DRG neurons (Tanaka *et al.*, (1998) Neuroreport. 9, 967-972). This data suggests that changes in the sodium current profile contribute to inflammation evoked-pain.

D. Therapies for Chronic Pain:

15 A variety of classes of drugs (anticonvulsants such as phenytoin and carbamazepine; anti-arrhythmics such as mexitine; local anesthetics such as lidocaine) act on Na^+ channels. Since the various Na^+ channels produce sodium currents with different properties, selective blockade or activation (or other modulation) of specific channel subtypes is expected to be of significant therapeutic value. Moreover, the selective expression of certain α -subunit isoforms
20 (PN1, SNS, NaN) in specific types of neurons provides a means for selectively altering their behavior.

 Nociceptive neurons of the DRG are the major source of the PNS TTX-R Na^+ current. Thus, the Na^+ channels producing TTX-R currents provide a relatively specific target for the manipulation of pain-producing neurons. The molecular structure of one TTX-R
25 channel in these DRG neurons, SNS, has been identified but, prior to our research, it has not been determined whether there are other TTX-R channels in these neurons. If such channels could be identified, they would be ideal candidates as target molecules that are preferentially expressed in nociceptive neurons, and whose modulation would attenuate pain transmission.

SUMMARY OF THE INVENTION

The present invention includes an isolated nucleic acid which encodes a voltage gated Na⁺ channel that is preferentially expressed in dorsal root ganglia or trigeminal ganglia (the NaN channel). (In our preceding U.S. Provisional Application 60/072,990, this NaN channel
5 was referred to by its previous name "NaX"). In a preferred embodiment, the isolated nucleic acid comprises the sequence shown in Figure 1 (SEQ ID NO: 1), Figure 7A (SEQ ID NO: 4), Figure 8A (SEQ ID NO: 6), Figure 11A (SEQ ID NO: 41), allelic variants of said sequences or nucleic acids that hybridize to the foregoing sequences under stringent conditions.

In another embodiment, the invention includes an expression vector comprising an
10 isolated nucleic acid which encodes the voltage gated Na⁺ channel that is preferentially expressed in dorsal root ganglia or trigeminal ganglia either alone or with appropriate regulatory and expression control elements. In a preferred embodiment, the expression vector comprises an isolated nucleic acid having the sequence shown in Figure 1 (SEQ ID NO: 1), Figure 7A (SEQ ID NO: 4), Figure 8A (SEQ ID NO: 6), Figure 11A (SEQ ID NO: 41), allelic
15 variants of said sequences or nucleic acids that hybridize to the foregoing sequences under stringent conditions.

The present invention further includes a host cell transformed with an expression vector comprising an isolated nucleic acid which encodes a voltage gated Na⁺ channel that is preferentially expressed in dorsal root ganglia or trigeminal ganglia with appropriate
20 regulatory and expression control elements. In a preferred embodiment, the expression vector comprises an isolated nucleic acid having the sequence shown in Figure 1 (SEQ ID NO: 1), Figure 7A (SEQ ID NO: 4), Figure 8A (SEQ ID NO: 6), Figure 11A (SEQ ID NO: 41), allelic variants of said sequences or nucleic acids that hybridize to the foregoing sequences under stringent conditions.

25 The present invention also includes an isolated voltage gated Na⁺ channel that is preferentially expressed in dorsal root ganglia or trigeminal ganglia. In a preferred embodiment, the channel has the amino acid sequence of Figure 2 (SEQ ID NO: 3), Figure 7B (SEQ ID NO: 5), Figure 8B (SEQ ID NO: 8) or Figure 11B (SEQ ID NO: 42), or is encoded by a nucleic acid having the sequence shown in Figure 2 (SEQ ID NO: 3), Figure 7B (SEQ ID

NO: 5), Figure 8B (SEQ ID NO: 8) or Figure 11B (SEQ ID NO: 42), allelic variants of said sequences or nucleic acids that hybridize to the foregoing sequences under stringent conditions. Peptide fragments of the channel are also included.

Another aspect of the invention is a method to identify an agent that modulates the
5 activity of the NaN channel, comprising the steps of bringing the agent into contact with a cell that expresses the Na⁺ channel on its surface and measuring depolarization, or any resultant changes in the sodium current. The measuring step may be accomplished with voltage clamp measurements, by measuring depolarization, the level of intracellular sodium or by measuring sodium influx.

10 Another aspect of the invention is a method to identify an agent that modulates the transcription or translation of mRNA encoding the NaN channel. The method comprises the steps of bringing the agent into contact with a cell that expresses the Na⁺ channel on its surface and measuring the resultant level of expression of the Na⁺ channel.

The invention also includes a method to treat pain, paraesthesia and hyperexcitability
15 phenomena in an animal or human subject by administering an effective amount of an agent capable of modulating, such as by inhibiting or enhancing, Na⁺ current flow through NaN channels in DRG or trigeminal neurons. The method may include administering an effective amount of an agent capable of modulating the transcription or translation of mRNA encoding the NaN channel.

20 Another aspect of the invention is an isolated nucleic acid that is antisense to the nucleic acids described above. In a preferred embodiment, the antisense nucleic acids are of sufficient length to modulate the expression of NaN channel mRNA in a cell containing the mRNA.

Another aspect of the invention is a scintigraphic method to image the loci of pain
25 generation or provide a measure the level of pain associated with DRG or trigeminal neuron mediated hyperexcitability in an animal or human subject by administering labeled monoclonal antibodies or other labeled ligands specific for the NaN Na⁺ channel.

Another aspect of the invention is a method to identify tissues, cells and cell types that express the NaN sodium channel. This method comprises the step of detecting NaN on the cell surface, or en route to the cell surface, or the presence of NaN encoding mRNA.

The present invention further includes a method of producing a transformed cell that
5 expresses an exogenous NaN encoding nucleic acid, comprising the step of transforming the cell with an expression vector comprising an isolated nucleic acid having the sequence shown in Figures 1, 7A, 8A or 11A, allelic variants of said sequences or nucleic acids that hybridize to the foregoing sequences under stringent conditions, together with appropriate regulatory and expression control elements. The invention also includes a method of producing
10 recombinant NaN protein, comprising the step of culturing the transformed host under conditions in which the NaN sodium channel or protein is expressed, and recovering the NaN protein.

The invention also includes an isolated antibody specific for the *NaN* channel or polypeptide fragment thereof. The isolated antibody may be labeled.

15 Another aspect of the invention includes a therapeutic composition comprising an effective amount of an agent capable of decreasing rapidly repriming sodium current flow in axotomized, inflamed or otherwise injured DRG neurons or in normal DRG neurons that are being driven to fire at high frequency. The invention also includes a method to treat acute pain or acute or chronic neuropathic or inflammatory pain and hyperexcitability phenomena in
20 an animal or a human patient by administering the therapeutic composition.

The present invention also includes a method to screen candidate compounds for use in treating pain and hyperexcitability phenomena by testing their ability to alter the expression or activity of an NaN channel mRNA or protein in axotomized, inflamed or otherwise injured DRG neurons.

BRIEF DESCRIPTION OF THE DRAWING FIGURES

Figure 1 shows the sequence of the rat *NaN* cDNA (SEQ ID NO: 3).

Figure 2 shows the putative amino acid sequence of the rat *NaN* cDNA (SEQ ID NO: 3). Predicted transmembrane segments of domains I - IV are underlined. The amino acid
5 serine "S" in DI-SS2, implicated in the TTX-R phenotype, is in bold face type.

Figure 3 presents a schematic diagram of predicted secondary structure of the *NaN* α -subunit.

Figure 4 shows the results of RT-PCR analysis for α -*NaN* in extracts of various tissues using *NaN*-specific primers. *NaN* is abundantly expressed in dorsal root and
10 trigeminal ganglia. Low levels of *NaN* are detected in cerebral hemisphere and retina tissues. No detectable *NaN* signal is seen in cerebellum, optic nerve, spinal cord, sciatic nerve, superior cervical ganglia, skeletal muscle, cardiac muscle, adrenal gland, uterus, liver and kidney.

Figure 5 shows the tissue distribution of α -*NaN* by *in situ* hybridization. Trigeminal
15 ganglion neurons show moderate-to-high hybridization signal (A). Dorsal root ganglion neurons show moderate-to-high hybridization signal in small neurons (B). Hybridization signal is attenuated in large neurons (arrow). (C) Sense probe shows no signal in DRG neurons. No hybridization signal is seen in spinal cord, cerebellum and liver (D-F). All tissues are from adult Sprague-Dawley rat (scale bars = 50 micrometer).

20 Figure 6 shows the predicted lengths of domain I amplification products of rat α -subunits and their subunit-specific restriction enzyme profile.

Figure 7 sets forth the (A) nucleotide (SEQ ID NO: 4) and (B) amino acid (SEQ ID NO: 5) sequences of the murine NaN.

Figure 8 is a partial (A) nucleotide sequence (SEQ ID NO: 6) of the human *NaN* and
25 partial (B) amino acid sequence (SEQ ID NO: 8) of the human NaN protein.

Figure 9 shows cultures of DRG neurons obtained from L4/5 ganglia of adult rats that were reacted with antibody to NaN and then processed for immunofluorescent localization. (A-B) NaN immunostaining is prominent within the cell bodies of DRG neurons. (C) NaN is

present in the neuritic outgrowths, as well as the cell bodies, of DRG neurons. Nomarski (D) and fluorescent (D') images of a neuron that does not express NaN protein.

Figure 10 shows the location of *Scn1 la* and related genes on distal mouse chromosome 9. (A) Haplotypes from the Jackson BSS backcross. Black boxes represent C57BL/6J alleles and white boxes represent SPRET/Ei alleles. The number of animals with each haplotype is given below each column. Missing data was inferred from adjacent data when typing was ambiguous. (B) Map of distal chromosome 9 based on data in (A). Positions of *Scn5a* and *Scn10a* from the MGD consensus map and the locations of the human orthologs are indicated. Numbers are cM positions on the consensus map
(<http://www.informatics.jax.org/bin/ccr/index>).

Figure 11 shows the (A) cDNA nucleotide sequence (SEQ ID NO: 41) of the human *NaN* gene spanning the complete open reading frame and (B) sets forth the amino acid sequence (SEQ ID NO: 42) of the full length human NaN protein.

DETAILED DESCRIPTION

The present invention relates to a novel gene that Applicants have discovered, called *NaN*. *NaN* encodes a previously unidentified protein, referred to herein as NaN, that belongs to the α -subunit voltage-gated sodium channel protein family and that produces a TTX-R sodium current. Such channels underlie the generation and propagation of impulses in excitable cells like neurons and muscle fibers. *NaN* is a novel sodium channel, with a sequence distinct from other, previously identified, channels. The preferential expression of *NaN* on sensory, but not other neurons, makes it a very useful target for diagnostic and/or therapeutic uses in relation to acute and/or chronic pain pathologies..

A. Definitions

This specification uses several technical terms and phrases which are intended to have the following meanings:

The phrase "modulate" or "alter" refers to up- or down-regulating the level or activity of a particular receptor, ligand or current flow. For example an agent might modulate Na^+ current flow by inhibiting (decreasing) or enhancing (increasing) Na^+ current flow. Similarly,

an agent might modulate the level of expression of the NaN sodium channel or the activity of the NaN channels that are expressed.

The phrase "sodium current" or "Na⁺ current" means the flow of sodium ions across a cell membrane, often through channels (specialized protein molecules) that are specifically
5 permeable to certain ions, in this case sodium ions.

The phrase "voltage gated" means that the ion channel opens when the cell membrane is in a particular voltage range. Voltage-sensitive sodium channels open when the membrane is depolarized. They then permit Na⁺ ions to flow into the cell, producing further depolarization. This permits the cell to generate electrical impulses (also known as "action
10 potentials").

The phrase "rapidly repriming" means that the currents recover from inactivation more rapidly than do such currents in most other voltage gated sodium channel family members.

The terms "TTX-R" and "TTX-S" means that the flow of current through a cell membrane is, respectively, resistant or sensitive to tetrodotoxin (a neurotoxin produced in
15 certain species) at a concentration of about 100 nM.

The phrase "peripheral nervous system (PNS)" means the part of the nervous system outside of the brain and spinal cord, *i.e.*, the spinal roots and associated ganglia such as dorsal root ganglia (DRG) and trigeminal ganglia, and the peripheral nerves.

The phrase "inhibits Na⁺ current flow" means that an agent has decreased such current
20 flow relative to a control cell not exposed to that agent. A preferred inhibitor will selectively inhibit such current flow, without affecting the current flow of other sodium channels; or it will inhibit Na⁺ current in the channel of interest to a much larger extent than in other channels.

The phrase "enhances Na⁺ current flow" means that an agent has increased such
25 current flow relative to a control cell not exposed to that agent. A preferred agent will selectively increase such current flow, without affecting the current flow of other sodium channels; or it will increase Na⁺ current in the channel of interest to a much larger extent than in other channels.

The phrase “specifically hybridizes” refers to nucleic acids which hybridize under highly stringent or moderately stringent conditions to the nucleic acids encoding the NaN sodium channel, such as the DNA sequence of SEQ ID NO: 1, 4, 6 or 41.

The phrase “isolated nucleic acid” refers to nucleic acids that have been separated from
5 or substantially purified relative to contaminant nucleic acids encoding other polypeptides. “Nucleic acids” refers to all forms of DNA and RNA, including cDNA molecules and antisense RNA molecules.

The phrase “RT-PCR” refers to the process of reverse transcription of RNA (RT) using the enzyme reverse transcriptase, followed by the amplification of certain cDNA templates
10 using the polymerase chain reaction (PCR); PCR requires generic or gene-specific primers and thermostable DNA polymerase, for example, *Taq* DNA polymerase.

The phrase “preferentially expressed” means that voltage gated Na⁺ channel is expressed in the defined tissues in detectably greater quantities than in other tissues. For instance, a voltage gated Na⁺ channel that is preferentially expressed in dorsal root ganglia or
15 trigeminal ganglia is found in detectably greater quantities in dorsal root ganglia or trigeminal ganglia when compared to other tissues or cell types. The quantity of the voltage gated Na⁺ channel may be detected by any available means, including the detection of specific RNA levels and detection of the channel protein with specific antibodies.

B. Characterization of the NaN Sodium Channel

20 The present invention relates to a previously unidentified, voltage-gated sodium channel α -subunit (*NaN*), predicted to be TTX-R, voltage-gated, and preferentially expressed in sensory neurons innervating the body (dorsal root ganglia or DRG) and the face (trigeminal ganglia). The predicted open reading frame (ORF), the part of the sequence coding for the NaN protein molecule, has been determined with the putative amino acid sequence from
25 different species (rat, mouse, human) presented in Figures 2 (SEQ ID NO: 3), 7B (SEQ ID NO: 5), 8B (SEQ ID NO: 8) or 11B (SEQ ID NO: 42).

All of the relevant landmark sequences of voltage-gated sodium channels are present in NaN at the predicted positions, indicating that *NaN* belongs to the sodium channel family.

But NaN is distinct from all other previously identified Na channels, sharing a sequence identity of less than 53% with each one of them. *NaN* is distinct from *SNS*, the only other TTX-R Na⁺ channel subunit that has been identified, until our discovery, in PNS. We have identified and cloned *NaN* without using any primers or probes that are based upon or specific to *SNS*. Moreover, *NaN* and *SNS* share only 47% similarity of their predicted open reading frame (ORF), comparable to the limited similarity of *NaN* to all subfamily 1 members.

The low sequence similarity to existing α -subunits clearly identifies *NaN* as a novel gene, not simply a variant of an existing channel. Sequence variations compared to the other voltage-gated channels indicate that *NaN* may be the prototype of a novel and previously unidentified, third class of TTX-R channels that may possess distinct properties compared to *SNS*. *NaN* and *SNS*, which are present in nociceptive DRG and trigeminal neurons, may respond to pharmacological interventions in different ways. The preferential expression of *NaN* in sensory DRG and trigeminal neurons provides a target for selectively modifying the behavior of these nerve cells while not affecting other nerve cells in the brain and spinal cord. A further elucidation of the properties of *NaN* channels will be important to understand more fully the effects of drugs designed to modulate the function of the "TTX-R" currents which are characteristic of DRG nociceptive neurons and which contribute to the transmission of pain messages, and to abnormal firing patterns after nerve injury and in other painful conditions..

C. NaN Nucleic Acids

Nucleic acid molecules of the invention include the nucleotide sequences set forth in Figures 1, 7A, 8A and 11A as well as nucleotide sequences that encode the amino acid sequences of Figures 2, 7B, 8B and 11B. Nucleic acids of the claimed invention also include nucleic acids which specifically hybridize to nucleic acids comprising the nucleotide sequences set forth in Figures 1, 7A, 8A and 11A, or nucleotide sequences which encode the amino acid sequences of Figures 2, 7B, 8B and 11B. A nucleic acid which specifically hybridizes to a nucleic acid comprising that sequence remains stably bound to said nucleic acid under highly stringent or moderately stringent conditions. Stringent and moderately stringent conditions are those commonly defined and available, such as those defined by

Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press or Ausubel *et al.*, (1995) Current Protocols in Molecular Biology, Greene Publishing. The precise level of stringency is not important, rather, conditions should be selected that provide a clear, detectable signal when specific hybridization has occurred.

5 Hybridization is a function of sequence identity (homology), G+C content of the sequence, buffer salt content, sequence length and duplex melt temperature (T_m) among other variables (see Maniatis *et al.*, (1982) Molecular Cloning, Cold Spring Harbor Press). With similar sequence lengths, the buffer salt concentration and temperature provide useful variables for assessing sequence identity (homology) by hybridization techniques. For
10 example, where there is at least 90 percent homology, hybridization is commonly carried out at 68°C in a buffer salt such as 6×SCC diluted from 20×SSC (see Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press). The buffer salt utilized for final Southern blot washes can be used at a low concentration, *e.g.*, 0.1×SSC and at a relatively high temperature, *e.g.*, 68°C, and two sequences will form a hybrid duplex
15 (hybridize). Use of the above hybridization and washing conditions together are defined as conditions of high stringency or highly stringent conditions. Moderately stringent conditions can be utilized for hybridization where two sequences share at least about 80 percent homology. Here, hybridization is carried out using 6×SSC at a temperature of about 50-55°C. A final wash salt concentration of about 1-3×SSC and at a temperature of about
20 60-68°C are used. These hybridization and washing conditions define moderately stringent conditions.

 In particular, specific hybridization occurs under conditions in which a high degree of complementarity exists between two nucleic acid molecules. With specific hybridization, complementarity will generally be at least about 70%, 75%, 80%, 85%, preferably about
25 90-100%, or most preferably about 95-100%. When referring the human NaN sequence of SEQ ID NO:41 and 42, preferred homologous sequences will typically encode an NaN protein exhibiting at least about 81% amino acid sequence similarity or at least about 75% or 76% sequence identity to SEQ ID NO: 42. A more preferred human NaN sequence will contain a positively changed residue at amino acid 670, preferably an arginine residue.

As used herein, homology or identity is determined by **BLAST** (Basic Local Alignment Search Tool) analysis using the algorithm employed by the programs **blastp**, **blastn**, **blastx**, **tblastn** and **tblastx** (Karlin *et al.*, (1990) Proc. Natl. Acad. Sci. USA 87, 2264-2268 and Altschul, (1993) J. Mol. Evol. 36, 290-300, both of which are herein
5 incorporated by reference) which are tailored for sequence similarity searching. The approach used by the **BLAST** program is to first consider similar segments between a query sequence and a database sequence, then to evaluate the statistical significance of all matches that are identified and finally to summarize only those matches which satisfy a preselected threshold of significance. For a discussion of basic issues in similarity searching of sequence databases
10 (see Altschul *et al.*, Nat. Genet. (1994) 6, 119-129) which is herein incorporated by reference. The search parameters for **histogram**, **descriptions**, **alignments**, **expect** (*i.e.*, the statistical significance threshold for reporting matches against database sequences), **cutoff**, **matrix** and **filter** are at the default settings. The default scoring matrix used by **blastp**, **blastx**, **tblastn**, and **tblastx** is the **BLOSUM62** matrix (Henikoff *et al.*, (1992) Proc. Natl. Acad. Sci. USA 89,
15 10915-10919, herein incorporated by reference). For **blastn**, the scoring matrix is set by the ratios of **M** (*i.e.*, the reward score for a pair of matching residues) to **N** (*i.e.*, the penalty score for mismatching residues), wherein the default values for **M** and **N** are 5 and -4, respectively.

The nucleic acids of the present invention can be used in a variety of ways in accordance with the present invention. For example, they can be used as nucleic acid probes
20 to screen other cDNA and genomic DNA libraries so as to select by hybridization other DNA sequences that encode homologous *NaN* sequences. Contemplated nucleic acid probes could be RNA or DNA labeled with radioactive nucleotides or by non-radioactive methods (for example, biotin). Screening may be done at various stringencies (through manipulation of the hybridization T_m , usually using a combination of ionic strength, temperature and/or presence
25 of formamide) to isolate close or distantly related homologs. The nucleic acids may also be used to generate primers to amplify cDNA or genomic DNA using polymerase chain reaction (PCR) techniques. The nucleic acid sequences of the present invention can also be used to identify adjacent sequences in the genome, for example, flanking sequences and regulatory elements of *NaN*. The nucleic acids may also be used to generate antisense primers or

constructs that could be used to modulate the level of gene expression of NaN. The amino acid sequence may be used to design and produce antibodies specific to NaN that could be used to localize NaN to specific cells and to modulate the function of NaN channels expressed on the surface of cells.

5 Nucleic acids of the invention also include recombinantly prepared altered NaN sequences. For instance, fusion proteins may be prepared with the open reading frames herein disclosed, or functional fragments thereof, and any available fusion protein. Nucleic acid molecules may also be prepared that encode chimeric NaN proteins, for instance, chimeric proteins comprising individual domains from different species. Such chimeric proteins
10 include, but are not limited to, human NaN chimeras containing mouse or rat domains, or mouse or rat chimeras containing human domains. Preferred chimeras include human NaN with a rat or mouse domain surrounding the residue equivalent to amino acid 670 of human NaN.

D. Vectors and Transformed Host Cells

15 The present invention also comprises recombinant vectors containing and capable of replicating and directing the expression of nucleic acids encoding a NaN sodium channel in a compatible host cell. For example, the insertion of a DNA in accordance with the present invention into a vector using enzymes such as T4 DNA ligase, may be performed by any conventional means. Such an insertion is easily accomplished when both the DNA and the
20 desired vector have been cut with the same restriction enzyme or enzymes, since complementary DNA termini are thereby produced. If this cannot be accomplished, it may be necessary to modify the cut ends that are produced by digesting back single-stranded DNA to produce blunt ends, or by achieving the same result by filling in the single-stranded termini with an appropriate DNA polymerase. In this way, blunt-end ligation may be carried out.
25 Alternatively, any site desired may be produced by ligating nucleotide sequences (linkers) onto the DNA termini. Such linkers may comprise specific oligonucleotide sequences that encode restriction site recognition sequences.

Any available vectors and the appropriate compatible host cells may be used (Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press; Ausubel *et al.*, (1995) Current Protocols in Molecular Biology, Greene Publishing). Commercially available vectors, for instance, those available from New England Biolabs,
5 Promega, Stratagene or other commercial sources are included.

The transformation of appropriate cell hosts with an rDNA (recombinant DNA) molecule of the present invention is accomplished by well known methods that typically depend on the type of vector used and host system employed. Frog oocytes can be injected with RNA and will express channels, but in general, expression in a mammalian cell line
10 (such as HEK293 or CHO cells) is preferred. With regard to transformation of prokaryotic host cells, electroporation and salt treatment methods are typically employed (see, for example, Cohen *et al.*, (1972) Proc. Natl. Acad. Sci. USA 69, 2110-2114; and Maniatis *et al.*, (1982) Molecular Cloning - A Laboratory Manual, Cold Spring Harbor Press). With regard to transformation of vertebrate cells with vectors containing rDNAs, electroporation, cationic
15 lipid or salt treatment methods are typically employed (Graham *et al.*, (1973) Virology 52, 456-467; Wigler *et al.*, Proc. Natl. Acad. Sci. USA (1979) 76, 1373-1376).

Successfully transformed cells, *i.e.*, cells that contain an rDNA molecule of the present invention, can be identified by well known techniques. For example, cells resulting from the introduction of an rDNA of the present invention can be cloned to produce single colonies.
20 Cells from those colonies can be harvested, lysed and their DNA content examined for the presence of the rDNA using conventional methods (Southern, (1975) J. Mol. Biol. 98, 503-517) or the proteins produced from the cell assayed via an immunological method. If tags such as green fluorescent protein are employed in the construction of the recombinant DNA, the transfected cells may also be detected *in vivo* by the fluorescence of such molecules by cell
25 sorting.

For transient expression of recombinant channels, transformed host cells for the measurement of Na⁺ current or intracellular Na⁺ levels are typically prepared by co-transfecting constructs into cells such as HEK293 cells with a fluorescent reporter plasmid (such as pGreen Lantern-1, Life Technologies) using the calcium-phosphate precipitation

technique (Ukomadu *et al.*, (1992) Neuron 8, 663-676). HEK293 cells are typically grown in high glucose DMEM (Life Technologies) supplemented with 10% fetal calf serum (Life Technologies). After forty-eight hours, cells with green fluorescence are selected for recording (Dib-Hajj *et al.*, (1997) FEBS Lett. 416, 11-14).

- 5 For preparation of cell lines continuously expressing recombinant channels, the *NaV* construct is cloned into other vectors that carry a selectable marker in mammalian cells. Transfections are carried out using the calcium phosphate precipitation technique (Ukomadu *et al.*, (1992) Neuron 8, 663-676). Human embryonic kidney (HEK-293), chinese hamster ovary (CHO) cells, derivatives of either or other suitable cell lines are grown under standard
- 10 tissue culture conditions in Dulbecco's modified Eagle's medium supplemented with 10% fetal bovine serum. The calcium phosphate-DNA mixture is added to the cell culture medium and left for 15-20 hours, after which time the cells are washed with fresh medium. After 48 hours, antibiotic (G418) is added to select for cells which have acquired neomycin resistance. After 2-3 weeks in G418, 10-20 isolated cell colonies are harvested using sterile 10 ml pipette
- 15 tips. Colonies are grown for another 4-7 days, split and subsequently tested for channel expression using whole-cell patch-clamp recording techniques and RT-PCR.

E. Method of Measuring Na⁺ Current Flow

- Na⁺ currents are measured using patch clamp methods (Hamill *et al.*, (1981) Pflügers Arch. 391, 85-100), as described by Rizzo *et al.*, (1994) J. Neurophysiol. 72, 2796-2815 and
- 20 Dib-Hajj *et al.*, (1997) FEBS Lett. 416, 11-14. For these recordings data are acquired on a MacIntosh Quadra 950 or similar computer, using a program such as Pulse (v 7.52, HEKA, German). Fire polished electrodes typically (0.8-1.5 MW) are fabricated from capillary glass using a Sutter P-87 puller or a similar instrument. In the most rigorous analyses, cells are usually only considered for analysis if initial seal resistance is <5 Gohm, they have high
- 25 leakage currents (holding current <0.1 nA at -80 mV), membrane blebs, and an access resistance <5 Mohm. Access resistance is usually monitored throughout the experiment and data are not used if resistance changes occur. Voltage errors are minimized using series resistance compensation and the capacitance artifact is canceled using computer controlled

amplifier circuitry or other similar methods. For comparisons of the voltage dependence of activation and inactivation, cells with a maximum voltage error of ± 10 mV after compensation are used. Linear leak subtraction is usually used for voltage clamp recordings. Membrane currents are typically filtered at 5 KHz and sampled at 20 KHz. The pipette solution contains
5 a standard solution such as: 140 mM CsF, 2 mM MgCl, 1 mM EGTA, and 10 mM Na-HEPES (pH 7.3). The standard bathing solution is usually 140 mM NaCl, 3 mM KCl, 2 mM MgCl₂, 1 mM CaCl₂, 10 mM HEPES, and 10 mM glucose (pH 7.3).

Voltage clamp studies on transformed cells or DRG neurons, using methods such as intracellular patch-clamp recordings, can provide a quantitative measure of the sodium current
10 density (and thus the number of sodium channels in a cell), and channel physiological properties. These techniques, which measure the currents that flow through ion channels such as sodium channels, are described in Rizzo *et al.*, (1995) Neurobiol. Dis. 2, 87-96. Alternatively, the blockage or enhancement of sodium channel function can be measured using optical imaging with sodium-sensitive dyes or with isotopically labeled Na. These
15 methods which are described in Rose *et al.*, (1997) J. Neurophysiol. 78, 3249-3258 and by (Kimelberg & Waltz, (1988) The Neuronal Microenvironment (Boulton *et al.*, editors) Humana Press), measure the increase in intracellular concentration of sodium ions that occurs when sodium channels are open.

F. Measurement of Intracellular Sodium [Na⁺]

20 The effects of various agents on cells that express Na⁺ can be determined using ratiometric imaging of [Na⁺]_i using SBFI or other similar ion-sensitive dyes. In this method, as described by Sontheimer *et al.*, (1994) J. Neurosci. 14, 2464-2475, cytosolic-free Na⁺ is measured using an indicator for Na⁺, such as SBFI (sodium-binding benzofuran isophthalate (Harootunian *et al.*, (1989) J. Biol. Chem. 264, 19458-19467)); or a similar dye. Cells are first loaded with the
25 membrane-permeable acetoxymethyl ester form of the dye (which is dissolved in dimethyl sulfoxide (DMSO) at a stock concentration of 10 mM). Recordings are obtained on the stage of a microscope using a ratiometric imaging setup (*e.g.*, from Georgia Instruments). Excitation light is provided at appropriate wavelengths (*e.g.*, 340:385 nm). Excitation light is

passed to the cells through a dichroic reflector (400 nm) and emitted light above 450 nm is collected. Fluorescence signals are amplified, *e.g.*, by an image intensifier (GenIISyS) and collected with a CCD camera, or similar device, interfaced to a frame grabber. To account for fluorescence rundown, the fluorescence ratio 340:385 is used to assay cytosolic-free Na^+ .

- 5 For calibration of SBFI's fluorescence, cells are perfused with calibration solutions containing known Na^+ concentrations (typically 0 and 30 mM, or 0, 30, and 50 mM $[\text{Na}^+]_i$), and with ionophores such as gramicidin and monensin (see above) after each experiment. As reported by Rose & Ransom, (1996) *J. Physiol. (Lond)* 491, 291-305, the 345/390 nm fluorescence ratio of intracellular SBFI changes monotonically with changes in $[\text{Na}^+]_i$.
- 10 Experiments are typically repeated on multiple (typically at least four) different coverslips, providing statistically significant measurements of intracellular sodium in control cells, and in cells exposed to various concentrations of agents that may block, inhibit or enhance Na^+ .

G. Method to Measure Na^+ Influx via Measuring ^{22}Na or ^{86}Rb

- ^{22}Na is a gamma emitter and can be used to measure Na^+ flux (Kimelberg & Waltz, 15 (1988) *The Neuronal Microenvironment* (Boulton *et al.*, editors) Humana Press), and $^{86}\text{Rb}^+$ can be used to measure Na^+/K^+ -ATPase activity (Sontheimer *et al.*, (1994) *J. Neurosci.* 14, 2464-2475). $^{86}\text{Rb}^+$ ions are taken up by the Na^+/K^+ -ATPase-like K^+ ions, but have the advantage of a much longer half-life than $^{42}\text{K}^+$ (Kimelberg & Mayhew (1975) *J. Biol. Chem.* 250, 100-104). Thus, measurement of the unidirectional ouabain-sensitive $^{86}\text{Rb}^+$ uptake 20 provides a quantitative method for assaying Na^+/K^+ -ATPase activity which provides another indicator of the electrical firing of nerve cells. Following incubation of cells expressing *NaV* with the isotope $^{22}\text{Na}^+$, the cellular content of the isotope is measured by liquid scintillation counting or a similar method, and cell protein is determined using a method such as the bicinchoninic acid protein assay (Smith *et al.*, (1985) *Anal. Biochem.* 150, 76-85) following 25 the modifications described by Goldschmidt & Kimelberg (1989) *Anal. Biochem.* 177, 41-45 for cultured cells. ^{22}Na and $^{86}\text{Rb}^+$ fluxes are determined in the presence and absence of agents that may block, inhibit, or enhance *NaV*. This permits determination of the actions of these agents on *NaV*.

H. Method to Identify Agents that Modulate NaN-Mediated Current

Several approaches can be used to identify agents that are able to modulate (*i.e.*, block or augment) the Na⁺ current through the NaN sodium channel. In general, to identify such agents, a model cultured cell line that expresses the NaN sodium channel is utilized, and
5 one or more conventional assays are used to measure Na⁺ current. Such conventional assays include, for example, patch clamp methods, the ratiometric imaging of [Na⁺], and the use of ²²Na and ⁸⁶Rb as described above.

In one embodiment of the present invention, to evaluate the activity of a candidate compound to modulate Na⁺ current, an agent is brought into contact with a suitable
10 transformed host cell that expresses *NaN*. After mixing or appropriate incubation time, the Na⁺ current is measured to determine if the agent inhibited or enhanced Na⁺ current flow.

Agents that inhibit or enhance Na⁺ current are thereby identified. A skilled artisan can readily employ a variety of art-recognized techniques for determining whether a particular agent modulates the Na⁺ current flow.

15 Because Na⁺ is preferentially expressed in pain-signaling cells, one can also design agents that block, inhibit, or enhance Na⁺ channel function by measuring the response of laboratory animals, treated with these agents, to acute, inflammatory or chronic pain. In one embodiment of this aspect of the invention, laboratory animals such as rats are treated with an agent for instance, an agent that blocks or inhibits (or is thought to block or inhibit) NaN. The
20 response to various painful stimuli are then measured using tests such as the tail-flick test and limb withdrawal reflex, and are compared to untreated controls. These methods are described by Dubner, (1994) Textbook of Pain (Wall & Melzack, editors) Churchill Livingstone Publishers. In another embodiment of this aspect of the invention, laboratory animals such as rats are subjected to localized injection of pain-producing inflammatory agents such as
25 formalin (Dubuisson & Dennis (1977) Pain 4, 161-74), Freund's adjuvant (Iadarola *et al.*, (1988) Pain 35, 313-326) or carageenan, or are subjected to nerve constriction (Bennett & Xie, (1988) Pain 33, 87-107; Kim & Chung (1992) Pain 50, 355-363) or nerve transection (Seltzer *et al.*, (1990) Pain 43, 205-218) which produce persistent pain. The response to various normal and painful stimuli are then measured, for example, by measuring the latency to

withdrawal from a warm or hot stimulus (Dubner, (1994) Textbook of Pain (Wall & Melzack, editors) Churchill Livingstone Publishers) so as to compare control animals and animals treated with agents that are thought to modify NaN.

The preferred inhibitors and enhancers of NaN preferably will be selective for the NaN
5 Na⁺ channel. They may be totally specific (like tetrodotoxin, TTX, which inhibits sodium channels but does not bind to or directly effect any other channels or receptors), or relatively specific (such as lidocaine which binds to and blocks several types of ion channels, but has a predilection for sodium channels). Total specificity is not required for an inhibitor or
10 enhancer to be efficacious. The ratio of its effect on sodium channels vs. other channels and receptors, may often determine its effect and effects on several channels, in addition to the targeted one, may be efficacious (Stys *et al.*, (1992) J. Neurophysiol. 67, 236-240).
Modulators of NaN may be combined with or coadministered with agents that modulate other channels expressed in primary sensory neurons, including but not limited to PN1/hNE and SNS/PN3 (Waxman (1999) Pain Supplement 6:S133-140).

15 It is contemplated that modulating agents of the present invention can be, as examples, peptides, small molecules, naturally occurring and other toxins and vitamin derivatives, as well as carbohydrates. A skilled artisan can readily recognize that there is no limit as to the structural nature of the modulating agents of the present invention. Screening of libraries of molecules may reveal agents that modulate NaN or current flow through it. Similarly,
20 naturally occurring toxins (such as those produced by certain fish, amphibians and invertebrates) can be screened. Such agents can be routinely identified by exposing a transformed host cell or other cell which expresses a sodium channel to these agents and measuring any resultant changes in Na⁺ current.

I. Recombinant Protein Expression, Synthesis and Purification

25 Recombinant NaN proteins can be expressed, for example, in *E. coli* strains HB101, DH5a or the protease deficient strain such as CAG-456 and purified by conventional techniques.

The peptide agents of the invention can be prepared using standard solid phase (or solution phase) peptide synthesis methods, as is known in the art. In addition, the DNA encoding these peptides may be synthesized using commercially available oligonucleotide synthesis instrumentation and produced recombinantly using standard recombinant production systems. The production using solid phase peptide synthesis is necessitated if non-gene-encoded amino acids are to be included.

J. Antibodies and Immunodetection

Another class of agents of the present invention are antibodies immunoreactive with the Na⁺ channel. These antibodies may block, inhibit, or enhance the Na⁺ current flow through the channel. Antibodies can be obtained by immunization of suitable mammalian subjects with peptides, containing as antigenic regions, those portions of NaN, particularly (but not necessarily) those that are exposed extracellularly on the cell surface. Such immunological agents also can be used in competitive binding studies to identify second generation inhibitory agents. The antibodies may also be useful in imaging studies, once appropriately labeled by conventional techniques.

K. Production of Transgenic Animals

Transgenic animals containing and mutant, knock-out or modified *NaN* genes are also included in the invention. Transgenic animals wherein both *NaN* and the *SNS/PN3* gene are modified, disrupted or in some form modified are also included in the present invention. Transgenic animals are genetically modified animals into which recombinant, exogenous or cloned genetic material has been experimentally transferred. Such genetic material is often referred to as a "transgene". The nucleic acid sequence of the transgene, in this case a form of *NaN*, may be integrated either at a locus of a genome where that particular nucleic acid sequence is not otherwise normally found or at the normal locus for the transgene. The transgene may consist of nucleic acid sequences derived from the genome of the same species or of a different species than the species of the target animal.

The term "germ cell line transgenic animal" refers to a transgenic animal in which the genetic alteration or genetic information was introduced into a germ line cell, thereby

conferring the ability of the transgenic animal to transfer the genetic information to offspring. If such offspring in fact possess some or all of that alteration or genetic information, then they too are transgenic animals.

The alteration or genetic information may be foreign to the species of animal to which
5 the recipient belongs, foreign only to the particular individual recipient, or may be genetic information already possessed by the recipient. In the last case, the altered or introduced gene may be expressed differently than the native gene.

Transgenic animals can be produced by a variety of different methods including transfection, electroporation, microinjection, gene targeting in embryonic stem cells and
10 recombinant viral and retroviral infection (see, *e.g.*, U.S. Patent No. 4,736,866; U.S. Patent No. 5,602,307; Mullins *et al.*, (1993) Hypertension 22, 630-633; Brenin *et al.*, (1997) Surg. Oncol. 6, 99-110; Tuan (1997) Recombinant Gene Expression Protocols, Humana Press).

A number of recombinant or transgenic mice have been produced, including those which express an activated oncogene sequence (U.S. Patent No. 4,736,866); express simian
15 SV 40 T-antigen (U.S. Patent No. 5,728,915); lack the expression of interferon regulatory factor 1 (IRF-1) (U.S. Patent No. 5,731,490); exhibit dopaminergic dysfunction (U.S. Patent No. 5,723,719); express at least one human gene which participates in blood pressure control (U.S. Patent No. 5,731,489); display greater similarity to the conditions existing in naturally occurring Alzheimer's disease (U.S. Patent No. 5,720,936); have a reduced capacity to
20 mediate cellular adhesion (U.S. Patent No. 5,602,307); possess a bovine growth hormone gene (Clutter *et al.*, (1996) Genetics 143, 1753-1760); or, are capable of generating a fully human antibody response (McCarthy, (1997) Lancet 349, 405).

While mice and rats remain the animals of choice for most transgenic experimentation, in some instances it is preferable or even necessary to use alternative animal species.

25 Transgenic procedures have been successfully utilized in a variety of non-murine animals, including sheep, goats, pigs, dogs, cats, monkeys, chimpanzees, hamsters, rabbits, cows and guinea pigs (see Kim *et al.*, (1997) Mol. Reprod. Dev. 46, 515-526; Houdebine, (1995) Reprod. Nutr. Dev. 35, 609-617; Petters (1994) Reprod. Fertil. Dev. 6, 643-645; Schnieke *et al.*, (1997) Science 278, 2130-2133; Amoah, (1997) J. Animal Sci. 75, 578-585).

The method of introduction of nucleic acid fragments into recombination competent mammalian cells can be by any method which favors co-transformation of multiple nucleic acid molecules. Detailed procedures for producing transgenic animals are readily available to one skilled in the art, including the disclosures in U.S. Patent No. 5,489,743 and U.S. Patent
5 No. 5,602,307.

The specific examples presented below are illustrative only and are not intended to limit the scope of the invention.

EXAMPLES

Example 1: Cloning and Characterization of the Rat *NaN* Coding Sequence

10 a. RNA Preparation

Dorsal root ganglia (DRG) from the lumbar region (L4-L5) were dissected from adult Sprague-Dawley rats and total cellular RNA was isolated by the single step guanidinium isothiocyanate-acid phenol procedure (Chomczynski, (1987) Anal. Biochem. 162, 156-159). For analytical applications, DRG tissues were dissected from a few animals at a time. The
15 quality and relative yield of the RNA was assessed by electrophoresis in a 1% agarose gel. Because of the limited starting material (four DRGs weigh on average 10 mg), quantifying the RNA yield was not attempted. PolyA⁺ RNA was purified from about 300 mg of total DRG RNA (28 animals) using the PolyA⁺Tract isolation system according to the manufacturers recommendations (Promega). Half of the purified RNA was used for the preparation of
20 Marathon cDNA (see below) without further quantification.

b. Reverse Transcription

For analytical applications, first strand cDNA was synthesized essentially as previously described (Dib-Hajj *et al.*, (1996) FEBS Lett. 384, 78-82). Briefly, total RNA was reverse transcribed in a 25 ml final volume using 1mM random hexamer (Boehringer
25 Mannheim) and 500 units SuperScript II reverse transcriptase (Life Technologies) in the presence of 100 units of RNase Inhibitor (Boehringer Mannheim). The reaction buffer

consisted of 50 mM Tris-HCl (pH 8.3), 75 mM KCl, 3 mM MgCl₂, 10 mM DTT and 125 mM dNTP. The reaction was allowed to proceed at 37°C for 90 minutes, 42°C for 30 minutes, then terminated by heating to 65°C for 10 minutes.

c. First-Strand cDNA Synthesis

5 The Marathon cDNA synthesis protocol was followed according to the manufacturer's instruction as summarized below (all buffers and enzymes are purchased from the manufacturer (Clontech):

Combine the following reagents in a sterile 0.5-ml microcentrifuge tube: 1 mg (1-4 ml) PolyA⁺ RNA sample, one ml cDNA Synthesis Primer (10 mM) and sterile water to a final
10 volume of 5 ml. Mix contents and spin the tube briefly in a microcentrifuge. Incubate the mixture at 70°C for two minutes, then immediately quench the tube on ice for two minutes. Touch-spin the tube briefly to collect the condensation. Add the following to each reaction tube: 2 ml 5× First-Strand Buffer, 1 ml dNTP Mix (10 mM), 1 ml [α -³²P]dCTP (1 mCi/ml), 1 ml AMV Reverse Transcriptase (20 units/ml) for a 10 ml volume. The radiolabeled dCTP is
15 optional (used to determine yield of cDNA) and is replaced by sterile H₂O if not used. Mix the contents of the tube by gently pipetting and touch-spin the tube to collect the contents at the bottom. Incubate the mixture at 42°C for one hour in an air incubator to reduce condensation and enhance the yield of the first strand cDNA. Place the tube on ice to terminate first-strand synthesis.

20 d. Second-Strand cDNA Synthesis

Combine the following components in the reaction tube from above: 48.4 ml Sterile water, 16 ml 5× Second-Strand Buffer, 1.6 ml dNTP Mix (10 mM), 4 ml 20× Second-Strand Enzyme Cocktail for an 80 ml total volume. Mix the contents thoroughly with gentle pipetting and spin the tube briefly in a microcentrifuge. Incubate the mixture at 16°C for 1.5
25 hours then add 2 ml (10 units) of T4 DNA Polymerase, mix thoroughly with gentle pipetting and incubate the mixture at 16°C for 45 minutes. Add 4 ml of the EDTA/Glycogen mix to terminate second-strand synthesis. Extract the mixture with an equal volume of buffer-

saturated (pH 7.5) phenol:chloroform:isoamyl alcohol (25:24:1). Mix the contents thoroughly by vortexing and spin the tube in a microcentrifuge at maximum speed (up to 14,000 rpm or 13000×g), 4°C for ten minutes to separate layers. Carefully transfer the top aqueous layer to a clean 0.5-ml tube. Extract the aqueous layer with 100 ml of chloroform:isoamyl alcohol
5 (24:1), vortex, and spin the tube as before to separate the layers. Collect the top layer into a clean 0.5-ml microcentrifuge tube. Ethanol precipitate the double-stranded cDNA by adding one-half volume of 4 M Ammonium Acetate and 2.5 volumes of room-temperature 95% ethanol. Mix thoroughly by vortexing and spin the tube immediately in a microcentrifuge at top speed, room temperature for twenty minutes. Remove the supernatant carefully and wash
10 the pellet with 300 ml of 80% ethanol. Spin the tube as before for 10 minutes and carefully remove the supernatant. Air dry the pellet for up to 10 minutes and dissolve the cDNA in 10 ml of sterile H₂O and store at -20°C. Analyze the yield and size of cDNA by running 2 ml of the cDNA solution on a 1.2% agarose/EtBr gel with suitable DNA size markers (for example, the 1 kilobp ladder, Gibco-BRL). If EtBr staining does not show a signal and [α -³²P]dCTP
15 was included in the reaction, dry the agarose gel on a vacuum gel drying system and expose an x-ray film to the gel overnight at -70°C.

e. Adaptor Ligation

Combine these reagents in a 0.5-ml microcentrifuge test tube, at room temperature, and in the following order: 5 ml double-stranded cDNA, 2 ml Marathon cDNA Adaptor (10
20 mM), 2 ml 5× DNA Ligation Buffer, 1 ml T4 DNA Ligase (1 unit/ml) for a 10 ml final volume. Mix the contents thoroughly with gentle pipetting and spin the tube briefly in a microcentrifuge. Incubate at either: 16°C overnight; or room temperature (19-23°C) for three to four hours. Inactivate the ligase enzyme by heating the mixture at 70°C for five minutes. Dilute 1 ml of this reaction mixture with 250 ml of Tricine-EDTA buffer and use for RACE
25 protocols. Store the undiluted adaptor-ligated cDNA at -20°C for future use.

f. PCR

For the initial discovery of *NaN*, we used generic primers designed against highly conserved sequences in domain 1 (D1) of α -subunits I, II and III and later added more primers to accommodate the new α -subunits that were discovered. Thus, generic primers were used that recognize conserved sequences in all known Na^+ channels. The middle of the amplified region shows significant sequence and length polymorphism (Figure 6) and (Gu *et al.*, (1997) J. Neurophysiol. 77, 236-246; Fjell *et al.*, (1997) Mol. Brain Res. 50, 197-204). Due to codon degeneracy, 4 forward primers (F1-F4) and 3 reverse primers (R1-R3) were designed to ensure efficient priming from all templates that might have been present in the cDNA pool (Table 1); however, any of these primers may bind to multiple templates depending on the stringency of the reaction. Forward primer F1 matches subunits αI , αIII ; αNa6 ; αPN1 ; αm1 , αrH1 and $\alpha\text{SNS/PN3}$. Sequences of individual subunits show 1 or 2 mismatches to this primer: T to C at position 16 and A to G at position 18 (αNa6); C to R at position 6 (αm1); A to G at position 18 (αrH1) and T to C at position 3 (αSNS). Forward primer F2 matches subunit αII . Forward primer F3 perfectly matches αNa6 and also matches αrH1 with a single mismatch of C to T at position 16. Reverse primer R1 matches subunits αI , αII , αIII , αNa6 , αPN1 , αm1 and αrH1 . This primer has mismatches compared to 4 subunits: G to A at position 3, A to G at position 4 and T to G at position 7 (αI); T to C at position 1 and A to G at position 19 (αPN1); G to A at position 3 and A to G at position 7 (αm1); an extra G after position 3, GC to CT at positions 14-15, and A to T at position 21 (αrH1). Reverse primer R2 matches subunit $\alpha\text{SNS/PN3}$.

Table 1: Generic and *NaV*-specific primers used for the identification and cloning of *NaV*. All primers except the marathon primers, were synthesized at the department of Pathology, Program for Critical Technologies in Molecular Medicine, Yale University.

Forward Primers	Reverse Primers
5 1. GACCCRTGGAATTGGTTGGA (SEQ ID NO: 9)	1. CAAGAAGGCCAGCTGAAGGTGTC (SEQ ID NO: 15)
2. AATCCCTGGAATTGGTTGGA (SEQ ID NO: 10)	2. GAGGAATGCCACGCAAAGGAATC (SEQ ID NO: 16)
10 3. GACCCGTGGAAGTGGTTAGA (SEQ ID NO: 11)	3. AAGAAGGGACCAGCCAAAGTTGTC (SEQ ID NO: 17)
4. GATCTTTGGAAGTGGCTTGA (SEQ ID NO: 12)	4. ACYTCCATRCANWCCCACAT (SEQ ID NO: 18)
5. AACATAGTGCTGGAGTTCAGG (SEQ ID NO: 13)	5. AGRAARTCNAGCCARCACCA (SEQ ID NO: 19)
15 6. GTGGCCTTTGGATTCCGGAGG (SEQ ID NO: 14)	6. TCTGCTGCCGAGCCAGGTA (SEQ ID NO: 20)
	7. CTGAGATAACTGAAATCGCC (SEQ ID NO: 21)
Marathon AP-1 CCATCCTAATACGACTCACTATAGGGC (SEQ ID NO: 22)	
Marathon AP-2 ACTCACTATAGGGCTCGAGCGGC (SEQ ID NO: 23)	

The respective mouse atypical sodium channel mNa_v2.3 sequence was used to design forward
 20 primer F4 and reverse primer R3 to amplify the analogous sequence from aNaG, the presumed
 rat homolog of mNa_v2.3 (Felipe *et al.*, (1994) J. Biol. Chem. 269, 30125-301231). The
 amplified sequence was cloned into the *Srf*I site of the vector pCR-Script (Stratagene). The
 nucleotide sequence of this fragment shows 88% identity to the respective sequence of
 mNa_v2.3 (Dib-Hajj & Waxman, unpublished). The restriction enzyme *Xba*I was found to be
 25 unique to this subunit. Recently, the sequence of a full length cDNA clone of putative sodium
 channel, NaG-like (SCL-11:Y09164), subunit was published (Akopian *et al.*, (1997) FEBS

Lett. 400, 183-187). The published sequence is 99% identical to our sequence and confirms the size and restriction enzyme polymorphism of the NaG PCR product.

The predicted lengths of amplified products and subunit-specific restriction enzyme recognition sites are shown in Figure 6. All subunit sequences are based on Genbank database
5 (accession numbers: α I: X03638; α II: X03639; α III: Y00766; α Na6: L39018; α hNE-Na: X82835; α m1 M26643; α rH1 M27902 and α SNS X92184; mNa 2.3 L36719).

Subsequently, amplification of NaN sequences 3' terminal to the aforementioned fragment was achieved using NaN-specific primers and two generic reverse primers, R4 and R5. The sequence of the R4 primer was based on the amino acid sequence MWV/DCMEV (SEQ ID
10 NO: 38) located just N-terminal to domain II S6 segment (see schematic diagram of Figure 3 of voltage-gated sodium channel α -subunits for reference). The sequence of the R5 primer is based on the amino acid sequence AWCWLDFL (SEQ ID NO: 43) which forms the N-terminal portion of domain III S3 segment.

Amplification was typically performed in 60 μ l volume using one μ l of the first
15 strand cDNA, 0.8mM of each primer and 1.75 units of Expand Long Template DNA polymerase enzyme mixture (Boehringer Mannheim). Compared to conventional and thermostable DNA polymerases, Expand Long Template enzyme mixture increases the yield of the PCR products without an increase in non-specific amplification (Barnes, (1994) Proc. Natl. Acad. Sci. USA 91, 2216-2220; Cheng *et al.*, (1994) Proc. Natl. Acad. Sci. USA 91,
20 5695-5699). The PCR reaction buffer consisted of 50 mM Tris-HCl (pH 9.2), 16 mM $(\text{NH}_4)_2\text{SO}_4$, 2.25 mM MgCl₂, 2% (v/v) DMSO and 0.1% Tween 20. As described previously (Dib-Hajj *et al.*, (1996) FEBS Lett. 384, 78-82), amplification was carried out in two stages using a programmable thermal cycler (PTC-200, MJ Research). First, a denaturation step at 94°C for four minutes, an annealing step at 60°C for two minutes and an elongation step at
25 72°C for 90 seconds. Second, a denaturation step at 94°C for one minute, an annealing step at 60°C for one minute and an elongation step at 72°C for 90 seconds. The second stage was repeated 33 times for a total of 35 cycles, with the elongation step in the last cycle extended to ten minutes

Primary RACE amplification was performed in 50 µl final volume using 4 µl diluted DRG marathon cDNA template, 0.2 mM marathon AP-1 and NaN-specific primers, 3.5 units Expand Long Template enzyme mixture. Extension period was adjusted at 1 minute per 800 base pairs based on the expected product. 5' and 3' RACE amplification was performed using primer pairs marathon AP-1/*NaN*-specific R6 and *NaN*-specific F5/marathon AP-1, respectively. The PCR reaction buffer consisted of 50 mM Tris-HCl (pH 9.2), 16 mM (NH₄)₂SO₄, 3.0 mM MgCl₂, 2% (v/v) DMSO and 0.1% Tween 20. Amplification in three stages was performed in a programmable thermal cycler (PTC-200, MJ Research). An initial denaturation step at 92°C was carried out for two minutes. This was followed by 35 cycles consisting of denaturation at 92°C for 20 seconds, annealing step at 60°C for one minute, and an elongation step at 68°C. Finally, an elongation step at 68°C was carried out for five minutes. Nested amplification was performed using 2 µl of a 1/500 diluted primary RACE product in a final volume of 50 µl under similar conditions to the primary RACE reactions. Primer pairs AP-2/*NaN*-specific R7 and *NaN*-specific F6/marathon AP-2 were used for nested 5' and 3' RACE, respectively. Secondary RACE products were band isolated from 1% agarose gels and purified using Qiaex gel extraction kit (Qiagen).

A schematic diagram of the putative structure of *NaN* is shown in Figure 3. The length of the intracellular loops is highly variable both in sequence and length among the various subunits. The exception is the loop between domains III and IV.

20 Example 2: Determination of the Putative Rat Amino Acid Sequence for the NaN Channel

NaN-related clones and secondary RACE fragments were sequenced at the W. M. Keck Foundation Biotechnology Resource Lab, DNA sequencing group at Yale University. Sequence analysis including determination of the predicted amino acid sequence was performed using commercial softwares, Lasergene (DNASTar) and GCG. The putative amino acid sequence of *NaN* is shown in Figure 2. Predicted transmembrane segments of domains I - IV are underlined.

Example 3: Determination of the Murine NaN Sequence

Total RNA extraction from trigeminal ganglia of mice, purification of polyA⁺ RNA, and Marathon cDNA construction were done as previously described for the rat. The initial amplification was performed using rat *NaN* primers. The forward primer corresponds to
5 nucleotides 765-787 of the rat sequence (5' CCCTGCTGCGCTCGGTGAAGAAG 3') (SEQ ID NO: 24), and the reverse primer corresponds to nucleotides 1156-1137 (negative strand) of the rat sequence (5' GACAAAGTAGATCCCAGAGG 3') (SEQ ID NO: 25). The amplification produced a fragment of the expected size. The sequence of this fragment demonstrated high similarity to rat *NaN*. Other fragments were amplified using different rat
10 primers and primers designed based on the new mouse *NaN* sequence that was being produced. Finally, longer fragments were amplified using mouse Marathon cDNA template and mouse *NaN*-specific primers in combination with adaptor primers that were introduced during the Marathon cDNA synthesis. These fragments were sequenced using primer walking and assembled into Figure 7A.

15 Mouse *NaN* nucleotide sequence, like rat *NaN*, lacks the out-of-frame ATG at the -8 position relative to the translation initiation codon ATG at position 41 (Figure 7A). Translation termination codon TGA is at position 5314. A polyadenylation signal (AATAAA) is present at position 5789 and a putative 23 nucleotide polyA tail is present beginning at position 5800. The sequence encodes an ORF of 1765 amino acids (Figure 7B), which is 90%
20 similar to rat *NaN*. The gene encoding *NaN* has been named *Scn11a*.

Chromosomal localization of mouse NaN

A genetic polymorphism between strains C57BL/6J and SPRET/Ei was identified by SSCP analysis of a 274 bp fragment from the 3'UTR of *Scn11a*. Genotyping of 94 animals from the BSS backcross panel (Rowe *et al.*, (1994) Mamm. Genome 5, 253-274)
25 demonstrated linkage of *Scn11a* with markers on distal chromosome 9 (Figure 10). No recombinants were observed between *Scn11a* and the microsatellite marker *D9Mit19*. Comparison of our data with the MGD consensus map of mouse chromosome 9 revealed close linkage of *Scn11a* with the two other TTX-R voltage-gated sodium channels, *Scn5a* (George

et al., (1995) Cytogenet. Cell. Genet. 68, 67-70) and *Scn10a* (Kozak & Sangameswaran, (1996) Mamm. Genome 7, 787-788; Souslova *et al.*, (1997) Genomics 41, 201-209).

Example 4: Determination of a Partial and Complete Human NaN Coding Sequence

Human DRG tissue was obtained from a transplant donor. Total RNA extraction and
5 cDNA synthesis were performed as described previously.

Forward primer corresponds to sequence 310-294 (minus strand) of EST AA446878. The sequence of the primer is 5' CTCAGTAGTTGGCATGC 3' (SEQ ID NO: 26). Reverse primer corresponds to sequence 270-247 (minus strand) of EST AA88521 1. The sequence of the primer is 5' GGAAAGAAGCACGACCACACAGTC 3' (SEQ ID NO: 27). Amplification
10 was performed as previously described. PCR amplification was successful and a 2.1 kbp fragment was obtained. This fragment was gel purified and sent for sequencing by primer walking, similar to what is done for mouse *NaN*. The sequence of the ESTs is extended in both directions; the additional sequence shows highest similarity to rat and mouse *NaN*, compared to the rest of the subunits.

15 The sequence of a human 2.1 kbp fragment was obtained using the PCR forward and reverse primers for sequencing from both ends of the fragment. Two additional primers were used to cover the rest of the sequence. The sequence was then extended in the 5' direction using forward primer 1 (above) and human *NaN* reverse primer (5'-GTGCCGTAAACATGAGACTGTCG3') (SEQ ID NO: 44) near the 5' end of the 2.1 kbp
20 fragment. The partial amino acid sequence is set forth in Figure 8B.

The partial ORF of the human *NaN* consists 1241 amino acids. The sequence is 64% identical to the corresponding sequence of rat *NaN* (73% similar, allowing for conservative substitutions) using the advanced BLAST program available at the National Center for Biotechnology Information. Using the Clustal method of alignment (Lasergene software,
25 DNASTar) the human *NaN* is 68% and 69% similar to mouse and rat *NaN*, respectively. The respective mouse and rat sequences are 88% similar.

Further sequencing revealed the cDNA sequence spanning the full length open reading frame for the human *NaN* gene. This extended sequence is shown in Figure 11A (SEQ ID

NO: 41). In addition to the features noted with regard to the partial cDNA sequence (Figure 8A), notable features of the extended sequence include a translation start codon (ATG) at position 31 and a translation termination codon at position 5400. A recognizable polyadenylation signal has not been observed and presumably is located 3' of the disclosed
 5 sequence. The putative amino acid sequence of the human Nan protein is set forth in Figure 11B (SEQ ID NO: 42).

Example 5: Isolation of an Alternative Splicing Variant of Rat NaN

A rat NaN cDNA that encodes a C-terminal truncated version of the full-length rat NaN in Figures 1 and 2 was isolated by sequencing the insert of a rat cDNA clone. The
 10 variant NaN cDNA encodes an NaN protein lacking the 387 C-terminal amino acids of the full length NaN and containing a novel 94 amino acid stretch at the C-terminal end. The new sequence arises from the use of a cryptic donor splice site in exon 23 and a novel exon 23', which is located in intron 23. The novel C terminal amino acids are: AAGQAMRKQG
 DILGPNIHQF SQSSETPFLG CPQQRTCVSF VRPQRVLRVP WFPWRTVTF
 15 LSRPRSSESS AWLGLVESSG WSGLPGESGP SSSL (SEQ ID NO: 28). The N-terminal amino acids of the truncated variant are identical to amino acids 1-1378 of the full length rat NaN of Figure 2. The alternative exon and the splicing pattern was confirmed by comparing the cDNA sequence and the genomic sequence in the respective region.

20 **Example 6: Methods to Isolate Other NaN Sequences**

a. Isolation of NaN sequences from genomic DNA

The genomic structure of three voltage-gated Na⁺ channel α -subunits have already been determined (George *et al.*, (1993) Genomics 15, 598-606; Souslova *et al.*, (1997) Genomics 41, 201-209; McClatchey *et al.*, (1992) Hum. Mol. Genet. 1, 521-527; Wang *et al.*,
 25 (1996) Genomics 34, 9-16). These genes bear remarkable similarity in their organization and provide a predictable map of most of the exon/intron boundaries. Based on the available rat, mouse and human cDNA sequence of NaN, disclosed herein, PCR primers are designed to amplify NaN homologous sequences from other species using standard PCR protocols.

Alternatively, commercially available genomic DNA libraries are screened with *NaN*-specific probes (based on the rat, mouse, or more preferably, the human sequence) using standard library screening procedures (Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press; Ausubel *et al.*, (1995) Current Protocols in Molecular Biology, Greene Publishing). This strategy yields genomic DNA isolates that can then be sequenced and the exon/intron boundaries determined by homology to the rat, mouse or human cDNA sequence.

b. Isolation of full length *NaN* sequences allelic variants from autopsy or biopsy tissues

For isolation of human ganglia total RNA, a full length *NaN* human cDNA homologue is isolated from human dorsal root ganglia or trigeminal ganglia or other cranial ganglia from post-mortem human material, foetuses or biopsy or surgical tissues. Total ribonucleic acid (RNA) is isolated from these tissues by extraction in guanidinium isothiocyanate (Saiki *et al.*, (1985) Science 230, 1350-1354) as described in Example 1.

For Determination of the full length transcript size of the human homologue of the rat *NaN* sodium channel cDNA, the method of determining transcript size is as described in Example 9.

Example 7: Production of human DRG cDNA library

A cDNA library from human DRG or trigeminal ganglia polyA+ RNA was prepared in Example 4 using standard molecular biology techniques (Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press; Ausubel *et al.*, (1995) Current Protocols in Molecular Biology, Greene Publishing.

PolyA+ mRNA is hybridized to an oligo(dT) primer and the RNA is copied by reverse transcriptase into single strand cDNA. Then, the RNA in the RNA-DNA hybrid is fragmented by RNase H as *E. coli* DNA polymerase I synthesizes the second-strand fragment. The ends of the double stranded cDNA are repaired, linkers carrying specific restriction enzyme site (for example, Eco RI) are ligated to the ends using *E. coli* DNA ligase. The pool

of the cDNA insert is then ligated into one of a variety of bacteriophage vectors that are commercially available like Lambda-Zap (Stratagene). The procedures are summarized in more detail as follows:

a. First strand cDNA Synthesis

- 5 Dissolve 10 mg poly(A) + RNA at a concentration of 1 mg/ml in sterile water. Heat the RNA for two to five minutes at 65-70°C then quench immediately on ice. In a separate tube add in the following order (180 ml total) : 20 ml of 5 mM dNTPs (500 µM final each), 40 ml 5× RT buffer (1×final), 10 ml 200 mM DTT (10 mM final), 20 ml 0.5 mg/ml oligo (dT)₁₂₋₁₈ (50 mg/ml final), 60 ml deionized water, 10 ml (10 units) RNasin (50 units/ml
- 10 final). Mix by vortexing, briefly microcentrifuge, and add the mixture to the tube containing the RNA. Add 20 ml (200 U) AMV or MMLV reverse transcriptase for a final concentration of 1000 units/ml in 200 ml. Mix by pipetting up and down several times and remove 10 ml to a separate tube containing 1 ml of α³²P dCTP. Typically, incubate both tubes at room temperature for five minutes, then place both tubes at 42°C for one and a half hours. This
- 15 radiolabeled aliquot is removed to determine incorporation and permit an estimation of recovery; this reaction is stopped by adding 1 ml of 0.5 M EDTA (pH 8.0) and stored frozen at -20°C. The radiolabeled reaction will be used later to estimate the yield and average size of the cDNA inserts. The main reaction is stopped by adding 4 ml of 0.5 M EDTA (pH 8.0) and 200 ml buffered phenol. The mixture is vortexed well, microcentrifuged at room temperature
- 20 for one minute to separate phases, and the upper aqueous layer is transferred to a fresh tube. Back extract the phenol layer with 1×TE buffer (10 mM Tris, 1 mM EDTA, pH 7.5) and pool the aqueous layers from the two extractions. This back extraction of the phenol layer improves the yield. The cDNA is ethanol precipitated using 7.5 M ammonium acetate (final concentration 2.0 to 2.5 M) and 95% ethanol. Place in dry ice/ethanol bath fifteen minutes,
- 25 warm to 4°C, and microcentrifuge at ten minutes at full speed at 4°C to pellet nucleic acids. The small, yellow-white pellet is then washed with ice-cold 70% ethanol, and microcentrifuged for three minutes at full speed, 4°C. Again, the supernatant is removed and the pellet briefly dried.

b. Second strand synthesis

Typically, the pellet from the first-strand synthesis is resuspended in 284 ml water and these reagents are added in the following order (400 ml total): Four ml of 5 mM dNTPs (50 μ M final each), 80 ml 5 \times second-strand buffer (1 \times final), 12 ml 5 mM β -NAD (150 μ M final), 2 ml 10 uCi/ml α -³²P dCTP (50 μ Ci/ml final). Mix by vortexing, briefly
5 microcentrifuge, and add: 4 ml (4 units) RNase H (10 units/ml final), 4 ml (20 units) *E. coli* DNA ligase (50 units/ml final), and 10 ml (100 units) *E. coli* DNA polymerase I (250 units/ml final). Mix by pipetting up and down, briefly microcentrifuge, and incubate twelve to sixteen hours at 14°C. After second-strand synthesis, remove 4 ml of the reaction to
10 determine the yield from the incorporation of the radiolabel into acid-insoluble material. Extract the second-strand synthesis reaction with 400 ml buffered phenol and back extract the phenol phase with 200 ml TE buffer (pH 7.5) as described above. The double stranded cDNA is then ethanol precipitated as described above.

To complete the second-strand synthesis the double-stranded cDNA ends are
15 rendered blunt using a mixture of enzymes. Resuspend the pellet in 42 ml water then add these reagents in the following order (80 ml total): 5 ml 5 mM dNTPs (310 μ M final each), 16 ml 5 \times TA buffer (1 \times final), 1 ml 5 mM β -NAD (62 μ M final). Mix by vortexing, microcentrifuge briefly, and add: 4 ml of 2 mg/ml RNase A (100 ng/ml final), 4 ml (4 units) RNase H (50 units/ml final), 4 ml (20 units) *E. coli* DNA ligase (250 units/ml final) and 4 ml
20 (8 units) T4 DNA polymerase (100 units/ml final). Mix as above and incubate forty-five minutes at 37°C. Add 120 ml TE buffer (pH 7.5) and 1 ml of 10 mg/ml tRNA. Extract with 200 ml buffered phenol and back extract the phenol layer with 100 ml TE buffer as described above. Pool the two aqueous layers and ethanol precipitate as described above.

c. Addition of linkers to double stranded cDNA

25 Combine these reagents in a 0.5 ml microcentrifuge test tube, at room temperature, and in the following order: 100 ng double stranded cDNA, 2 ml linkers/adaptors (10 mM), 2 ml 5 \times DNA Ligation Buffer, 1 ml T4 DNA Ligase (unit/ml) for a 10 ml final volume. Mix the contents thoroughly with gentle pipetting and spin the tube briefly in a microcentrifuge.

Incubate at either: 16°C overnight; or room temperature (19-23°C) for three to four hours. Inactivate the ligase enzyme by heating the mixture at 70°C for five minutes. This cDNA is typically digested by *Eco* RI to prepare the cohesive ends of the cDNA for ligation into the vector and to cleave linker concatemers. Typically this reaction consists of the 10 ml of the
5 cDNA, 2 ml of 10× *Eco* RI buffer (depending on the company of source), 2 ml of *Eco* RI (10 units/ml) and sterile water to a final volume of 20 ml. The mixture is incubated at 37°C for two to four hours.

d. Size fractionation of cDNA

Size exclusion columns are typically used to remove linker molecules and short
10 cDNA fragments (350 bp). For example, a 1-ml Sepharose CL-4B column is prepared in a plastic column plugged with a small piece of sterilized glass wool (a 5 ml plastic pipet will work fine). The column is equilibrated with 0.1 M sodium chloride in 1×TE (10mM Tris, 1 mM EDTA, pH 7.5). The cDNA is then loaded onto the column and 200 µl fractions are collected. 2 µl aliquots of each fraction are analyzed by gel electrophoresis and
15 autoradiography to determine the peak of cDNA elution. Typically, fractions containing the first half of the peak are pooled and purified by ethanol precipitation and resuspending in 10 µl distilled water.

e. Cloning of cDNA into bacteriophage vector

Bacteriophage vectors designed for the cloning and propagation of cDNA are
20 provided ready-digested with *Eco* RI and with phosphatased ends from commercial sources (e.g., lambda gt10 from Stratagene). The prepared cDNA is ligated into lambda vectors following manufacturer's instructions. Ligated vector/cDNA molecules are packaged into phage particles using packaging extracts available commercially.

Example 8: Screening of Human cDNA Library

a. Labeling of cDNA fragments (probes) for library screening

An RNA probe is used that recognizes nucleotide sequences that are specific to *NaN*, such as 1371-1751 of *NaN*. Other nucleotide sequences can be developed on the basis of the *NaN* sequence (Figures 2, 7 & 8) such as nucleotides 765-1160 of the human nucleotide sequence. A *Hind* III/*Bam* HI fragment of *NaN* was inserted in pBluescript (SK+) vector (Stratagene). The sequence of the resulting construct was verified by sequencing. The orientation of the insert is such that the 5' and 3' ends of the construct delineated by the *Hind* III and *Bam* HI restriction enzyme sites, respectively, are proximal to T7 and T3 RNA polymerase promoters, respectively. Digoxigenin-labeled Sense (linearized at the *Hind* III site and transcribed by T7 RNA polymerase) and antisense (linearized at the *Bam* HI site and transcribed by T3 RNA polymerase) transcripts were prepared in vitro using MEGAscript transcription kit (Ambion) according to manufacturer specifications. Briefly, 1 µg linearized template was transcribed with the respective RNA polymerase in a 20 µl final volume containing the following reagents: 1× enzyme mixture containing the respective RNA polymerase and RNase inhibitor and reaction buffer (Ambion), 7.5 mM ATP, GTP and CTP nucleotides, 5.625 mM UTP and 1.725 mM Dig-11UTP (Boehringer Mannheim). *In vitro* transcription was carried out at 37°C for three hours in a water bath. DNA template was removed by adding 1 µl of RNase-free DNase I (2 units/µl) to each reaction and incubating further at 37°C for fifteen minutes. The reaction was then stopped by adding 30 µl nuclease-free water) and 25 µl of LiCl precipitation solution (7.5 M Lithium Chloride, 50 mM EDTA).

The mixture was incubated at -20°C for thirty minutes. The RNA transcripts were pelleted in a microfuge at 13000×g, 4°C for fifteen minutes. The supernatant was removed and the pellet washed once with 100 µl of 75% ethanol. The mixture was re-centrifuged at 13000×g, room temperature for five minutes. The pellet was then air-dried in a closed chamber and subsequently dissolved in 100 µl of RNase-free water. The transcript yield and integrity were determined by comparison to a control DIG-labeled RNA on agarose-formaldehyde gel as described in the DIG/Genius kit according to manufacturer

recommendations (Boehringer Mannheim). Alternatively, a skilled artisan can design radioactive probes for autoradiographic analysis.

Other regions of the rat, mouse or human *NaN* sodium channel cDNA, like 3' untranslated sequences, can also be used as probes in a similar fashion for cDNA library screening or Northern blot analysis. Specifically, a probe is made using commercially available kits, such as the Pharmacia oligo labeling kit, or Genius kit (Boehringer Mannheim).

b. cDNA library screening

Recombinant plaques containing full length human homologues of the *NaN* sodium channel are detected using moderate stringency hybridization washes (50-60°C, 5×SSC, thirty minutes), using non-radioactive (see above) or radiolabeled DNA or cRNA *NaN*-specific probes derived from the 3' untranslated or other regions as described above. Libraries are screened using standard protocols (Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press; Ausubel *et al.*, (1995) Current Protocols in Molecular Biology, Greene Publishing) involving the production of nitrocellulose or nylon membrane filters carrying recombinant phages. The recombinant DNA is then hybridized to *NaN*-specific probes (see above). Moderate stringency washes are carried out.

Plaques which are positive on duplicate filters (*i.e.*, not artefacts or background) are selected for further purification. One or more rounds of screening after dilution to separate the phage are typically performed. Resulting plaques are then purified, DNA is extracted and the insert sizes of these clones characterized. The clones are cross-hybridized to each other using standard techniques (Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press) and distinct positive clones identified.

Typically, overlapping clones that encode the channel are isolated. Standard cloning techniques are then used to produce a full length cDNA construct that contains any 5' untranslated sequence, the start codon ATG, the coding sequence, a stop codon and any 3' untranslated sequence, a poly A consensus sequence and possibly a poly A run. If overlapping clones do not produce sufficient fragments to assemble a full length cDNA clone, alternative

methods like RACE (PCR-based) could be used to generate the missing pieces or a full length clone.

c. Characterization of a human homologue full-length clone

- A *NaN*-specific cDNA sequence from a full-length clone is used as a probe in
- 5 Northern blot analysis to determine the messenger RNA size in human tissue for comparison with the rat and mouse messenger RNA size. Confirmation of biological activity of the cloned cDNA is carried out using methods similar to those described for the rat *NaN*.

Example 9: Polymerase chain reaction (PCR) approaches to clone other full length human *NaN* sodium channels using DNA sequences derived from rat, mouse or human amino acid

10 sequences

Total RNA and poly A+ RNA is isolated from human dorsal root ganglia or trigeminal ganglia or other cranial ganglia from post-mortem human material or foetuses or biopsy/surgical tissues as described above. Preparation of cDNA and PCR-based methods are then used as described previously in Example 1.

- 15 Using degenerate PCR primers derived from the rat, mouse or human *NaN*-specific coding sequence (see Figures 2 (SEQ ID NO: 3), 7B (SEQ ID NO: 5), 8B (SEQ ID NO: 8) and 11B (SEQ ID NO: 41)), the cDNA is amplified using the polymerase chain reaction (Saiki *et al.*, (1985) Science 230, 1350-1354). A skilled artisan could utilize the many variables which can be manipulated in a PCR reaction to derive the homologous sequences required.
- 20 These include, but are not limited to, varying cycle and step temperatures, cycle and step times, number of cycles, thermostable polymerase, and Mg^{2+} concentration. A greater specificity can be achieved using nested primers derived from further conserved sequences from the *NaN* sodium channel.

- Amplification is typically performed in 60 μ l volume using 1 μ l of the first strand
- 25 cDNA, 0.8 mM of each primer and 1.75 units of Expand Long Template DNA polymerase enzyme mixture (Boehringer Mannheim). Compared to conventional and thermostable DNA polymerases, Expand Long Template enzyme mixture increases the yield of the PCR products

without an increase in non-specific amplification (Barnes, (1994) Proc. Natl. Acad. Sci. USA 91, 2216-2220; Cheng *et al.*, (1994) Proc. Natl. Acad. Sci. USA 91, 5695-5699). The PCR reaction buffer consists of 50 mM Tris-HCl (pH 9.2), 16 mM $(\text{NH}_4)_2\text{SO}_4$, 2.25 mM MgCl₂, 2% (v/v) DMSO and 0.1% Tween 20. As described previously (Dib-Hajj *et al.*, (1996) FEBS Lett. 384, 78-82), amplification is carried out in two stages using a programmable thermal cycler (PTC-200, MJ Research). First, a denaturation step at 94°C for four minutes, an annealing step at 60°C for two minutes and an elongation step at 72°C for ninety seconds. Second, a denaturation step at 94°C for one minute, an annealing step at 60°C for one minute and an elongation step at 72°C for ninety seconds. The second stage is repeated 33 times for a total of 35 cycles, with the elongation step in the last cycle extended to 10 minutes. In addition, control reactions are performed alongside the samples. These should be: (1) all components without cDNA, (negative control) and (2) all reaction components with primers for constitutively expressed product, *e.g.*, GAPDH.

The products of the PCR reactions are examined on 1-1.6% (w/v) agarose gels. Bands on the gel (visualized by staining with ethidium bromide and viewing under UV light) representing amplification products of the approximate predicted size are then cut from the gel and the DNA purified.

The resulting DNA may be sequenced directly or is ligated into suitable vectors such as, but not limited to, pCR II (Invitrogen) or pGEMT (Promega). Clones are then sequenced to identify those containing sequence with similarity to the rat, mouse or partial human *NaV* sodium channel sequence.

Example 10: Clone analysis

Candidate clones from Example 9 are further characterized by conventional techniques. The biological activity of expression products is also confirmed using conventional techniques.

Example 11: Isolation of full length *NaN* sequences from human fetal tissues

Commercially available human fetal cDNA libraries and/or cDNA pools are screened with *NaN*-specific primers (by PCR) or probes (library screening) using PCR standard PCR protocols and standard library screening procedures as described above.

5 Example 12: Northern Blot of rat DRG or Trigeminal Neurons with Fragments of Rat *NaN*

10-30 µg total DRG and/or RNA from DRG or trigeminal (for positive tissues) and cerebral hemisphere, cerebellum and liver (for negative tissues) is electrophoresed in denaturing 1% agarose-formaldehyde gel or agarose-glyoxal gel, and then is transferred to a nylon membrane as described in achieved in multiple steps, as detailed in standard molecular biology manuals (Sambrook *et al.*, (1989) Molecular Cloning - A Laboratory Approach, Cold Spring Harbor Press; Ausubel *et al.*, (1995) Current Protocols in Molecular Biology, Greene Publishing). Radiolabeled (specific activity of $>10^8$ dpm/µg) or Digoxigenin-labeled RNA probes are typically used for Northern analysis. An antisense RNA probe (see Example 20, which describes *in situ* hybridization with a *NaN*-specific probe) is created by *in vitro* synthesis from a sense DNA fragment. The membrane carrying the immobilized RNA in wetted with 6×SSC, and the membrane is placed RNA-side-up in a hybridization tube. One ml formamide prehybridization/hybridization solution per 10 cm² of membrane is added. Prehybridization and hybridization are usually carried out in glass tubes in a commercial hybridization oven. The tubes are place in a hybridization oven and incubated, with rotation, at 60°C for fifteen minutes to one hour. The desired volume of probe is pipeted into the hybridization tube, and the incubation is continued with rotation overnight at 60°C. The probe concentration in the hybridization solution should be 10 ng/ml if the specific activity is 10^8 dpm/µg or 2 ng/ml if the specific activity is 10^9 dpm/µg (use 2-10 ng/ml of Digoxigenin labeled probe).

25 The hybridization solution is poured off and an equal volume of 2×SSC/0.1% SDS is added. Incubation with rotation for 5 minutes at room temperature is carried out. The wash solution is changed, and this step is repeated. To reduce background, it may be beneficial to double the volume of the wash solutions. The wash solution is replaced with an equal volume

of 0.2×SSC/0.1% SDS and the tube is incubated for five minutes with rotation at room temperature. The wash solution is changed and this step is repeated (this is a low-stringency wash). For moderate or high stringency conditions, further washes are done with wash solutions pre-warmed to moderate (42°C) or high (68°C) temperatures. The final wash
5 solution is removed and the membrane rinsed in 2×SSC at room temperature. Autoradiography is then performed for up to one week. Alternatively, signal is detected using chemiluminescence technology (Amersham) if non-radioactive probes are used. The transcript size is calculated from the signal from the gel in comparison with gel molecular weight standard markers.

10 Example 13: Tissue specific distribution of *NaN* by RT-PCR

NaN-specific forward (5' CCCTGCTGCGCTCGGTGAAGAA 3') (SEQ ID NO: 39) and reverse primer (5' GACAAAGTAGATCCCAGAGG 3') (SEQ ID NO: 25), were used in RT-PCR assays using cDNA template prepared from multiple rat. These primers amplify *NaN* sequence between nucleotides 765 and 1156 (392 bp) and are *NaN*-specific as judged by
15 lack of similarity to sequences in the database (using programs like BLASTN from the National Center for Biotechnology Information). Amplification was typically performed in a 60 µl volume using 1 µl of the first strand of cDNA, 0.8 µM of each primer and 1.75 units of Expand Long Template DNA polymerase enzyme mixture (Boehringer Mannheim). Compared to conventional and thermostable DNA polymerases, Expand Long Template
20 enzyme mixture increases the yield of the PCR products without an increase in non-specific amplification (Barnes, (1994) Proc. Natl. Acad. Sci. USA 91, 2216-2220; Cheng *et al.*, (1994) Proc. Natl. Acad. Sci. USA 91, 5695-5699). The PCR reaction buffer consisted of 50 mM Tris-HCl (pH 9.2), 16mM (NH₄)₂SO₄, 2.25 mM MgCl₂, 2% (v/v) DMSO and 0.1% Tween 20. As described previously (Dib-Hajj *et al.*, (1996) FEBS Lett. 384, 78-82), amplification was
25 carried out in two stages using a programmable thermal cycler (PTC-200, MJ Research). First, a denaturation step is performed at 94°C for four minutes, followed by an annealing step at 60°C for two minutes, and then an elongation step at 72°C for ninety seconds. Second, a denaturation step is performed at 94°C for one minute, followed by an annealing step at 60°C

for one minute, and then an elongation step at 72°C for ninety seconds. The second stage was repeated 33 times for a total of 25-35 cycles, with the elongation step in the last cycle extended to ten minutes.

Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as an internal
5 control to ensure that a lack of *NaN* signals in different tissues was not due to degraded templates or presence of PCR inhibitors. Rat GAPDH sequences were co-amplified using primers which amplify a 66 bp product that corresponds to nucleotides 328-994 (accession number: M17701). The amplified product spans multiple exon/intron splice sites, based on the structure of the human gene (Benham *et al.*, (1987) Nature 328, 275-278). DnaseI
10 treatment was routinely performed prior to reverse transcription to prevent amplification of GAPDH sequences from genomic processed pseudogenes that may have contaminated the total RNA preparation (Ercolani *et al.*, (1988) J. Biol. Chem. 263, 15335-15341).

NaN is primarily and preferentially expressed in DRG and trigeminal ganglia neurons. Figure 4 shows the results of screening by RT-PCR for the expression of *NaN* in
15 various neuronal and non-neuronal tissues. Lanes 1, 2, 4, 9 and 16 show a single amplification product co-migrating with the 400 bp marker, consistent with *NaN*-specific product of 392 bp. Lanes 1, 2, 4, 9 and 16 contain products using DRG, cerebral hemisphere, retina, and trigeminal ganglia, respectively. Using this assay, *NaN* was not detected in cerebellum, optic nerve, spinal cord, sciatic nerve, superior cervical ganglia, skeletal muscle,
20 cardiac muscle, adrenal gland, uterus, liver or kidney (lanes 3, 5-8, and 10-15, respectively). The attenuated *NaN* signal in cerebral hemisphere and retina, and the absence of this signal in the remaining tissues is not due to degraded RNA or the presence of PCR inhibitors in the cDNA templates as comparable GAPDH amplification products were obtained in a parallel set of PCR reaction (data not shown).

25 **Example 14: Transformation of a Host Cell with the *NaN* Coding Sequence**

Transformed host cells for the measurement of Na⁺ current or intracellular Na⁺ levels are usually prepared by co-transfecting constructs into cells such as HEK293 cells with a fluorescent reporter plasmid (pGreen Lantern-1, Life Technologies, Inc.) using the

calcium-phosphate precipitation technique (Ukomadu *et al.*, (1992) Neuron 8, 663-676).

HEK293 cells are typically grown in high glucose DMEM (Life Technologies) supplemented with 10% fetal calf serum (Life Technologies). After 48 hours, cells with green fluorescence are selected for recording (Dib-Hajj *et al.*, (1997) FEBS Lett. 416, 11-14).

- 5 For preparation of cell lines continuously expressing recombinant channels, the *NaN* construct is cloned into other vectors that carry a selectable marker in mammalian cells. Transfections are carried out using the calcium phosphate precipitation technique (Ukomadu *et al.*, (1992) Neuron 8, 663-676). Human embryonic kidney (HEK-293), chinese hamster ovary (CHO) cells, or other suitable cell lines are grown under standard tissue culture
- 10 conditions in Dulbeccos's modified Eagle's medium supplemented with 10% fetal bovine serum. The calcium phosphate-DNA mixture is added to the cell culture medium and left for fifteen to twenty hours, after which time the cells are washed with fresh medium. After forty-eight hours, antibiotic (G418) is added to select for cells which have acquired neomycin resistance. After two weeks in G418, 10-20 isolated cell colonies are harvested using sterile
- 15 10ml pipette tips. Colonies are grown for another four to seven days, split and subsequently tested for channel expression using whole-cell patch-clamp recording techniques and RT-PCR.

Example 15: Production of NaN specific Antibodies

- Antibodies specific to the rat, mouse or human NaN are produced with an
- 20 immunogenic NaN-specific peptide by raising polyclonal antibodies in rabbits. In one example, the peptide **CGPNPASNKDCCFEKEKDSED** (rat amino acids 285-304) (SEQ ID NO: 40) was selected because it fits the criteria for immunogenecity and surface accessibility. This peptide sequence does not match any peptide in the public databases. The underlined cysteine (C) residue was changed to Alanine (A) to prevent disulfide bond formation. This
- 25 amino acid change is not expected to significantly affect the specificity of the antibodies.

Peptide synthesis, rabbit immunization, and affinity purification of the antipeptide antibodies were performed using standard procedures. Purified antibodies were then tested on

DRG neurons in culture. Immunostaining procedures using these antibodies before and after blocking with excess peptide were performed according to standard procedures.

DRG neurons after sixteen to twenty-four hours in culture were processed for immunocytochemical detection of NaN protein as follows. Coverslips were washed with
5 complete saline solution (137 mM NaCl, 5.3 mM KCl, 1 ITIM M902 25 mM sorbitol, 10 mM HEPES, 3 mM CaCl_2 (pH 7.2)), fixed with 4% paraformaldehyde in 0.14 M phosphate buffer for ten minutes at 4°C, washed with three five minutes with phosphate-buffered saline (PBS), and blocked with PBS containing 20% normal goat serum, 1% bovine serum albumin and 0.1
10 % Triton X-100 for fifteen minutes. The coverslips were incubated in anti-NaN antibody (1:100 dilution) at 4°C overnight. Following overnight incubation, coverslips were washed extensively in PBS and then incubated with goat anti-rabbit IgG-conjugated to Cy3 (1:3000; Amersham) for two hours at room temperature. The coverslips were rinsed with PBS and mounted onto glass slides with Aqua-poly-mount. The neurons were examined with a Leitz Aristoplan light microscope equipped with epifluorescence and images were captured with a
15 Dage DC330T color camera and Scion CG-7 color PCI frame grabber (see Figure 7).

Example 16: NaN expression is altered in a neuropathic pain model

The CCI model of neuropathic pain was used to study the plasticity of sodium channel expression in DRG neurons. Twenty two adult, female Sprague-Dawley rats, weighing 240-260 g were anesthetized with pentobarbital sodium (50 mg/kg ip) and the right sciatic nerve
20 exposed at the mid-thigh. Four chromic gut (4-0) ligatures were tied loosely around the nerve as described by Bennett & Xie, (1988) Pain 33, 87-107. The incision site was closed in layers and a bacteriostatic agent administered intramuscularly.

Previous studies have shown that transection of the sciatic nerve induces dramatic changes in sodium currents of axotomized DRG neurons, which is paralleled by significant
25 changes to transcripts of various sodium channels expressed in these neurons. Sodium currents that are TTX-R and the transcripts of two TTX-R sodium channels (SNS/PN3 and NaN) are significantly attenuated while a rapidly repriming silent TTX-S current emerges and the transcript of α -III sodium channel, which produces a TTX-S current, is up-regulated. We

have discovered that CCI-induced changes in DRG neurons, fourteen days post-surgery, mirror those of axotomy. Transcripts of NaN and SNS, the two sensory neuron-specific TTX-R channels, are significantly down-regulated as is the TTX-R sodium current, while transcripts of the TTX-S α -III sodium channel are up-regulated, in small diameter DRG neurons. These changes may be partly responsible for making DRG neurons hyperexcitable, that contributes to the hyperalgesia that results from this injury.

Example 17: Assays for agents which modulate the activity of the NaN channel using patch clamp methods

Cells lines expressing the cloned Na⁺ channel are used to assay for agents which modulate the activity of the NaN channel, *e.g.*, agents which block or inhibit the channel or enhance channel opening. Since the channel activation is voltage dependent, depolarizing conditions may be used for observation of baseline activity that is modified by the agent to be tested. Depolarization may be achieved by any means available, for example, by raising the extracellular potassium ion concentration to about 20 to 40 nM, or by repeated electrical pulses.

The agent to be tested is incubated with HEK 293 or other transformed cells that express the Na⁺ channel (Dib-Hajj *et al.*, (1997) FEBS Lett. 416, 11-14). After incubation for a sufficient period of time, the agent induced changes in Na⁺ channel activity can be measured by patch-clamp methods (Hamill *et al.*, (1981) Pflügers Arch. 391, 85-100). Data for these measurements are acquired on a MacIntosh Quadra 950, or similar computer, using a program such as Pulse (v 7.52, HEKA, German). Fire-polished electrodes (0.8-1.5 MW) are fabricated from capillary glass using a Sutter P-87 puller or a similar instrument. Cells are usually only considered for analysis if initial seal resistance is <5 Gohm, they have high leakage currents (holding current <0.1 nA at -80 mV), membrane blebs, and an access resistance <5 Mohm. Access resistance is monitored and data is not used if resistance changes occur. Voltage errors are minimized using series resistance compensation and the capacitance artifact will be canceled as necessary using computer-controlled amplifier circuitry or other similar methods.

For comparisons of the voltage dependence of activation and inactivation, cells with a maximum voltage error of <10 mV after compensation are usually used. Linear leak subtraction is used for voltage clamp recordings. Membrane currents are typically filtered at 5 KHz and sampled at 20 KHz. The pipette solution contains a standard solution such as: 140 mM CsF, 2 mM MgCl₂, 1 mM EGTA, and 10 mM Na-HEPES (pH 7.3). The standard bathing solution is a standard solution such as 140 mM NaCl, 3 mM KCl, 2 mM MgCl₂, 1 mM CaCl₂, 10 mM HEPES, and 10 mM glucose (pH 7.3).

Tetrodotoxin (TTX)-resistant and TTX-sensitive Na⁺ currents are measured by exposure to appropriate concentrations of TTX and/or by pre-pulse protocols which distinguish between TTX-sensitive and TTX-resistant currents on the basis of their distinct steady-state inactivation properties (Cummins & Waxman (1997) J. Neurophysiol. 17, 3503-3514; Sontheimer & Waxman, (1992) J. Neurophysiol. 68, 1001-1011).

Data are collected using standard pulse protocols and are analyzed to measure sodium current properties that include voltage-dependence, steady-state characteristics, kinetics, and re-priming. Measurements of current amplitude and cell capacitance provides an estimate of Na⁺ current density, thereby permitting comparisons of channel density under different conditions (Cummins & Waxman (1997) J. Neurophysiol. 17, 3503-3514,30). Cells are studied in the current clamp mode to study patterns of spontaneous and evoked action potential generation, threshold for firing, frequency response characteristics, and response to de- and hyperpolarization, and other aspects of electrogenesis (Sontheimer & Waxman, (1992) J. Neurophysiol. 68, 1001-1011). These measurements are carried out both in control cells expressing *NaV* and in cells expressing *NaV* that also have been exposed to the agent to be tested.

Example 18: Assays for agents which modulate the activity of the NaN channel by the measurement of Intracellular Sodium [Na⁺]

The agent to be tested is incubated with cells exhibiting NaN channel activity. After incubation for a sufficient period of time, the agent induced changes in Na⁺ channel are measured by ratiometric imaging of [Na⁺]_i using SBFI. In this method, cytosolic-free Na⁺ is

measured using an indicator for Na^+ , such as SBFI (sodium-binding benzofuran isophthalate (Harootunian *et al.*, (1989) J. Biol. Chem. 264, 19458-19467)); or a similar dye. Cells are first loaded with the membrane-permeable acetoxymethyl ester form of SBFI (SBFI/AM) or a similar dye (usually dissolved in dimethyl sulfoxide (DMSO) at a stock concentration of 10 mM). Recordings are obtained on the stage of a microscope using a commercially available ratiometric imaging setup (*e.g.*, from Georgia Instruments). Excitation light is provided at appropriate wavelengths (*e.g.*, 340:385 nm). Excitation light is passed to the cells through a dichroic reflector (400 nm) and emitted light above 450 nm was collected. Fluorescence signals are amplified, *e.g.*, by an image intensifier (GenIISyS) and collected with a CCD camera, or similar device, interfaced to a frame grabber. To account for fluorescence rundown, the fluorescence ratio 340:385 is used to assay cytosolic-free Na^+ .

For calibration of SBFI's fluorescence, cells are perfused with calibration solutions containing known Na^+ concentrations (typically 0 and 30 mM, or 0, 30, and 50 mM $[\text{Na}^+]$, and gramicidin and monensin. As reported by Rose and Ransom (Rose & Ransom, (1996) J. Physiol. (Lond) 491, 291-305), the 345/390 nm fluorescence ratio of intracellular SBFI changes monotonically with changes in $[\text{Na}^+]$. Experiments are repeated on multiple (typically at least four) different coverslips, providing statistically significant measurements of intracellular sodium in control cells, and in cells exposed to various concentrations of agents that may block, inhibit or enhance the activity of the channel. Use of this method is illustrated in Sontheimer *et al.*, (1994) J. Neurosci. 14, 2464-2475.

Example 19: Assays for agents which modulate the activity of the Na^+ channel by scintigraphic imaging

Cells lines expressing the cloned Na^+ channel are used to assay for agents which modulate the activity of the Na^+ channel, *e.g.*, agents which block the channel or enhance channel opening. For example, the agent to be tested is incubated with HEK 293 or other transformed cells that express the Na^+ channel (Dib-Hajj *et al.*, (1997) FEBS Lett. 416, 11-14). After incubation for a sufficient period of time, the agent induced changes in Na^+ channel activity are detected by the measurement of Na^+ influx by isotopic methods. ^{22}Na is a

gamma emitter and can be used to measure Na^+ flux (Kimelberg & Waltz, (1988) The Neuronal Microenvironment (Boulton *et al.*, editors) Humana Press) and $^{86}\text{Rb}^+$ can be used to measure Na^+/K^+ ATPase activity which provides a measure of Na channel activity (Sontheimer *et al.*, (1994) J. Neurosci. 14, 2464-2475) $^{86}\text{Rb}^+$ ions are taken up by the Na^+/K^+ ATPase like
5 K^+ ions, but have the advantage of a much longer half-life than $^{42}\text{K}^+$ (Kimelberg & Mayhew (1975) J. Biol. Chem. 250, 100-104). Thus, measurement of the unidirectional ouabain-sensitive $^{86}\text{Rb}^+$ uptake provides a quantitative method for assaying Na^+/K^+ -ATPase activity which follows action potentials.

Following incubation of cell expressing *NaN* to the isotope, the cellular content of the
10 isotope is measured by liquid scintillation counting or a similar method, and cell protein is determined using a method such as the bicinchoninic acid protein assay (Smith *et al.*, (1985) Anal. Biochem. 150, 76-85) following the modifications (Goldschmidt & Kimelberg (1989) Anal. Biochem. 177, 41-45) for cultured cells. ^{22}Na and $^{86}\text{Rb}^+$ fluxes are determined in the presence and absence of agents that may block, inhibit, or enhance Na^+ . This permits
15 determination of the actions of these agents on *NaN*.

Example 20: In situ hybridization

a. Probes

Probes are prepared as described above in Example 5.

b. DRG Neuron Culture

20 Cultures of DRG neurons from adult rats were established as described previously (Rizzo *et al.*, (1994) J. Neurophysiol. 72, 2796-2815). Briefly, lumbar ganglia (L4, L5) from adult Sprague Dawley female rats were freed from their connective sheaths and incubated sequentially in enzyme solutions containing collagenase and then papain. The tissue was triturated in culture medium containing 1:1 Dulbecco's modified Eagle's medium (DMEM)
25 and Hank's F12 medium and 10% fetal calf serum, 1.5 mg/ml trypsin inhibitor, 1.5 mg/ml bovine serum albumin, 100 units/ml penicillin and 0.1 mg/ml streptomycin and plated at a density of 500-1000 cells/mm² on polyornithine/laminin coated coverslips. The cells were

maintained at 37°C in a humidified 95% air/5% CO₂ incubator overnight and then processed for *in situ* hybridization cytochemistry as described previously (Black *et al.*, (1994) Brain Res. Mol. Brain Res. 23, 235-245; Zur *et al.*, (1995) Brain Res. Mol. Brain Res. 30, 97-105). Trigeminal ganglia can be cultured by a skilled artisan using similar methods.

5 c. Tissue Preparation

Adult female Sprague Dawley rats were deeply anesthetized, *e.g.*, with chloral hydrate and perfused through the heart, first with a phosphate-buffered saline (PBS) solution and then with a 4% paraformaldehyde in 0.14 M Sorensen's phosphate buffer (pH 7.4) at 4°C. Following perfusion fixation, dorsal root ganglia at levels L4 and L5 and trigeminal ganglia
10 were collected and placed in fresh fixative at 4°C. After two to four hours, the tissue was transferred to a solution containing 4% paraformaldehyde and 30% sucrose in 0.14 M phosphate buffer and stored overnight at 4°C. Fifteen µm sections were cut and placed on poly-L-lysine-coated slides. The slides were processed for *in situ* hybridization cytochemistry as previously described (Waxman *et al.*, (1994) J. Neurophysiol. 72, 466-470; Black *et al.*,
15 (1994) Brain Res. Mol. Brain Res. 23, 235-245). Following *in situ* hybridization cytochemistry, the slides were dehydrated, cleared and mounted with Permount. The results are shown in Figure 5.

Sections of DRG hybridized with *NaV* sense riboprobe showed no specific labeling (panel C, Figure 5). In DRG (panel A, Figure 5) and trigeminal (panel B) sections hybridized
20 with a *NaV* antisense riboprobe, with the *NaV* signal present in most small (<30 µm diam.) neurons; in contrast, most large (>30 µm diam.) neurons did not exhibit *NaV* hybridization signal. Sections of spinal cord, cerebellum and liver hybridized with an antisense *NaV* riboprobe showed no specific signal (panels D, E and F respectively).

Example 21: Microsatellite Sequences

25 The following are the murine intronic microsatellite sequences. These microsatellites may be polymorphic in the human SCN11a gene and could be used as markers to screen for

It should be understood that the foregoing discussion and examples merely present a detailed description of certain preferred embodiments. It will be apparent to those of ordinary skill in the art that various modification and equivalents can be made without departing from the spirit and scope of the invention. The documents cited and referred to in this patent
5 specification are hereby incorporated by reference in their entirety.

CLAIMS

1. An isolated nucleic acid molecule selected from the group consisting of a nucleic acid molecule comprising SEQ ID NO: 41, a nucleic acid molecule which encodes the amino acid sequence of SEQ ID NO: 42, a nucleic acid molecule which encodes an allelic variant of
5 SEQ ID NO:42, a nucleic acid molecule which encodes a human protein exhibiting at least about 76% amino acid sequence identity to SEQ ID NO:42 and a nucleic acid molecule that hybridizes to one of the foregoing sequences under stringent conditions.
2. The isolated nucleic acid of claim 1, wherein the nucleic acid encodes a voltage gated Na⁺ channel that is preferentially expressed in dorsal root ganglia or trigeminal ganglia.
3. The isolated nucleic acid of claim 2, wherein the nucleic acid encodes the human NaN sodium channel.
4. An expression vector comprising the isolated nucleic acid of any one of claims 1 to 3, alone or together with appropriate regulatory and expression control elements.
5. A host cell transformed with the expression vector of claim 4.
6. A Na⁺ channel encoded by an isolated nucleic acid molecule of any one of claims 1 to 3.
7. The Na⁺ channel of claim 6, comprising the amino acid sequence of SEQ ID NO:42.
8. An isolated protein consisting of the amino acid sequence of SEQ ID NO: 42 or a peptide fragment thereof.

9. A protein comprised within a membrane fragment isolated from the host cell of claim 5.

10. A method to identify an agent that modulates the activity of the Na^+ channel of claim 6, comprising the steps of bringing the agent into contact with a cell that expresses the Na^+ channel on its surface and measuring any resultant changes in the sodium current, resultant change in membrane potential or change in intracellular Na^+ .

11. The method of claim 10, wherein the measuring step is accomplished by voltage clamp measurements or measurement of membrane potential.

12. The method of claim 10, wherein the measuring step is accomplished by measuring the level of intracellular sodium.

13. The method of claim 10, wherein the measuring step is accomplished by measuring sodium influx.

14. The method of claim 13, wherein the sodium influx is measured using ^{22}Na or ^{86}Rb .

15. A method to identify an agent that modulates the transcription or translation of mRNA encoding the Na^+ channel of claim 6, comprising the steps of bringing the agent into contact with a cell that expresses the Na^+ channel and measuring the resultant level of expression of the Na^+ channel.

16. A method to treat pain, paraesthesia and/or hyperexcitability phenomena in an animal or human subject by administering an effective amount of an agent that alters Na^+ current flow through Na^+ channels in DRG or trigeminal neurons.

17. A method to treat pain, paraesthesia and/or hyperexcitability phenomena in an animal or human subject by administering an effective amount of an agent that modulates the transcription or translation of mRNA encoding the Na⁺ channel of claim 6.

18. An isolated nucleic acid that is antisense to the nucleic acid of claim 1 and of sufficient length to modulate the expression of NaN channel in a cell containing the mRNA.

19. A scintigraphic method to image the loci of pain generation or provide a measure of the level of pain associated with DRG or trigeminal neuron mediated hyperexcitability in an animal or human subject by administering labeled monoclonal antibodies or other labeled ligands specific for the human NaN Na⁺ channel.

20. A method to identify tissues, cells or cell types that express the human NaN sodium channel, comprising the step of detecting human NaN on the cell surface or intracellularly.

21. A method to identify tissues, cells or cell types that express human NaN comprising the step of detecting the presence therein of human NaN encoding mRNA.

22. A method of producing a transformed cell that expresses an exogenous NaN encoding nucleic acid, comprising the step of transforming the cell with an expression vector comprising the isolated nucleic acid of any of claims 1 to 3, together with appropriate regulatory or expression control elements.

23. An isolated antibody specific for the human NaN channel or polypeptide fragment thereof.

24. The isolated antibody of claim 23, wherein the antibody is labeled.

25. A method of producing recombinant NaN protein, comprising the step of culturing the transformed host of claim 5 under conditions in which the NaN sodium channel or protein is expressed.

26. A therapeutic composition comprising an effective amount of an agent capable of altering, such as by increasing or decreasing, the rapidly repriming current flow in axotomized, inflamed or otherwise injured DRG neurons.

27. A method to treat acute pain or acute or chronic neuropathic or inflammatory pain and hyperexcitability phenomena in an animal or a human patient by administering the therapeutic composition of claim 26.

28. A method to screen candidate compounds for use in treating pain and hyperexcitability phenomena by testing their ability to upregulate or downregulate the NaN channel mRNA in axotomized, inflamed or otherwise injured DRG neurons.

29. A chimeric NaN channel.

30. A chimeric channel of claim 29, wherein at least one human domain has been substituted with the corresponding domain from the NaN channel of another species.

31. A chimeric channel of claim 30, wherein the species is rat or mouse.

32. A nucleic acid molecule encoding a chimeric NaN channel of any one of claims 29 through 31.

33. An NaN channel protein comprising a positively charged amino acid at a position corresponding to residue 670 of SEQ ID NO:42.

34. An NaN channel protein of claim 33, wherein the positively charged amino acid is arginine.

35. An isolated nucleic acid molecule encoding a channel protein of either of claims 33 or 34.

36. A therapeutic composition of claim 26, further comprising at least one second agent that modulates a channel in primary sensory neurons.

37. A therapeutic composition of claim 36, wherein the composition comprises agents which modulate NaN and at least one channel selected from the group consisting of PN1/hNE and SNS/PN3.

SEQUENCE LISTING

<110> Yale University
 Dib-Hajj, Sulayman
 Waxman, Stephen G.

<120> Modulation of Sodium Channels in Dorsal Root Ganglia

<130> 44574-5004-02-WO

<140>

<141>

<150> US 09/354,147

<151> 1999-07-16

<160> 44

<170> PatentIn Ver. 2.1

<210> 1

<211> 5875

<212> DNA

<213> Rattus norvegicus

<220>

<221> CDS

<222> (41)..(5335)

<223> cDNA sequence for rat NaN

<220>

<221> unsure

<222> (1996)..(4042)

<223> n = a or c or g or t

<400> 1

```

acggtgccct gatcctctgt accaggaaga caggggaag atg gag gag agg tac    55
                                     Met Glu Glu Arg Tyr
                                     1          5

tac ccg gtg atc ttc ccg gac gag cgg aat ttc cgc ccc ttc act tcc    103
Tyr Pro Val Ile Phe Pro Asp Glu Arg Asn Phe Arg Pro Phe Thr Ser
                10          15          20

gac tct ctg gct gcc ata gag aag cgg att gct atc caa aag gag agg    151
Asp Ser Leu Ala Ala Ile Glu Lys Arg Ile Ala Ile Gln Lys Glu Arg
                25          30          35

aag aag tcc aaa gac aag gcg gca gct gag ccc cag cct cgg cct cag    199
Lys Lys Ser Lys Asp Lys Ala Ala Ala Glu Pro Gln Pro Arg Pro Gln
                40          45          50

ctt gac cta aag gcc tcc agg aag tta cct aag ctt tat ggt gac att    247
Leu Asp Leu Lys Ala Ser Arg Lys Leu Pro Lys Leu Tyr Gly Asp Ile
                55          60          65

```

ccc cct gag ctt gta gcg aag cct ctg gaa gac ctg gac cca ttc tac	295
Pro Pro Glu Leu Val Ala Lys Pro Leu Glu Asp Leu Asp Pro Phe Tyr	
70 75 80 85	
aaa gac cat aag aca ttc atg gtg ttg aac aag aag aga aca att tat	343
Lys Asp His Lys Thr Phe Met Val Leu Asn Lys Lys Arg Thr Ile Tyr	
90 95 100	
cgc ttc agc gcc aag cgg gcc ttg ttc att ctg ggg cct ttt aat ccc	391
Arg Phe Ser Ala Lys Arg Ala Leu Phe Ile Leu Gly Pro Phe Asn Pro	
105 110 115	
ctc aga agc tta atg att cgt atc tct gtc cat tca gtc ttt agc atg	439
Leu Arg Ser Leu Met Ile Arg Ile Ser Val His Ser Val Phe Ser Met	
120 125 130	
ttc atc atc tgc acg gtg atc atc aac tgt atg ttc atg gcg aat tct	487
Phe Ile Ile Cys Thr Val Ile Ile Asn Cys Met Phe Met Ala Asn Ser	
135 140 145	
atg gag aga agt ttc gac aac gac att ccc gaa tac gtc ttc att ggg	535
Met Glu Arg Ser Phe Asp Asn Asp Ile Pro Glu Tyr Val Phe Ile Gly	
150 155 160 165	
att tat att tta gaa gct gtg att aaa ata ttg gca aga ggc ttc att	583
Ile Tyr Ile Leu Glu Ala Val Ile Lys Ile Leu Ala Arg Gly Phe Ile	
170 175 180	
gtg gat gag ttt tcc ttc ctc cga gat ccg tgg aac tgg ctg gac ttc	631
Val Asp Glu Phe Ser Phe Leu Arg Asp Pro Trp Asn Trp Leu Asp Phe	
185 190 195	
att gtc att gga aca gcg atc gca act tgt ttt ccg ggc agc caa gtc	679
Ile Val Ile Gly Thr Ala Ile Ala Thr Cys Phe Pro Gly Ser Gln Val	
200 205 210	
aat ctt tca gct ctt cgt acc ttc cga gtg ttc aga gct ctg aag gcg	727
Asn Leu Ser Ala Leu Arg Thr Phe Arg Val Phe Arg Ala Leu Lys Ala	
215 220 225	
att tca gtt atc tca ggt ctg aag gtc atc gta ggt gcc ctg ctg cgc	775
Ile Ser Val Ile Ser Gly Leu Lys Val Ile Val Gly Ala Leu Leu Arg	
230 235 240 245	
tcg gtg aag aag ctg gta gac gtg atg gtc ctc act ctc ttc tgc ctc	823
Ser Val Lys Lys Leu Val Asp Val Met Val Leu Thr Leu Phe Cys Leu	
250 255 260	
agc atc ttt gcc ctg gtc ggt cag cag ctg ttc atg gga att ctg aac	871
Ser Ile Phe Ala Leu Val Gly Gln Gln Leu Phe Met Gly Ile Leu Asn	
265 270 275	
cag aag tgt att aag cac aac tgt ggc ccc aac cct gca tcc aac aag	919
Gln Lys Cys Ile Lys His Asn Cys Gly Pro Asn Pro Ala Ser Asn Lys	
280 285 290	

gat tgt ttt gaa aag gaa aaa gat agc gaa gac ttc ata atg tgt ggt Asp Cys Phe Glu Lys Glu Lys Asp Ser Glu Asp Phe Ile Met Cys Gly 295 300 305	967
acc tgg ctc ggc agc aga ccc tgt ccc aat ggt tct acg tgc gat aaa Thr Trp Leu Gly Ser Arg Pro Cys Pro Asn Gly Ser Thr Cys Asp Lys 310 315 320 325	1015
acc aca ttg aac cca gac aat aat tat aca aag ttt gac aac ttt ggc Thr Thr Leu Asn Pro Asp Asn Asn Tyr Thr Lys Phe Asp Asn Phe Gly 330 335 340	1063
tgg tcc ttt ctc gcc atg ttc cgg gtt atg act caa gac tcc tgg gag Trp Ser Phe Leu Ala Met Phe Arg Val Met Thr Gln Asp Ser Trp Glu 345 350 355	1111
agg ctt tac cga cag atc ctg cgg acc tct ggg atc tac ttt gtc ttc Arg Leu Tyr Arg Gln Ile Leu Arg Thr Ser Gly Ile Tyr Phe Val Phe 360 365 370	1159
ttc ttc gtg gtg gtc atc ttc ctg ggc tcc ttc tac ctg ctt aac cta Phe Phe Val Val Val Ile Phe Leu Gly Ser Phe Tyr Leu Leu Asn Leu 375 380 385	1207
acc ctg gct gtt gtc acc atg gct tat gaa gaa cag aac aga aat gta Thr Leu Ala Val Val Thr Met Ala Tyr Glu Glu Gln Asn Arg Asn Val 390 395 400 405	1255
gct gct gag aca gag gcc aag gag aaa atg ttt cag gaa gcc cag cag Ala Ala Glu Thr Glu Ala Lys Glu Lys Met Phe Gln Glu Ala Gln Gln 410 415 420	1303
ctg tta agg gag gag aag gag gct ctg gtt gcc atg gga att gac aga Leu Leu Arg Glu Glu Lys Glu Ala Leu Val Ala Met Gly Ile Asp Arg 425 430 435	1351
agt tcc ctt aat tcc ctt caa gct tca tcc ttt tcc ccg aag aag agg Ser Ser Leu Asn Ser Leu Gln Ala Ser Ser Phe Ser Pro Lys Lys Arg 440 445 450	1399
aag ttt ttc ggt agt aag aca aga aag tcc ttc ttt atg aga ggg tcc Lys Phe Phe Gly Ser Lys Thr Arg Lys Ser Phe Phe Met Arg Gly Ser 455 460 465	1447
aag acg gcc caa gcc tca gcg tct gat tca gag gac gat gcc tct aaa Lys Thr Ala Gln Ala Ser Ala Ser Asp Ser Glu Asp Asp Ala Ser Lys 470 475 480 485	1495
aat cca cag ctc ctt gag cag acc aaa cga ctg tcc cag aac ttg cca Asn Pro Gln Leu Leu Glu Gln Thr Lys Arg Leu Ser Gln Asn Leu Pro 490 495 500	1543
gtg gat ctc ttt gat gag cac gtg gac ccc ctc cac agg cag aga gcg Val Asp Leu Phe Asp Glu His Val Asp Pro Leu His Arg Gln Arg Ala 505 510 515	1591

ctg agc gct gtc agt atc tta acc atc acc atg cag gaa caa gaa aaa	1639
Leu Ser Ala Val Ser Ile Leu Thr Ile Thr Met Gln Glu Gln Glu Lys	
520 525 530	
ttc cag gag cct tgt ttc cca tgt ggg aaa aat ttg gcc tct aag tac	1687
Phe Gln Glu Pro Cys Phe Pro Cys Gly Lys Asn Leu Ala Ser Lys Tyr	
535 540 545	
ctg gtg tgg gac tgt agc cct caa tgg ctg tgc ata aag aag gtc ctg	1735
Leu Val Trp Asp Cys Ser Pro Gln Trp Leu Cys Ile Lys Lys Val Leu	
550 555 560 565	
cgg acc atc atg acg gat ccc ttt act gag ctg gcc atc acc atc tgc	1783
Arg Thr Ile Met Thr Asp Pro Phe Thr Glu Leu Ala Ile Thr Ile Cys	
570 575 580	
atc atc atc aat acc gtt ttc tta gcc gtg gag cac cac aac atg gat	1831
Ile Ile Ile Asn Thr Val Phe Leu Ala Val Glu His His Asn Met Asp	
585 590 595	
gac aac tta aag acc ata ctg aaa ata gga aac tgg gtt ttc acg gga	1879
Asp Asn Leu Lys Thr Ile Leu Lys Ile Gly Asn Trp Val Phe Thr Gly	
600 605 610	
att ttc ata gcg gaa atg tgt ctc aag atc atc gcg ctc gac cct tac	1927
Ile Phe Ile Ala Glu Met Cys Leu Lys Ile Ile Ala Leu Asp Pro Tyr	
615 620 625	
cac tac ttc cgg cac ggc tgg aat gtt ttt gac agc atc gtg gcc ctc	1975
His Tyr Phe Arg His Gly Trp Asn Val Phe Asp Ser Ile Val Ala Leu	
630 635 640 645	
ctg agt ctc gct gat gtg ctn tac aac aca ctg tct gat aac aat agg	2023
Leu Ser Leu Ala Asp Val Xaa Tyr Asn Thr Leu Ser Asp Asn Asn Arg	
650 655 660	
tct ttc ttg gct tcc ctc aga gtg ctg agg gtc ttc aag tta gcc aaa	2071
Ser Phe Leu Ala Ser Leu Arg Val Leu Arg Val Phe Lys Leu Ala Lys	
665 670 675	
tcc tgg ccc acg tta aac act ctc att aag atc atc ggc cac tcc gtg	2119
Ser Trp Pro Thr Leu Asn Thr Leu Ile Lys Ile Ile Gly His Ser Val	
680 685 690	
ggc gcg ctt gga aac ctg act gtg gtc ctg act atc gtg gtc ttc atc	2167
Gly Ala Leu Gly Asn Leu Thr Val Val Leu Thr Ile Val Val Phe Ile	
695 700 705	
ttt tct gtg gtg ggc atg cgg ctc ttc ggc acc aag ttt aac aag acc	2215
Phe Ser Val Val Gly Met Arg Leu Phe Gly Thr Lys Phe Asn Lys Thr	
710 715 720 725	
gcc tac gcc acc cag gag cgg ccc agg cgg cgc tgg cac atg gat aat	2263
Ala Tyr Ala Thr Gln Glu Arg Pro Arg Arg Arg Trp His Met Asp Asn	
730 735 740	

ttc tac cac tcc ttc ctg gtg gtg ttc cgc atc ctc tgt ggg gaa tgg	2311
Phe Tyr His Ser Phe Leu Val Val Phe Arg Ile Leu Cys Gly Glu Trp	
745 750 755	
atc gag aac atg tgg ggc tgc atg cag gat atg gac ggc tcc ccg ttg	2359
Ile Glu Asn Met Trp Gly Cys Met Gln Asp Met Asp Gly Ser Pro Leu	
760 765 770	
tgc atc att gtc ttt gtc ctg ata atg gtg atc ggg aag ctt gtg gtg	2407
Cys Ile Ile Val Phe Val Leu Ile Met Val Ile Gly Lys Leu Val Val	
775 780 785	
ctt aac ctc ttc att gcc ttg ctg ctc aat tcc ttc agc aat gag gag	2455
Leu Asn Leu Phe Ile Ala Leu Leu Leu Asn Ser Phe Ser Asn Glu Glu	
790 795 800 805	
aag gat ggg agc ctg gaa gga gag acc agg aaa acc aaa gtg cag cta	2503
Lys Asp Gly Ser Leu Glu Gly Glu Thr Arg Lys Thr Lys Val Gln Leu	
810 815 820	
gcc ctg gat cgg ttc cgc cgg gcc ttc tcc ttc atg ctg cac gct ctt	2551
Ala Leu Asp Arg Phe Arg Arg Ala Phe Ser Phe Met Leu His Ala Leu	
825 830 835	
cag agt ttt tgt tgc aag aaa tgc agg agg aaa aac tcg cca aag cca	2599
Gln Ser Phe Cys Cys Lys Lys Cys Arg Arg Lys Asn Ser Pro Lys Pro	
840 845 850	
aaa gag aca aca gaa agc ttt gct ggt gag aat aaa gac tca atc ctc	2647
Lys Glu Thr Thr Glu Ser Phe Ala Gly Glu Asn Lys Asp Ser Ile Leu	
855 860 865	
ccg gat gcg agg ccc tgg aag gag tat gat aca gac atg gct ttg tac	2695
Pro Asp Ala Arg Pro Trp Lys Glu Tyr Asp Thr Asp Met Ala Leu Tyr	
870 875 880 885	
act gga cag gcc ggg gct ccg ctg gcc cca ctc gca gag gta gag gac	2743
Thr Gly Gln Ala Gly Ala Pro Leu Ala Pro Leu Ala Glu Val Glu Asp	
890 895 900	
gat gtg gaa tat tgt ggt gaa ggc ggt gcc cta ccc acc tca caa cat	2791
Asp Val Glu Tyr Cys Gly Glu Gly Glu Ala Leu Pro Thr Ser Gln His	
905 910 915	
agt gct gga gtt cag gcc ggt gac ctc cct cca gag acc aag cag ctc	2839
Ser Ala Gly Val Gln Ala Gly Asp Leu Pro Pro Glu Thr Lys Gln Leu	
920 925 930	
act agc ccg gat gac caa ggg gtt gaa atg gaa gta ttt tct gaa gaa	2887
Thr Ser Pro Asp Asp Gln Gly Val Glu Met Glu Val Phe Ser Glu Glu	
935 940 945	
gat ctg cat tta agc ata cag agt cct cga aag aag tct gac gca gtg	2935
Asp Leu His Leu Ser Ile Gln Ser Pro Arg Lys Lys Ser Asp Ala Val	
950 955 960 965	

agc atg ctc tcg gaa tgc agc aca att gac ctg aat gat atc ttt aga	2983
Ser Met Leu Ser Glu Cys Ser Thr Ile Asp Leu Asn Asp Ile Phe Arg	
970 975 980	
aat tta cag aaa aca gtt tcc ccc aaa aag cag cca gat aga tgc ttt	3031
Asn Leu Gln Lys Thr Val Ser Pro Lys Lys Gln Pro Asp Arg Cys Phe	
985 990 995	
ccc aag ggc ctt agt tgt cac ttt cta tgc cac aaa aca gac aag aga	3079
Pro Lys Gly Leu Ser Cys His Phe Leu Cys His Lys Thr Asp Lys Arg	
1000 1005 1010	
aag tcc ccc tgg gtc ctg tgg tgg aac att cgg aaa acc tgc tac caa	3127
Lys Ser Pro Trp Val Leu Trp Trp Asn Ile Arg Lys Thr Cys Tyr Gln	
1015 1020 1025	
atc gtg aag cac agc tgg ttt gag agt ttc ata atc ttt gtt att ctg	3175
Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile Ile Phe Val Ile Leu	
1030 1035 1040 1045	
ctg agc agt gga gcg ctg ata ttt gaa gat gtc aat ctc ccc agc cgg	3223
Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val Asn Leu Pro Ser Arg	
1050 1055 1060	
ccc caa gtt gag aaa tta cta agg tgt acc gat aat att ttc aca ttt	3271
Pro Gln Val Glu Lys Leu Leu Arg Cys Thr Asp Asn Ile Phe Thr Phe	
1065 1070 1075	
att ttc ctc ctg gaa atg atc ctg aag tgg gtg gcc ttt gga ttc cgg	3319
Ile Phe Leu Leu Glu Met Ile Leu Lys Trp Val Ala Phe Gly Phe Arg	
1080 1085 1090	
agg tat ttc acc agt gcc tgg tgc tgg ctt gat ttc ctc att gtg gtg	3367
Arg Tyr Phe Thr Ser Ala Trp Cys Trp Leu Asp Phe Leu Ile Val Val	
1095 1100 1105	
gtg tct gtg ctc agt ctc atg aat cta cca agc ttg aag tcc ttc cgg	3415
Val Ser Val Leu Ser Leu Met Asn Leu Pro Ser Leu Lys Ser Phe Arg	
1110 1115 1120 1125	
act ctg cgg gcc ctg aga cct ctg cgg gcg ctg tcc cag ttt gaa gga	3463
Thr Leu Arg Ala Leu Arg Pro Leu Arg Ala Leu Ser Gln Phe Glu Gly	
1130 1135 1140	
atg aag gtt gtc gtc tac gcc ctg atc agc gcc ata cct gcc att ctc	3511
Met Lys Val Val Val Tyr Ala Leu Ile Ser Ala Ile Pro Ala Ile Leu	
1145 1150 1155	
aat gtc ttg ctg gtc tgc ctc att ttc tgg ctc gta ttt tgt atc ttg	3559
Asn Val Leu Leu Val Cys Leu Ile Phe Trp Leu Val Phe Cys Ile Leu	
1160 1165 1170	
gga gta aat tta ttt tct ggg aag ttt gga agg tgc att aac ggg aca	3607
Gly Val Asn Leu Phe Ser Gly Lys Phe Gly Arg Cys Ile Asn Gly Thr	
1175 1180 1185	

7

tgt gtg gtc gtg gtt ctt tct atc att agt acc ctg gtt tcc cgc ttg Cys Val Val Val Leu Ser Ile Ile Ser Thr Leu Val Ser Arg Leu 1415 1420 1425	4327
gag gac agt gac att tct ttc ccg ccc acg ctc ttc aga gtc gtc cgc Glu Asp Ser Asp Ile Ser Phe Pro Pro Thr Leu Phe Arg Val Val Arg 1430 1435 1440 1445	4375
ttg gct cgg att ggt cga atc ctc agg ctg gtc cgg gct gcc cgg gga Leu Ala Arg Ile Gly Arg Ile Leu Arg Leu Val Arg Ala Ala Arg Gly 1450 1455 1460	4423
atc agg acc ctc ctc ttt gct ttg atg atg tct ctc ccc tct ctc ttc Ile Arg Thr Leu Leu Phe Ala Leu Met Met Ser Leu Pro Ser Leu Phe 1465 1470 1475	4471
aac atc ggt ctg ctg ctc ttc ctg gtg atg ttc att tac gcc atc ttt Asn Ile Gly Leu Leu Leu Phe Leu Val Met Phe Ile Tyr Ala Ile Phe 1480 1485 1490	4519
ggg atg agc tgg ttt tcc aaa gtg aag aag ggc tcc ggg atc gac gac Gly Met Ser Trp Phe Ser Lys Val Lys Lys Gly Ser Gly Ile Asp Asp 1495 1500 1505	4567
atc ttc aac ttc gag acc ttt acg ggc agc atg ctg tgc ctc ttc cag Ile Phe Asn Phe Glu Thr Phe Thr Gly Ser Met Leu Cys Leu Phe Gln 1510 1515 1520 1525	4615
ata acc act tcg gct ggc tgg gat acc ctc ctc aac ccc atg ctg gag Ile Thr Thr Ser Ala Gly Trp Asp Thr Leu Leu Asn Pro Met Leu Glu 1530 1535 1540	4663
gca aaa gaa cac tgc aac tcc tcc tcc caa gac agc tgt cag cag ccg Ala Lys Glu His Cys Asn Ser Ser Ser Gln Asp Ser Cys Gln Gln Pro 1545 1550 1555	4711
cag ata gcc gtc gtc tac ttc gtc agt tac atc atc atc tcc ttc ctc Gln Ile Ala Val Val Tyr Phe Val Ser Tyr Ile Ile Ile Ser Phe Leu 1560 1565 1570	4759
atc gtg gtc aac atg tac atc gct gtg atc ctc gag aac ttc aac aca Ile Val Val Asn Met Tyr Ile Ala Val Ile Leu Glu Asn Phe Asn Thr 1575 1580 1585	4807
gcc acg gag gag agc gag gac cct ctg gga gag gac gac ttt gaa atc Ala Thr Glu Glu Ser Glu Asp Pro Leu Gly Glu Asp Asp Phe Glu Ile 1590 1595 1600 1605	4855
ttc tat gag gtc tgg gag aag ttt gac ccc gag gcg tcg cag ttc atc Phe Tyr Glu Val Trp Glu Lys Phe Asp Pro Glu Ala Ser Gln Phe Ile 1610 1615 1620	4903
cag tat tcg gcc ctc tct gac ttt gcg gac gcc ctg ccg gag ccg ttg Gln Tyr Ser Ala Leu Ser Asp Phe Ala Asp Ala Leu Pro Glu Pro Leu 1625 1630 1635	4951

cgt gtg gcc aag ccg aat aag ttt cag ttt cta gtg atg gac ttg ccc 4999
 Arg Val Ala Lys Pro Asn Lys Phe Gln Phe Leu Val Met Asp Leu Pro
 1640 1645 1650

atg gtg atg ggc gac cgc ctc cat tgc atg gat gtt ctc ttt gct ttc 5047
 Met Val Met Gly Asp Arg Leu His Cys Met Asp Val Leu Phe Ala Phe
 1655 1660 1665

act acc agg gtc ctc ggg gac tcc agc ggc ttg gat acc atg aaa acc 5095
 Thr Thr Arg Val Leu Gly Asp Ser Ser Gly Leu Asp Thr Met Lys Thr
 1670 1675 1680 1685

atg atg gag gag aag ttt atg gag gcc aac cct ttt aag aag ctc tac 5143
 Met Met Glu Glu Lys Phe Met Glu Ala Asn Pro Phe Lys Lys Leu Tyr
 1690 1695 1700

gag ccc ata gtc acc acc acc aag agg aag gag gag gag caa ggc gcc 5191
 Glu Pro Ile Val Thr Thr Thr Lys Arg Lys Glu Glu Glu Gln Gly Ala
 1705 1710 1715

gcc gtc atc cag agg gcc tac cgg aaa cac atg gag aag atg gtc aaa 5239
 Ala Val Ile Gln Arg Ala Tyr Arg Lys His Met Glu Lys Met Val Lys
 1720 1725 1730

ctg agg ctg aag gac agg tca agt tca tcg cac cag gtg ttt tgc aat 5287
 Leu Arg Leu Lys Asp Arg Ser Ser Ser Ser His Gln Val Phe Cys Asn
 1735 1740 1745

gga gac ttg tcc agc ttg gat gtg gcc aag gtc aag gtt cac aat gac 5335
 Gly Asp Leu Ser Ser Leu Asp Val Ala Lys Val Lys Val His Asn Asp
 1750 1755 1760 1765

tgaaccctca tctccacccc tacctcactg cctcacagct tagcctccag cctctggcga 5395

gcaggcggca gactcactga acacaggccg ttcgatctgt gtttttggct gaacgaggtg 5455

acaggttggc gtccattttt aaatgactct tggaagatt tcatgtagag agatgttaga 5515

agggactgca aaggacaccg accataacgg aaggcctgga ggacagtcca acttacataa 5575

agatgagaaa caagaaggaa agatcccagg aaaacttcag attgtgttct cagtacattc 5635

cccaatgtgt ctgttcggtg ttttgagtat gtgacctgcc acatgtagct cttttttgca 5695

tgtacgtcaa aaccctgcag taagttaata gcttgctacg ggtgttccta ccagcatcac 5755

agaattgggt gtatgactca aacctaaaag catgactctg acttgctcagt cagcaccocg 5815

actttcagac gctccaatct ctgtcccagg tgtctaacga ataaataggt aaaagaaaaa 5875

<210> 2

<211> 1765

<212> PRT

<213> Rattus norvegicus

<400> 2

```

Met Glu Glu Arg Tyr Tyr Pro Val Ile Phe Pro Asp Glu Arg Asn Phe
 1           5           10           15

Arg Pro Phe Thr Ser Asp Ser Leu Ala Ala Ile Glu Lys Arg Ile Ala
          20           25           30

Ile Gln Lys Glu Arg Lys Lys Ser Lys Asp Lys Ala Ala Ala Glu Pro
          35           40           45

Gln Pro Arg Pro Gln Leu Asp Leu Lys Ala Ser Arg Lys Leu Pro Lys
          50           55           60

Leu Tyr Gly Asp Ile Pro Pro Glu Leu Val Ala Lys Pro Leu Glu Asp
          65           70           75           80

Leu Asp Pro Phe Tyr Lys Asp His Lys Thr Phe Met Val Leu Asn Lys
          85           90           95

Lys Arg Thr Ile Tyr Arg Phe Ser Ala Lys Arg Ala Leu Phe Ile Leu
          100          105          110

Gly Pro Phe Asn Pro Leu Arg Ser Leu Met Ile Arg Ile Ser Val His
          115          120          125

Ser Val Phe Ser Met Phe Ile Ile Cys Thr Val Ile Ile Asn Cys Met
          130          135          140

Phe Met Ala Asn Ser Met Glu Arg Ser Phe Asp Asn Asp Ile Pro Glu
          145          150          155          160

Tyr Val Phe Ile Gly Ile Tyr Ile Leu Glu Ala Val Ile Lys Ile Leu
          165          170          175

Ala Arg Gly Phe Ile Val Asp Glu Phe Ser Phe Leu Arg Asp Pro Trp
          180          185          190

Asn Trp Leu Asp Phe Ile Val Ile Gly Thr Ala Ile Ala Thr Cys Phe
          195          200          205

Pro Gly Ser Gln Val Asn Leu Ser Ala Leu Arg Thr Phe Arg Val Phe
          210          215          220

Arg Ala Leu Lys Ala Ile Ser Val Ile Ser Gly Leu Lys Val Ile Val
          225          230          235          240

Gly Ala Leu Leu Arg Ser Val Lys Lys Leu Val Asp Val Met Val Leu
          245          250          255

Thr Leu Phe Cys Leu Ser Ile Phe Ala Leu Val Gly Gln Gln Leu Phe
          260          265          270

Met Gly Ile Leu Asn Gln Lys Cys Ile Lys His Asn Cys Gly Pro Asn
          275          280          285

Pro Ala Ser Asn Lys Asp Cys Phe Glu Lys Glu Lys Asp Ser Glu Asp

```

290	295	300
Phe Ile Met Cys Gly Thr Trp Leu Gly Ser Arg Pro Cys Pro Asn Gly 305 310 315 320		
Ser Thr Cys Asp Lys Thr Thr Leu Asn Pro Asp Asn Asn Tyr Thr Lys 325 330 335		
Phe Asp Asn Phe Gly Trp Ser Phe Leu Ala Met Phe Arg Val Met Thr 340 345 350		
Gln Asp Ser Trp Glu Arg Leu Tyr Arg Gln Ile Leu Arg Thr Ser Gly 355 360 365		
Ile Tyr Phe Val Phe Phe Phe Val Val Val Ile Phe Leu Gly Ser Phe 370 375 380		
Tyr Leu Leu Asn Leu Thr Leu Ala Val Val Thr Met Ala Tyr Glu Glu 385 390 395 400		
Gln Asn Arg Asn Val Ala Ala Glu Thr Glu Ala Lys Glu Lys Met Phe 405 410 415		
Gln Glu Ala Gln Gln Leu Leu Arg Glu Glu Lys Glu Ala Leu Val Ala 420 425 430		
Met Gly Ile Asp Arg Ser Ser Leu Asn Ser Leu Gln Ala Ser Ser Phe 435 440 445		
Ser Pro Lys Lys Arg Lys Phe Phe Gly Ser Lys Thr Arg Lys Ser Phe 450 455 460		
Phe Met Arg Gly Ser Lys Thr Ala Gln Ala Ser Ala Ser Asp Ser Glu 465 470 475 480		
Asp Asp Ala Ser Lys Asn Pro Gln Leu Leu Glu Gln Thr Lys Arg Leu 485 490 495		
Ser Gln Asn Leu Pro Val Asp Leu Phe Asp Glu His Val Asp Pro Leu 500 505 510		
His Arg Gln Arg Ala Leu Ser Ala Val Ser Ile Leu Thr Ile Thr Met 515 520 525		
Gln Glu Gln Glu Lys Phe Gln Glu Pro Cys Phe Pro Cys Gly Lys Asn 530 535 540		
Leu Ala Ser Lys Tyr Leu Val Trp Asp Cys Ser Pro Gln Trp Leu Cys 545 550 555 560		
Ile Lys Lys Val Leu Arg Thr Ile Met Thr Asp Pro Phe Thr Glu Leu 565 570 575		
Ala Ile Thr Ile Cys Ile Ile Ile Asn Thr Val Phe Leu Ala Val Glu 580 585 590		

His His Asn Met Asp Asp Asn Leu Lys Thr Ile Leu Lys Ile Gly Asn
 595 600 605
 Trp Val Phe Thr Gly Ile Phe Ile Ala Glu Met Cys Leu Lys Ile Ile
 610 615 620
 Ala Leu Asp Pro Tyr His Tyr Phe Arg His Gly Trp Asn Val Phe Asp
 625 630 635 640
 Ser Ile Val Ala Leu Leu Ser Leu Ala Asp Val Xaa Tyr Asn Thr Leu
 645 650 655
 Ser Asp Asn Asn Arg Ser Phe Leu Ala Ser Leu Arg Val Leu Arg Val
 660 665 670
 Phe Lys Leu Ala Lys Ser Trp Pro Thr Leu Asn Thr Leu Ile Lys Ile
 675 680 685
 Ile Gly His Ser Val Gly Ala Leu Gly Asn Leu Thr Val Val Leu Thr
 690 695 700
 Ile Val Val Phe Ile Phe Ser Val Val Gly Met Arg Leu Phe Gly Thr
 705 710 715 720
 Lys Phe Asn Lys Thr Ala Tyr Ala Thr Gln Glu Arg Pro Arg Arg Arg
 725 730 735
 Trp His Met Asp Asn Phe Tyr His Ser Phe Leu Val Val Phe Arg Ile
 740 745 750
 Leu Cys Gly Glu Trp Ile Glu Asn Met Trp Gly Cys Met Gln Asp Met
 755 760 765
 Asp Gly Ser Pro Leu Cys Ile Ile Val Phe Val Leu Ile Met Val Ile
 770 775 780
 Gly Lys Leu Val Val Leu Asn Leu Phe Ile Ala Leu Leu Leu Asn Ser
 785 790 795 800
 Phe Ser Asn Glu Glu Lys Asp Gly Ser Leu Glu Gly Glu Thr Arg Lys
 805 810 815
 Thr Lys Val Gln Leu Ala Leu Asp Arg Phe Arg Arg Ala Phe Ser Phe
 820 825 830
 Met Leu His Ala Leu Gln Ser Phe Cys Cys Lys Lys Cys Arg Arg Lys
 835 840 845
 Asn Ser Pro Lys Pro Lys Glu Thr Thr Glu Ser Phe Ala Gly Glu Asn
 850 855 860
 Lys Asp Ser Ile Leu Pro Asp Ala Arg Pro Trp Lys Glu Tyr Asp Thr
 865 870 875 880
 Asp Met Ala Leu Tyr Thr Gly Gln Ala Gly Ala Pro Leu Ala Pro Leu
 885 890 895

Ala Glu Val Glu Asp Asp Val Glu Tyr Cys Gly Glu Gly Gly Ala Leu
 900 905 910

Pro Thr Ser Gln His Ser Ala Gly Val Gln Ala Gly Asp Leu Pro Pro
 915 920 925

Glu Thr Lys Gln Leu Thr Ser Pro Asp Asp Gln Gly Val Glu Met Glu
 930 935 940

Val Phe Ser Glu Glu Asp Leu His Leu Ser Ile Gln Ser Pro Arg Lys
 945 950 955 960

Lys Ser Asp Ala Val Ser Met Leu Ser Glu Cys Ser Thr Ile Asp Leu
 965 970 975

Asn Asp Ile Phe Arg Asn Leu Gln Lys Thr Val Ser Pro Lys Lys Gln
 980 985 990

Pro Asp Arg Cys Phe Pro Lys Gly Leu Ser Cys His Phe Leu Cys His
 995 1000 1005

Lys Thr Asp Lys Arg Lys Ser Pro Trp Val Leu Trp Trp Asn Ile Arg
 1010 1015 1020

Lys Thr Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile
 1025 1030 1035 1040

Ile Phe Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val
 1045 1050 1055

Asn Leu Pro Ser Arg Pro Gln Val Glu Lys Leu Leu Arg Cys Thr Asp
 1060 1065 1070

Asn Ile Phe Thr Phe Ile Phe Leu Leu Glu Met Ile Leu Lys Trp Val
 1075 1080 1085

Ala Phe Gly Phe Arg Arg Tyr Phe Thr Ser Ala Trp Cys Trp Leu Asp
 1090 1095 1100

Phe Leu Ile Val Val Val Ser Val Leu Ser Leu Met Asn Leu Pro Ser
 1105 1110 1115 1120

Leu Lys Ser Phe Arg Thr Leu Arg Ala Leu Arg Pro Leu Arg Ala Leu
 1125 1130 1135

Ser Gln Phe Glu Gly Met Lys Val Val Val Tyr Ala Leu Ile Ser Ala
 1140 1145 1150

Ile Pro Ala Ile Leu Asn Val Leu Leu Val Cys Leu Ile Phe Trp Leu
 1155 1160 1165

Val Phe Cys Ile Leu Gly Val Asn Leu Phe Ser Gly Lys Phe Gly Arg
 1170 1175 1180

Cys Ile Asn Gly Thr Asp Ile Asn Met Tyr Leu Asp Phe Thr Glu Val
 1185 1190 1195 1200

Pro Asn Arg Ser Gln Cys Asn Ile Ser Asn Tyr Ser Trp Lys Val Pro
 1205 1210 1215
 Gln Val Asn Phe Asp Asn Val Gly Asn Ala Tyr Leu Ala Leu Leu Gln
 1220 1225 1230
 Val Ala Thr Tyr Lys Gly Trp Leu Glu Ile Met Asn Ala Ala Val Asp
 1235 1240 1245
 Ser Arg Glu Lys Asp Glu Gln Pro Asp Phe Glu Ala Asn Leu Tyr Ala
 1250 1255 1260
 Tyr Leu Tyr Phe Val Val Phe Ile Ile Phe Gly Ser Phe Phe Thr Leu
 1265 1270 1275 1280
 Asn Leu Phe Ile Gly Val Ile Ile Asp Asn Phe Asn Gln Gln Gln Lys
 1285 1290 1295
 Lys Leu Gly Gly Gln Asp Ile Phe Met Thr Glu Glu Gln Lys Lys Tyr
 1300 1305 1310
 Tyr Asn Ala Met Lys Lys Leu Gly Thr Lys Lys Pro Gln Lys Pro Ile
 1315 1320 1325
 Pro Arg Pro Leu Asn Xaa Cys Gln Ala Phe Val Phe Asp Leu Val Thr
 1330 1335 1340
 Ser His Val Phe Asp Val Ile Ile Leu Gly Leu Ile Val Leu Asn Met
 1345 1350 1355 1360
 Ile Ile Met Met Ala Glu Ser Ala Asp Gln Pro Lys Asp Val Lys Lys
 1365 1370 1375
 Thr Phe Asp Ile Leu Asn Ile Ala Phe Val Val Ile Phe Thr Ile Glu
 1380 1385 1390
 Cys Leu Ile Lys Val Phe Ala Leu Arg Gln His Tyr Phe Thr Asn Gly
 1395 1400 1405
 Trp Asn Leu Phe Asp Cys Val Val Val Val Leu Ser Ile Ile Ser Thr
 1410 1415 1420
 Leu Val Ser Arg Leu Glu Asp Ser Asp Ile Ser Phe Pro Pro Thr Leu
 1425 1430 1435 1440
 Phe Arg Val Val Arg Leu Ala Arg Ile Gly Arg Ile Leu Arg Leu Val
 1445 1450 1455
 Arg Ala Ala Arg Gly Ile Arg Thr Leu Leu Phe Ala Leu Met Met Ser
 1460 1465 1470
 Leu Pro Ser Leu Phe Asn Ile Gly Leu Leu Leu Phe Leu Val Met Phe
 1475 1480 1485
 Ile Tyr Ala Ile Phe Gly Met Ser Trp Phe Ser Lys Val Lys Lys Gly
 1490 1495 1500

Ser Gly Ile Asp Asp Ile Phe Asn Phe Glu Thr Phe Thr Gly Ser Met
 1505 1510 1515 1520
 Leu Cys Leu Phe Gln Ile Thr Thr Ser Ala Gly Trp Asp Thr Leu Leu
 1525 1530 1535
 Asn Pro Met Leu Glu Ala Lys Glu His Cys Asn Ser Ser Ser Gln Asp
 1540 1545 1550
 Ser Cys Gln Gln Pro Gln Ile Ala Val Val Tyr Phe Val Ser Tyr Ile
 1555 1560 1565
 Ile Ile Ser Phe Leu Ile Val Val Asn Met Tyr Ile Ala Val Ile Leu
 1570 1575 1580
 Glu Asn Phe Asn Thr Ala Thr Glu Glu Ser Glu Asp Pro Leu Gly Glu
 1585 1590 1595 1600
 Asp Asp Phe Glu Ile Phe Tyr Glu Val Trp Glu Lys Phe Asp Pro Glu
 1605 1610 1615
 Ala Ser Gln Phe Ile Gln Tyr Ser Ala Leu Ser Asp Phe Ala Asp Ala
 1620 1625 1630
 Leu Pro Glu Pro Leu Arg Val Ala Lys Pro Asn Lys Phe Gln Phe Leu
 1635 1640 1645
 Val Met Asp Leu Pro Met Val Met Gly Asp Arg Leu His Cys Met Asp
 1650 1655 1660
 Val Leu Phe Ala Phe Thr Thr Arg Val Leu Gly Asp Ser Ser Gly Leu
 1665 1670 1675 1680
 Asp Thr Met Lys Thr Met Met Glu Glu Lys Phe Met Glu Ala Asn Pro
 1685 1690 1695
 Phe Lys Lys Leu Tyr Glu Pro Ile Val Thr Thr Thr Lys Arg Lys Glu
 1700 1705 1710
 Glu Glu Gln Gly Ala Ala Val Ile Gln Arg Ala Tyr Arg Lys His Met
 1715 1720 1725
 Glu Lys Met Val Lys Leu Arg Leu Lys Asp Arg Ser Ser Ser Ser His
 1730 1735 1740
 Gln Val Phe Cys Asn Gly Asp Leu Ser Ser Leu Asp Val Ala Lys Val
 1745 1750 1755 1760
 Lys Val His Asn Asp
 1765

<210> 3
 <211> 1765
 <212> PRT

<213> Rattus norvegicus

<220>

<223> putative amino acid seq. of rat NaN

<400> 3

```

Met Glu Glu Arg Tyr Tyr Pro Val Ile Phe Pro Asp Glu Arg Asn Phe
 1             5             10             15

Arg Pro Phe Thr Ser Asp Ser Leu Ala Ala Ile Glu Lys Arg Ile Ala
      20             25             30

Ile Gln Lys Glu Arg Lys Lys Ser Lys Asp Lys Ala Ala Ala Glu Pro
      35             40             45

Gln Pro Arg Pro Gln Leu Asp Leu Lys Ala Ser Arg Lys Leu Pro Lys
      50             55             60

Leu Tyr Gly Asp Ile Pro Pro Glu Leu Val Ala Lys Pro Leu Glu Asp
      65             70             75             80

Leu Asp Pro Phe Tyr Lys Asp His Lys Thr Phe Met Val Leu Asn Lys
      85             90             95

Lys Arg Thr Ile Tyr Arg Phe Ser Ala Lys Arg Ala Leu Phe Ile Leu
      100            105            110

Gly Pro Phe Asn Pro Leu Arg Ser Leu Met Ile Arg Ile Ser Val His
      115            120            125

Ser Val Phe Ser Met Phe Ile Ile Cys Thr Val Ile Ile Asn Cys Met
      130            135            140

Phe Met Ala Asn Ser Met Glu Arg Ser Phe Asp Asn Asp Ile Pro Glu
      145            150            155            160

Tyr Val Phe Ile Gly Ile Tyr Ile Leu Glu Ala Val Ile Lys Ile Leu
      165            170            175

Ala Arg Gly Phe Ile Val Asp Glu Phe Ser Phe Leu Arg Asp Pro Trp
      180            185            190

Asn Trp Leu Asp Phe Ile Val Ile Gly Thr Ala Ile Ala Thr Cys Phe
      195            200            205

Pro Gly Ser Gln Val Asn Leu Ser Ala Leu Arg Thr Phe Arg Val Phe
      210            215            220

Arg Ala Leu Lys Ala Ile Ser Val Ile Ser Gly Leu Lys Val Ile Val
      225            230            235            240

Gly Ala Leu Leu Arg Ser Val Lys Lys Leu Val Asp Val Met Val Leu
      245            250            255

Thr Leu Phe Cys Leu Ser Ile Phe Ala Leu Val Gly Gln Gln Leu Phe
      260            265            270

```

Met Gly Ile Leu Asn Gln Lys Cys Ile Lys His Asn Cys Gly Pro Asn
 275 280 285
 Pro Ala Ser Asn Lys Asp Cys Phe Glu Lys Glu Lys Asp Ser Glu Asp
 290 295 300
 Phe Ile Met Cys Gly Thr Trp Leu Gly Ser Arg Pro Cys Pro Asn Gly
 305 310 315 320
 Ser Thr Cys Asp Lys Thr Thr Leu Asn Pro Asp Asn Asn Tyr Thr Lys
 325 330 335
 Phe Asp Asn Phe Gly Trp Ser Phe Leu Ala Met Phe Arg Val Met Thr
 340 345 350
 Gln Asp Ser Trp Glu Arg Leu Tyr Arg Gln Ile Leu Arg Thr Ser Gly
 355 360 365
 Ile Tyr Phe Val Phe Phe Phe Val Val Val Ile Phe Leu Gly Ser Phe
 370 375 380
 Tyr Leu Leu Asn Leu Thr Leu Ala Val Val Thr Met Ala Tyr Glu Glu
 385 390 395 400
 Gln Asn Arg Asn Val Ala Ala Glu Thr Glu Ala Lys Glu Lys Met Phe
 405 410 415
 Gln Glu Ala Gln Gln Leu Leu Arg Glu Glu Lys Glu Ala Leu Val Ala
 420 425 430
 Met Gly Ile Asp Arg Ser Ser Leu Asn Ser Leu Gln Ala Ser Ser Phe
 435 440 445
 Ser Pro Lys Lys Arg Lys Phe Phe Gly Ser Lys Thr Arg Lys Ser Phe
 450 455 460
 Phe Met Arg Gly Ser Lys Thr Ala Gln Ala Ser Ala Ser Asp Ser Glu
 465 470 475 480
 Asp Asp Ala Ser Lys Asn Pro Gln Leu Leu Glu Gln Thr Lys Arg Leu
 485 490 495
 Ser Gln Asn Leu Pro Val Asp Leu Phe Asp Glu His Val Asp Pro Leu
 500 505 510
 His Arg Gln Arg Ala Leu Ser Ala Val Ser Ile Leu Thr Ile Thr Met
 515 520 525
 Gln Glu Gln Glu Lys Phe Gln Glu Pro Cys Phe Pro Cys Gly Lys Asn
 530 535 540
 Leu Ala Ser Lys Tyr Leu Val Trp Asp Cys Ser Pro Gln Trp Leu Cys
 545 550 555 560
 Ile Lys Lys Val Leu Arg Thr Ile Met Thr Asp Pro Phe Thr Glu Leu
 565 570 575

Ala Ile Thr Ile Cys Ile Ile Ile Asn Thr Val Phe Leu Ala Val Glu
 580 585 590
 His His Asn Met Asp Asp Asn Leu Lys Thr Ile Leu Lys Ile Gly Asn
 595 600 605
 Trp Val Phe Thr Gly Ile Phe Ile Ala Glu Met Cys Leu Lys Ile Ile
 610 615 620
 Ala Leu Asp Pro Tyr His Tyr Phe Arg His Gly Trp Asn Val Phe Asp
 625 630 635 640
 Ser Ile Val Ala Leu Leu Ser Leu Ala Asp Val Leu Tyr Asn Thr Leu
 645 650 655
 Ser Asp Asn Asn Arg Ser Phe Leu Ala Ser Leu Arg Val Leu Arg Val
 660 665 670
 Phe Lys Leu Ala Lys Ser Trp Pro Thr Leu Asn Thr Leu Ile Lys Ile
 675 680 685
 Ile Gly His Ser Val Gly Ala Leu Gly Asn Leu Thr Val Val Leu Thr
 690 695 700
 Ile Val Val Phe Ile Phe Ser Val Val Gly Met Arg Leu Phe Gly Thr
 705 710 715 720
 Lys Phe Asn Lys Thr Ala Tyr Ala Thr Gln Glu Arg Pro Arg Arg Arg
 725 730 735
 Trp His Met Asp Asn Phe Tyr His Ser Phe Leu Val Val Phe Arg Ile
 740 745 750
 Leu Cys Gly Glu Trp Ile Glu Asn Met Trp Gly Cys Met Gln Asp Met
 755 760 765
 Asp Gly Ser Pro Leu Cys Ile Ile Val Phe Val Leu Ile Met Val Ile
 770 775 780
 Gly Lys Leu Val Val Leu Asn Leu Phe Ile Ala Leu Leu Leu Asn Ser
 785 790 795 800
 Phe Ser Asn Glu Glu Lys Asp Gly Ser Leu Glu Gly Glu Thr Arg Lys
 805 810 815
 Thr Lys Val Gln Leu Ala Leu Asp Arg Phe Arg Arg Ala Phe Ser Phe
 820 825 830
 Met Leu His Ala Leu Gln Ser Phe Cys Cys Lys Lys Cys Arg Arg Lys
 835 840 845
 Asn Ser Pro Lys Pro Lys Glu Thr Thr Glu Ser Phe Ala Gly Glu Asn
 850 855 860
 Lys Asp Ser Ile Leu Pro Asp Ala Arg Pro Trp Lys Glu Tyr Asp Thr
 865 870 875 880

Asp Met Ala Leu Tyr Thr Gly Gln Ala Gly Ala Pro Leu Ala Pro Leu
 885 890 895
 Ala Glu Val Glu Asp Asp Val Glu Tyr Cys Gly Glu Gly Gly Ala Leu
 900 905 910
 Pro Thr Ser Gln His Ser Ala Gly Val Gln Ala Gly Asp Leu Pro Pro
 915 920 925
 Glu Thr Lys Gln Leu Thr Ser Pro Asp Asp Gln Gly Val Glu Met Glu
 930 935 940
 Val Phe Ser Glu Glu Asp Leu His Leu Ser Ile Gln Ser Pro Arg Lys
 945 950 955 960
 Lys Ser Asp Ala Val Ser Met Leu Ser Glu Cys Ser Thr Ile Asp Leu
 965 970 975
 Asn Asp Ile Phe Arg Asn Leu Gln Lys Thr Val Ser Pro Lys Lys Gln
 980 985 990
 Pro Asp Arg Cys Phe Pro Lys Gly Leu Ser Cys His Phe Leu Cys His
 995 1000 1005
 Lys Thr Asp Lys Arg Lys Ser Pro Trp Val Leu Trp Trp Asn Ile Arg
 1010 1015 1020
 Lys Thr Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile
 1025 1030 1035 1040
 Ile Phe Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val
 1045 1050 1055
 Asn Leu Pro Ser Arg Pro Gln Val Glu Lys Leu Leu Arg Cys Thr Asp
 1060 1065 1070
 Asn Ile Phe Thr Phe Ile Phe Leu Leu Glu Met Ile Leu Lys Trp Val
 1075 1080 1085
 Ala Phe Gly Phe Arg Arg Tyr Phe Thr Ser Ala Trp Cys Trp Leu Asp
 1090 1095 1100
 Phe Leu Ile Val Val Val Ser Val Leu Ser Leu Met Asn Leu Pro Ser
 1105 1110 1115 1120
 Leu Lys Ser Phe Arg Thr Leu Arg Ala Leu Arg Pro Leu Arg Ala Leu
 1125 1130 1135
 Ser Gln Phe Glu Gly Met Lys Val Val Val Tyr Ala Leu Ile Ser Ala
 1140 1145 1150
 Ile Pro Ala Ile Leu Asn Val Leu Leu Val Cys Leu Ile Phe Trp Leu
 1155 1160 1165
 Val Phe Cys Ile Leu Gly Val Asn Leu Phe Ser Gly Lys Phe Gly Arg
 1170 1175 1180

Cys Ile Asn Gly Thr Asp Ile Asn Met Tyr Leu Asp Phe Thr Glu Val
 1185 1190 1195 1200
 Pro Asn Arg Ser Gln Cys Asn Ile Ser Asn Tyr Ser Trp Lys Val Pro
 1205 1210 1215
 Gln Val Asn Phe Asp Asn Val Gly Asn Ala Tyr Leu Ala Leu Leu Gln
 1220 1225 1230
 Val Ala Thr Tyr Lys Gly Trp Leu Glu Ile Met Asn Ala Ala Val Asp
 1235 1240 1245
 Ser Arg Glu Lys Asp Glu Gln Pro Asp Phe Glu Ala Asn Leu Tyr Ala
 1250 1255 1260
 Tyr Leu Tyr Phe Val Val Phe Ile Ile Phe Gly Ser Phe Phe Thr Leu
 1265 1270 1275 1280
 Asn Leu Phe Ile Gly Val Ile Ile Asp Asn Phe Asn Gln Gln Gln Lys
 1285 1290 1295
 Lys Leu Gly Gly Gln Asp Ile Phe Met Thr Glu Glu Gln Lys Lys Tyr
 1300 1305 1310
 Tyr Asn Ala Met Lys Lys Leu Gly Thr Lys Lys Pro Gln Lys Pro Ile
 1315 1320 1325
 Pro Arg Pro Leu Asn Arg Cys Gln Ala Phe Val Phe Asp Leu Val Thr
 1330 1335 1340
 Ser His Val Phe Asp Val Ile Ile Leu Gly Leu Ile Val Leu Asn Met
 1345 1350 1355 1360
 Ile Ile Met Met Ala Glu Ser Ala Asp Gln Pro Lys Asp Val Lys Lys
 1365 1370 1375
 Thr Phe Asp Ile Leu Asn Ile Ala Phe Val Val Ile Phe Thr Ile Glu
 1380 1385 1390
 Cys Leu Ile Lys Val Phe Ala Leu Arg Gln His Tyr Phe Thr Asn Gly
 1395 1400 1405
 Trp Asn Leu Phe Asp Cys Val Val Val Val Leu Ser Ile Ile Ser Thr
 1410 1415 1420
 Leu Val Ser Arg Leu Glu Asp Ser Asp Ile Ser Phe Pro Pro Thr Leu
 1425 1430 1435 1440
 Phe Arg Val Val Arg Leu Ala Arg Ile Gly Arg Ile Leu Arg Leu Val
 1445 1450 1455
 Arg Ala Ala Arg Gly Ile Arg Thr Leu Leu Phe Ala Leu Met Met Ser
 1460 1465 1470
 Leu Pro Ser Leu Phe Asn Ile Gly Leu Leu Leu Phe Leu Val Met Phe
 1475 1480 1485

Ile Tyr Ala Ile Phe Gly Met Ser Trp Phe Ser Lys Val Lys Lys Gly
 1490 1495 1500
 Ser Gly Ile Asp Asp Ile Phe Asn Phe Glu Thr Phe Thr Gly Ser Met
 1505 1510 1515 1520
 Leu Cys Leu Phe Gln Ile Thr Thr Ser Ala Gly Trp Asp Thr Leu Leu
 1525 1530 1535
 Asn Pro Met Leu Glu Ala Lys Glu His Cys Asn Ser Ser Ser Gln Asp
 1540 1545 1550
 Ser Cys Gln Gln Pro Gln Ile Ala Val Val Tyr Phe Val Ser Tyr Ile
 1555 1560 1565
 Ile Ile Ser Phe Leu Ile Val Val Asn Met Tyr Ile Ala Val Ile Leu
 1570 1575 1580
 Glu Asn Phe Asn Thr Ala Thr Glu Glu Ser Glu Asp Pro Leu Gly Glu
 1585 1590 1595 1600
 Asp Asp Phe Glu Ile Phe Tyr Glu Val Trp Glu Lys Phe Asp Pro Glu
 1605 1610 1615
 Ala Ser Gln Phe Ile Gln Tyr Ser Ala Leu Ser Asp Phe Ala Asp Ala
 1620 1625 1630
 Leu Pro Glu Pro Leu Arg Val Ala Lys Pro Asn Lys Phe Gln Phe Leu
 1635 1640 1645
 Val Met Asp Leu Pro Met Val Met Gly Asp Arg Leu His Cys Met Asp
 1650 1655 1660
 Val Leu Phe Ala Phe Thr Thr Arg Val Leu Gly Asp Ser Ser Gly Leu
 1665 1670 1675 1680
 Asp Thr Met Lys Thr Met Met Glu Glu Lys Phe Met Glu Ala Asn Pro
 1685 1690 1695
 Phe Lys Lys Leu Tyr Glu Pro Ile Val Thr Thr Thr Lys Arg Lys Glu
 1700 1705 1710
 Glu Glu Gln Gly Ala Ala Val Ile Gln Arg Ala Tyr Arg Lys His Met
 1715 1720 1725
 Glu Lys Met Val Lys Leu Arg Leu Lys Asp Arg Ser Ser Ser Ser His
 1730 1735 1740
 Gln Val Phe Cys Asn Gly Asp Leu Ser Ser Leu Asp Val Ala Lys Val
 1745 1750 1755 1760
 Lys Val His Asn Asp
 1765

<210> 4

<211> 5822

<212> DNA

<213> Mus musculus

<220>

<221> CDS

<222> (19) .. (5313)

<220>

<221> unsure

<222> (5804)

<223> cDNA sequence of mouse NaN, n = a or c or g or t

<400> 4

```

tctgagccaa ggggtgaag atg gag gag agg tac tat cca gtg atc ttc cca 51
      Met Glu Glu Arg Tyr Pro Val Ile Phe Pro
        1             5             10

gac gag agg aat ttc cgc ccc ttc act ttc gac tct ttg gct gca ata 99
Asp Glu Arg Asn Phe Arg Pro Phe Thr Phe Asp Ser Leu Ala Ala Ile
      15             20             25

gag aag cgg atc acc atc caa aag gag aag aag aaa tcc aaa gac aag 147
Glu Lys Arg Ile Thr Ile Gln Lys Glu Lys Lys Lys Ser Lys Asp Lys
      30             35             40

gca gca act gag ccc cag cct cgg cct cag ctc gac cta aag gcc tcc 195
Ala Ala Thr Glu Pro Gln Pro Arg Pro Gln Leu Asp Leu Lys Ala Ser
      45             50             55

agg aag tta cct aag ctc tat ggc gac gtt ccc cct gac ctt ata gcg 243
Arg Lys Leu Pro Lys Leu Tyr Gly Asp Val Pro Pro Asp Leu Ile Ala
      60             65             70             75

aag ccc ctg gaa gat ctg gac cca ttt tac aaa gac cat aag aca ttc 291
Lys Pro Leu Glu Asp Leu Asp Pro Phe Tyr Lys Asp His Lys Thr Phe
      80             85             90

atg gta ttg aac aag aag aga aca atc tat cgc ttc agc gcc aag agg 339
Met Val Leu Asn Lys Lys Arg Thr Ile Tyr Arg Phe Ser Ala Lys Arg
      95             100            105

gcc ttg ttc att ctg ggg cct ttt aat ccc atc aga agc ttc atg att 387
Ala Leu Phe Ile Leu Gly Pro Phe Asn Pro Ile Arg Ser Phe Met Ile
      110            115            120

cgc atc tct gtc cat tca gtc ttc agc atg ttc att atc tgc aca gtg 435
Arg Ile Ser Val His Ser Val Phe Ser Met Phe Ile Ile Cys Thr Val
      125            130            135

atc atc aac tgt atg ttc atg gct aat aat tct tct gtg gac agt cgt 483
Ile Ile Asn Cys Met Phe Met Ala Asn Asn Ser Ser Val Asp Ser Arg
      140            145            150            155

cct agc agt aac att ccc gaa tac gtc ttc att ggg att tat gtt tta 531
Pro Ser Ser Asn Ile Pro Glu Tyr Val Phe Ile Gly Ile Tyr Val Leu

```

160	165	170	
gaa gct gtg att aaa ata ttg gca aga ggc ttc att gtg gat gag ttt Glu Ala Val Ile Lys Ile Leu Ala Arg Gly Phe Ile Val Asp Glu Phe 175 180 185			579
tcc tac ctc cga gat cct tgg aac tgg ctg gac ttc att gtc atc gga Ser Tyr Leu Arg Asp Pro Trp Asn Trp Leu Asp Phe Ile Val Ile Gly 190 195 200			627
aca gcg ata gcg cct tgt ttt ctc ggt aac aaa gtc aat aat ctt tcc Thr Ala Ile Ala Pro Cys Phe Leu Gly Asn Lys Val Asn Asn Leu Ser 205 210 215			675
act cta cgt acc ttc cga gtg ttg aga gct ctg aaa gcc att tct gta Thr Leu Arg Thr Phe Arg Val Leu Arg Ala Leu Lys Ala Ile Ser Val 220 225 230 235			723
atc tca ggt ctg aag gtc atc gtg ggt gcc ctg ctg cgc tcc gtg aag Ile Ser Gly Leu Lys Val Ile Val Gly Ala Leu Leu Arg Ser Val Lys 240 245 250			771
aag cta gtg gac gtg atg gtc ctc act ctc ttt tgc ctc agc atc ttt Lys Leu Val Asp Val Met Val Leu Thr Leu Phe Cys Leu Ser Ile Phe 255 260 265			819
gcc ctg gtt ggt cag cag ctc ttc atg gga att ctg agc cag aaa tgt Ala Leu Val Gly Gln Gln Leu Phe Met Gly Ile Leu Ser Gln Lys Cys 270 275 280			867
att aag gac gac tgt ggc cct aac gct ttt tcc aac aag gat tgc ttt Ile Lys Asp Asp Cys Gly Pro Asn Ala Phe Ser Asn Lys Asp Cys Phe 285 290 295			915
gta aaa gaa aat gat agc gag gac ttc ata atg tgt ggc aac tgg ctc Val Lys Glu Asn Asp Ser Glu Asp Phe Ile Met Cys Gly Asn Trp Leu 300 305 310 315			963
ggc aga aga tcc tgc ccc gat ggt tcc acg tgc aat aaa acc aca ttt Gly Arg Arg Ser Cys Pro Asp Gly Ser Thr Cys Asn Lys Thr Thr Phe 320 325 330			1011
aac cca gat tat aat tat aca aac ttt gac agc ttt ggc tgg tct ttt Asn Pro Asp Tyr Asn Tyr Thr Asn Phe Asp Ser Phe Gly Trp Ser Phe 335 340 345			1059
ctc gcc atg ttc cgg gtt atg act caa gac tcc tgg gag aag ctt tat Leu Ala Met Phe Arg Val Met Thr Gln Asp Ser Trp Glu Lys Leu Tyr 350 355 360			1107
cga cag atc ctt cgc acc tcc ggg atc tac ttt gtc ttc ttc ttc gtg Arg Gln Ile Leu Arg Thr Ser Gly Ile Tyr Phe Val Phe Phe Phe Val 365 370 375			1155
gtc gtc atc ttc ctg ggc tct ttc tac ctg ctt aac tta acc ctg gct Val Val Ile Phe Leu Gly Ser Phe Tyr Leu Leu Asn Leu Thr Leu Ala			1203

380	385	390	395	
gtc gtc acc atg gct tac gag gaa cag aac aga aat gtc gct gcc gag				1251
Val Val Thr Met Ala Tyr Glu Glu Gln Asn Arg Asn Val Ala Ala Glu				
	400	405	410	
aca gag gcc aag gag aag atg ttt cag gaa gcc cag cag ctg ttg agg				1299
Thr Glu Ala Lys Glu Lys Met Phe Gln Glu Ala Gln Gln Leu Leu Arg				
	415	420	425	
gag gaa aag gag gct ctg gtt gcc atg gga att gac aga act tcc ctt				1347
Glu Glu Lys Glu Ala Leu Val Ala Met Gly Ile Asp Arg Thr Ser Leu				
	430	435	440	
aat tcc ctc caa gct tcg tcc ttt tcc cca aag aag agg aag ttt ttt				1395
Asn Ser Leu Gln Ala Ser Ser Phe Ser Pro Lys Lys Arg Lys Phe Phe				
	445	450	455	
ggc agt aag aca aga aag tcc ttc ttt atg aga ggg tcc aag aca gcc				1443
Gly Ser Lys Thr Arg Lys Ser Phe Phe Met Arg Gly Ser Lys Thr Ala				
	460	465	470	475
cga gcc tca gcg tcc gat tca gag gac gat gcc tct aaa aac cca caa				1491
Arg Ala Ser Ala Ser Asp Ser Glu Asp Asp Ala Ser Lys Asn Pro Gln				
	480	485	490	
ctc ctt gag caa aca aaa cga cta tcc cag aac ttg ccc gta gaa ctc				1539
Leu Leu Glu Gln Thr Lys Arg Leu Ser Gln Asn Leu Pro Val Glu Leu				
	495	500	505	
ttt gat gag cac gtg gac ccc ctc cat agg cag aga gcg ctg agt gcc				1587
Phe Asp Glu His Val Asp Pro Leu His Arg Gln Arg Ala Leu Ser Ala				
	510	515	520	
gtc agt atc tta acc atc acc atg cag gaa caa gaa aaa tcc cag gag				1635
Val Ser Ile Leu Thr Ile Thr Met Gln Glu Gln Glu Lys Ser Gln Glu				
	525	530	535	
cct tgt ttc ccg tgt ggg aaa aac ttg gca tcc aag tac ctg gtg tgg				1683
Pro Cys Phe Pro Cys Gly Lys Asn Leu Ala Ser Lys Tyr Leu Val Trp				
	540	545	550	555
gaa tgt agc cct ccg tgg ctg tgc ata aag aag gtc ctg cag act atc				1731
Glu Cys Ser Pro Pro Trp Leu Cys Ile Lys Lys Val Leu Gln Thr Ile				
	560	565	570	
atg aca gac ccc ttc act gag ctg gcc atc acc atc tgc atc atc gtc				1779
Met Thr Asp Pro Phe Thr Glu Leu Ala Ile Thr Ile Cys Ile Ile Val				
	575	580	585	
aat act gtc ttc ttg gcc atg gaa cac cac aat atg gat aac tct tta				1827
Asn Thr Val Phe Leu Ala Met Glu His His Asn Met Asp Asn Ser Leu				
	590	595	600	
aaa gac ata ctg aaa ata gga aac tgg gtt ttc act gga att ttc ata				1875
Lys Asp Ile Leu Lys Ile Gly Asn Trp Val Phe Thr Gly Ile Phe Ile				

605	610	615	
gcg gaa atg tgt ctc aag atc att gcg cta gac cct tac cac tac ttc			1923
Ala Glu Met Cys Leu Lys Ile Ile Ala Leu Asp Pro Tyr His Tyr Phe			
620	625	630	635
cgg cac ggc tgg aac atc ttt gac agc att gtg gcc ctt gtg agt ctc			1971
Arg His Gly Trp Asn Ile Phe Asp Ser Ile Val Ala Leu Val Ser Leu			
	640	645	650
gct gac gtg ctc ttc cac aaa ctg tct aaa aac ctc tcc ttc ttg gct			2019
Ala Asp Val Leu Phe His Lys Leu Ser Lys Asn Leu Ser Phe Leu Ala			
	655	660	665
tcc ctc aga gtg ctg agg gtc ttc aag tta gcc aaa tcc tgg ccc aca			2067
Ser Leu Arg Val Leu Arg Val Phe Lys Leu Ala Lys Ser Trp Pro Thr			
	670	675	680
tta aac act ctc att aag atc atc ggc cac tcc gtg ggt gcg ctc gga			2115
Leu Asn Thr Leu Ile Lys Ile Ile Gly His Ser Val Gly Ala Leu Gly			
	685	690	695
aac ctg act gtg gtc cta acg atc gtg gtc ttc atc ttt tcc gtg gtt			2163
Asn Leu Thr Val Val Leu Thr Ile Val Val Phe Ile Phe Ser Val Val			
	700	705	710
ggc atg cgg ctc ttt ggt gcc aag ttt aac aag act tgc tcc acc tct			2211
Gly Met Arg Leu Phe Gly Ala Lys Phe Asn Lys Thr Cys Ser Thr Ser			
	720	725	730
ccg gag tcc ctc cgg cgc tgg cac atg ggt gat ttc tac cat tcc ttc			2259
Pro Glu Ser Leu Arg Arg Trp His Met Gly Asp Phe Tyr His Ser Phe			
	735	740	745
ctg gtg gtg ttc cgc atc ctc tgt ggg gag tgg atc gag aac atg tgg			2307
Leu Val Val Phe Arg Ile Leu Cys Gly Glu Trp Ile Glu Asn Met Trp			
	750	755	760
gaa tgc atg cag gag atg gaa ggc tcc ccg ctg tgt gtc atc gtc ttt			2355
Glu Cys Met Gln Glu Met Glu Gly Ser Pro Leu Cys Val Ile Val Phe			
	765	770	775
gtg ctg atc atg gtg gtc ggg aag ctc gtg gtg ctt aac ctc ttc att			2403
Val Leu Ile Met Val Val Gly Lys Leu Val Val Leu Asn Leu Phe Ile			
	780	785	790
gcc ttg ctg ctc aat tcc ttc agc aat gag gaa aag gat ggg aac cca			2451
Ala Leu Leu Leu Asn Ser Phe Ser Asn Glu Glu Lys Asp Gly Asn Pro			
	800	805	810
gaa gga gag acc agg aaa acc aaa gtg cag cta gcc ctg gat cgg ttc			2499
Glu Gly Glu Thr Arg Lys Thr Lys Val Gln Leu Ala Leu Asp Arg Phe			
	815	820	825
agc cga gcg ttc tac ttc atg gcg cgc gct ctt cag aat ttc tgt tgc			2547
Ser Arg Ala Phe Tyr Phe Met Ala Arg Ala Leu Gln Asn Phe Cys Cys			

830	835	840	
aag aga tgc agg agg caa aac tcg cca aag cca aat gag gca aca gaa Lys Arg Cys Arg Arg Gln Asn Ser Pro Lys Pro Asn Glu Ala Thr Glu 845 850 855			2595
agc ttt gct ggt gag agt aga gac aca gcc acc ctg gat aca agg tcc Ser Phe Ala Gly Glu Ser Arg Asp Thr Ala Thr Leu Asp Thr Arg Ser 860 865 870 875			2643
tgg aag gag tat gat tca gaa atg act ctg tac act ggg cag gcc ggg Trp Lys Glu Tyr Asp Ser Glu Met Thr Leu Tyr Thr Gly Gln Ala Gly 880 885 890			2691
gct cca ctg gcc cca ctg gca aaa gaa gag gac gat atg gaa tgt tgt Ala Pro Leu Ala Pro Leu Ala Lys Glu Glu Asp Asp Met Glu Cys Cys 895 900 905			2739
ggg gaa tgt gat gcc tca cct acc tca cag cct agt gag gaa gct cag Gly Glu Cys Asp Ala Ser Pro Thr Ser Gln Pro Ser Glu Glu Ala Gln 910 915 920			2787
gcc tgt gac ctc cct ctg aag acc aag cgg ctc ccc agc cca gat gac Ala Cys Asp Leu Pro Leu Lys Thr Lys Arg Leu Pro Ser Pro Asp Asp 925 930 935			2835
cac ggg gtt gaa atg gaa gtg ttt tcc gaa gaa gat ccg aat tta acc His Gly Val Glu Met Glu Val Phe Ser Glu Glu Asp Pro Asn Leu Thr 940 945 950 955			2883
ata cag agt gct cga aag aag tct gat gcg gca agc atg ctc tca gaa Ile Gln Ser Ala Arg Lys Lys Ser Asp Ala Ala Ser Met Leu Ser Glu 960 965 970			2931
tgc agc aca ata gac ctg aat gat atc ttt aga aat tta cag aaa aca Cys Ser Thr Ile Asp Leu Asn Asp Ile Phe Arg Asn Leu Gln Lys Thr 975 980 985			2979
gtt tcc ccc caa aag caa cca gat cga tgc ttt ccc aag ggc ctc agt Val Ser Pro Gln Lys Gln Pro Asp Arg Cys Phe Pro Lys Gly Leu Ser 990 995 1000			3027
tgt atc ttt cta tgt tgc aaa aca atc aaa aaa aag tcc ccc tgg gtc Cys Ile Phe Leu Cys Cys Lys Thr Ile Lys Lys Lys Ser Pro Trp Val 1005 1010 1015			3075
ctg tgg tgg aat ctt cgg aaa acc tgc tac caa atc gtg aag cat agc Leu Trp Trp Asn Leu Arg Lys Thr Cys Tyr Gln Ile Val Lys His Ser 1020 1025 1030 1035			3123
tgg ttt gag agc ttc ata att ttt gtc atc ctg ctg agc agc gga gca Trp Phe Glu Ser Phe Ile Ile Phe Val Ile Leu Leu Ser Ser Gly Ala 1040 1045 1050			3171
ctg ata ttc gaa gat gtc aat ctt ccc agc cgg ccc caa gtt gaa aaa Leu Ile Phe Glu Asp Val Asn Leu Pro Ser Arg Pro Gln Val Glu Lys			3219

1055	1060	1065	
tta ctg aag tgt acc gat aat att ttc aca ttt att ttt ctc ctg gaa Leu Leu Lys Cys Thr Asp Asn Ile Phe Thr Phe Ile Phe Leu Leu Glu 1070	1075	1080	3267
atg att ttg aag tgg gtg gcc ttt gga ttc cgg aag tat ttc acc agt Met Ile Leu Lys Trp Val Ala Phe Gly Phe Arg Lys Tyr Phe Thr Ser 1085	1090	1095	3315
gcc tgg tgc tgg ctc gat ttc ctc att gtg gtg gtg tct gtg ctc agc Ala Trp Cys Trp Leu Asp Phe Leu Ile Val Val Val Ser Val Leu Ser 1100	1105	1110	3363
ctc acg aac tta cca aac ttg aag tcc ttc cgg aat ctg cga gcg ctg Leu Thr Asn Leu Pro Asn Leu Lys Ser Phe Arg Asn Leu Arg Ala Leu 1120	1125	1130	3411
aga cct ctg cgg gca ctg tct cag ttt gaa gga atg aag gtt gtt gtc Arg Pro Leu Arg Ala Leu Ser Gln Phe Glu Gly Met Lys Val Val Val 1135	1140	1145	3459
aat gcc ctc atg agt gcc ata cct gcc atc ctc aat gtc ttg ctg gtc Asn Ala Leu Met Ser Ala Ile Pro Ala Ile Leu Asn Val Leu Leu Val 1150	1155	1160	3507
tgc ctc att ttc tgg ctc ata ttt tgt atc ctg gga gta aat ttt ttt Cys Leu Ile Phe Trp Leu Ile Phe Cys Ile Leu Gly Val Asn Phe Phe 1165	1170	1175	3555
tct ggg aag ttt gga aga tgc att aat gga aca gac ata aat aaa tat Ser Gly Lys Phe Gly Arg Cys Ile Asn Gly Thr Asp Ile Asn Lys Tyr 1180	1185	1190	3603
ttc aac gct tcc aat gtt cca aac caa agc caa tgt tta gtt agt aat Phe Asn Ala Ser Asn Val Pro Asn Gln Ser Gln Cys Leu Val Ser Asn 1200	1205	1210	3651
tac acg tgg aaa gtc ccg aat gtc aac ttt gac aac gtg ggg aat gcc Tyr Thr Trp Lys Val Pro Asn Val Asn Phe Asp Asn Val Gly Asn Ala 1215	1220	1225	3699
tac ctt gcc ctg ctg caa gtg gcg acc tat aag ggc tgg ctg gac att Tyr Leu Ala Leu Leu Gln Val Ala Thr Tyr Lys Gly Trp Leu Asp Ile 1230	1235	1240	3747
atg aat gca gct gtt gat tcc aga ggg aaa gat gag cag ccg gcc ttt Met Asn Ala Ala Val Asp Ser Arg Gly Lys Asp Glu Gln Pro Ala Phe 1245	1250	1255	3795
gag gcg aat cta tac gca tac ctt tac ttc gtg gtt ttt atc atc ttc Glu Ala Asn Leu Tyr Ala Tyr Leu Tyr Phe Val Val Phe Ile Ile Phe 1260	1265	1270	3843
ggc tca ttc ttt acc ctg aac ctc ttt atc ggt gtt att att gac aac Gly Ser Phe Phe Thr Leu Asn Leu Phe Ile Gly Val Ile Ile Asp Asn			3891

1280	1285	1290	
ttc aat cag cag cag aaa aag tta ggt ggc caa gac att ttt atg aca			3939
Phe Asn Gln Gln Gln Lys Lys Leu Gly Gly Gln Asp Ile Phe Met Thr			
1295	1300	1305	
gaa gaa cag aag aaa tat tac aat gca atg aaa aag tta gga acc aag			3987
Glu Glu Gln Lys Lys Tyr Tyr Asn Ala Met Lys Lys Leu Gly Thr Lys			
1310	1315	1320	
aag cct caa aag ccc atc cca agg ccc ctg aac aaa tgt caa gcc ttc			4035
Lys Pro Gln Lys Pro Ile Pro Arg Pro Leu Asn Lys Cys Gln Ala Phe			
1325	1330	1335	
gtg ttc gat ttg gtc aca agc cag gtc ttt gac gtc atc att ctg ggt			4083
Val Phe Asp Leu Val Thr Ser Gln Val Phe Asp Val Ile Ile Leu Gly			
1340	1345	1350	1355
ctt att gtc aca aac atg att atc atg atg gct gaa tct gaa ggc cag			4131
Leu Ile Val Thr Asn Met Ile Ile Met Met Ala Glu Ser Glu Gly Gln			
1360	1365	1370	
ccc aac gaa gtg aag aaa atc ttt gat att ctc aac ata gtc ttc gtg			4179
Pro Asn Glu Val Lys Lys Ile Phe Asp Ile Leu Asn Ile Val Phe Val			
1375	1380	1385	
gtc atc ttt acc gta gag tgt ctc atc aaa gtc ttt gct ttg agg caa			4227
Val Ile Phe Thr Val Glu Cys Leu Ile Lys Val Phe Ala Leu Arg Gln			
1390	1395	1400	
cac tac ttc acc aat ggc tgg aac tta ttt gat tgt gtg gtc gtg gtt			4275
His Tyr Phe Thr Asn Gly Trp Asn Leu Phe Asp Cys Val Val Val Val			
1405	1410	1415	
ott tcc atc att agt acc ttg gtt tct ggc ttg gag aac agc aac gtc			4323
Leu Ser Ile Ile Ser Thr Leu Val Ser Gly Leu Glu Asn Ser Asn Val			
1420	1425	1430	1435
ttc ccg ccc aca ctc ttc agg att gtc cgc ttg gct cgg atc ggt cga			4371
Phe Pro Pro Thr Leu Phe Arg Ile Val Arg Leu Ala Arg Ile Gly Arg			
1440	1445	1450	
atc ctc aga ctg gtc cgg gcg gct cga gga atc agg aca ctc ctt ttc			4419
Ile Leu Arg Leu Val Arg Ala Ala Arg Gly Ile Arg Thr Leu Leu Phe			
1455	1460	1465	
gcg ttg atg atg tct ctc ccc tct ctc ttc aac att ggt ctg ctt ctc			4467
Ala Leu Met Met Ser Leu Pro Ser Leu Phe Asn Ile Gly Leu Leu Leu			
1470	1475	1480	
ttt ctg gtg atg ttc att tat gcc atc ttt ggg atg aac tgg ttt tcc			4515
Phe Leu Val Met Phe Ile Tyr Ala Ile Phe Gly Met Asn Trp Phe Ser			
1485	1490	1495	
aaa gtg aag aga ggc tct ggg att gat gac atc ttc aac ttt gac act			4563
Lys Val Lys Arg Gly Ser Gly Ile Asp Asp Ile Phe Asn Phe Asp Thr			

1500	1505	1510	1515	
ttc tgc ggc agc atg ctc tgc ctc ttc cag ata acc act tca gcc ggc				4611
Phe Ser Gly Ser Met Leu Cys Leu Phe Gln Ile Thr Thr Ser Ala Gly				
1520	1525	1530		
tggtgatgctctctaaccccattgctggaa tca aaa gcc tct tgc aat				4659
Trp Asp Ala Leu Leu Asn Pro Met Leu Glu Ser Lys Ala Ser Cys Asn				
1535	1540	1545		
tcc tcc tcc caa gag agc tgt cag cag ccg cag ata gcc ata gtc tac				4707
Ser Ser Ser Gln Glu Ser Cys Gln Gln Pro Gln Ile Ala Ile Val Tyr				
1550	1555	1560		
ttc gtc agc tac atc atc atc tcc ttt ctc att gtg gtt aac atg tac				4755
Phe Val Ser Tyr Ile Ile Ile Ser Phe Leu Ile Val Val Asn Met Tyr				
1565	1570	1575		
ata gct gtg att cta gag aac ttc aac aca gcc aca gag gag agc gag				4803
Ile Ala Val Ile Leu Glu Asn Phe Asn Thr Ala Thr Glu Glu Ser Glu				
1580	1585	1590	1595	
gac ccc ctg ggc gaa gac gac ttt gag atc ttc tat gag atc tgg gag				4851
Asp Pro Leu Gly Glu Asp Asp Phe Glu Ile Phe Tyr Glu Ile Trp Glu				
1600	1605	1610		
aag ttt gac ccc gaa gca aca cag ttc atc cag tac tca tcc ctc tct				4899
Lys Phe Asp Pro Glu Ala Thr Gln Phe Ile Gln Tyr Ser Ser Leu Ser				
1615	1620	1625		
gac ttc gcc gac gcc ctg ccc gag ccg ttg cgt gtg gcc aag ccc aac				4947
Asp Phe Ala Asp Ala Leu Pro Glu Pro Leu Arg Val Ala Lys Pro Asn				
1630	1635	1640		
agg ttt cag ttt ctc atg atg gac ttg ccc atg gtg atg ggt gat cgc				4995
Arg Phe Gln Phe Leu Met Met Asp Leu Pro Met Val Met Gly Asp Arg				
1645	1650	1655		
ctc cat tgc atg gat gtt ctc ttt gct ttc acc acc agg gtc ctc ggg				5043
Leu His Cys Met Asp Val Leu Phe Ala Phe Thr Thr Arg Val Leu Gly				
1660	1665	1670	1675	
aac tcc agc ggc ttg gat acc atg aaa gcc atg atg gag gag aag ttc				5091
Asn Ser Ser Gly Leu Asp Thr Met Lys Ala Met Met Glu Glu Lys Phe				
1680	1685	1690		
atg gag gcc aat cct ttc aag aag ttg tac gag ccc att gtc acc acc				5139
Met Glu Ala Asn Pro Phe Lys Lys Leu Tyr Glu Pro Ile Val Thr Thr				
1695	1700	1705		
aca aag agg aag gag gag gag gaa tgt gcc gct gtc atc cag agg gcc				5187
Thr Lys Arg Lys Glu Glu Glu Cys Ala Ala Val Ile Gln Arg Ala				
1710	1715	1720		
tac cgg aga cac atg gag aag atg atc aag ctg aag ctg aaa ggc agg				5235
Tyr Arg Arg His Met Glu Lys Met Ile Lys Leu Lys Leu Lys Gly Arg				

1725 1730 1735
 tca agt tca tcg ctc cag gtg ttt tgc aat gga gac ttg tct agc ttg 5283
 Ser Ser Ser Ser Leu Gln Val Phe Cys Asn Gly Asp Leu Ser Ser Leu
 1740 1745 1750 1755
 gat gtg ccc aag atc aag gtt cat tgt gac tgaaaccccc acctgcacgc 5333
 Asp Val Pro Lys Ile Lys Val His Cys Asp
 1760 1765
 ctacctcaca gcctcacagc tcagcccccga gcctctggcg aacaagcggc ggactcacgc 5393
 aacaggccgt tcaacttggt tttttgggtg aaagaggtga taggttggtg tccattttta 5453
 aatgattctt ggaagattg aacgtcggaa catgttagaa aggactgcca aggacatcca 5513
 cagtaacgga aggcctgaag gacagttcaa attatgtaaa gaaacgagaa ggaaagggtca 5573
 catgtctggt cagttttaag tatgtgacct gccacatgta gtccttttgc atgttaagtg 5633
 agaagtcaaa accctgccat aagtaaatac ctttggtgca ggtgtttcta ccagtgtgc 5693
 ggatttgggt gtatgggtca aacctgaaag catgactctg acttgtcagc accccaactt 5753
 tcagaagctc tgatctctgt cctaggtggt tgacaaataa atacataaaa naaaaaaaaa 5813
 aaaaaaaaaa 5822

<210> 5
 <211> 1765
 <212> PRT
 <213> Mus musculus

<400> 5
 Met Glu Glu Arg Tyr Tyr Pro Val Ile Phe Pro Asp Glu Arg Asn Phe
 1 5 10 15
 Arg Pro Phe Thr Phe Asp Ser Leu Ala Ala Ile Glu Lys Arg Ile Thr
 20 25 30
 Ile Gln Lys Glu Lys Lys Lys Ser Lys Asp Lys Ala Ala Thr Glu Pro
 35 40 45
 Gln Pro Arg Pro Gln Leu Asp Leu Lys Ala Ser Arg Lys Leu Pro Lys
 50 55 60
 Leu Tyr Gly Asp Val Pro Pro Asp Leu Ile Ala Lys Pro Leu Glu Asp
 65 70 75 80
 Leu Asp Pro Phe Tyr Lys Asp His Lys Thr Phe Met Val Leu Asn Lys
 85 90 95
 Lys Arg Thr Ile Tyr Arg Phe Ser Ala Lys Arg Ala Leu Phe Ile Leu
 100 105 110

Gly Pro Phe Asn Pro Ile Arg Ser Phe Met Ile Arg Ile Ser Val His
 115 120 125
 Ser Val Phe Ser Met Phe Ile Ile Cys Thr Val Ile Ile Asn Cys Met
 130 135 140
 Phe Met Ala Asn Asn Ser Ser Val Asp Ser Arg Pro Ser Ser Asn Ile
 145 150 155 160
 Pro Glu Tyr Val Phe Ile Gly Ile Tyr Val Leu Glu Ala Val Ile Lys
 165 170 175
 Ile Leu Ala Arg Gly Phe Ile Val Asp Glu Phe Ser Tyr Leu Arg Asp
 180 185 190
 Pro Trp Asn Trp Leu Asp Phe Ile Val Ile Gly Thr Ala Ile Ala Pro
 195 200 205
 Cys Phe Leu Gly Asn Lys Val Asn Asn Leu Ser Thr Leu Arg Thr Phe
 210 215 220
 Arg Val Leu Arg Ala Leu Lys Ala Ile Ser Val Ile Ser Gly Leu Lys
 225 230 235 240
 Val Ile Val Gly Ala Leu Leu Arg Ser Val Lys Lys Leu Val Asp Val
 245 250 255
 Met Val Leu Thr Leu Phe Cys Leu Ser Ile Phe Ala Leu Val Gly Gln
 260 265 270
 Gln Leu Phe Met Gly Ile Leu Ser Gln Lys Cys Ile Lys Asp Asp Cys
 275 280 285
 Gly Pro Asn Ala Phe Ser Asn Lys Asp Cys Phe Val Lys Glu Asn Asp
 290 295 300
 Ser Glu Asp Phe Ile Met Cys Gly Asn Trp Leu Gly Arg Arg Ser Cys
 305 310 315 320
 Pro Asp Gly Ser Thr Cys Asn Lys Thr Thr Phe Asn Pro Asp Tyr Asn
 325 330 335
 Tyr Thr Asn Phe Asp Ser Phe Gly Trp Ser Phe Leu Ala Met Phe Arg
 340 345 350
 Val Met Thr Gln Asp Ser Trp Glu Lys Leu Tyr Arg Gln Ile Leu Arg
 355 360 365
 Thr Ser Gly Ile Tyr Phe Val Phe Phe Phe Val Val Val Ile Phe Leu
 370 375 380
 Gly Ser Phe Tyr Leu Leu Asn Leu Thr Leu Ala Val Val Thr Met Ala
 385 390 395 400
 Tyr Glu Glu Gln Asn Arg Asn Val Ala Ala Glu Thr Glu Ala Lys Glu
 405 410 415

Lys Met Phe Gln Glu Ala Gln Gln Leu Leu Arg Glu Glu Lys Glu Ala
 420 425 430
 Leu Val Ala Met Gly Ile Asp Arg Thr Ser Leu Asn Ser Leu Gln Ala
 435 440 445
 Ser Ser Phe Ser Pro Lys Lys Arg Lys Phe Phe Gly Ser Lys Thr Arg
 450 455 460
 Lys Ser Phe Phe Met Arg Gly Ser Lys Thr Ala Arg Ala Ser Ala Ser
 465 470 475 480
 Asp Ser Glu Asp Asp Ala Ser Lys Asn Pro Gln Leu Leu Glu Gln Thr
 485 490 495
 Lys Arg Leu Ser Gln Asn Leu Pro Val Glu Leu Phe Asp Glu His Val
 500 505 510
 Asp Pro Leu His Arg Gln Arg Ala Leu Ser Ala Val Ser Ile Leu Thr
 515 520 525
 Ile Thr Met Gln Glu Gln Glu Lys Ser Gln Glu Pro Cys Phe Pro Cys
 530 535 540
 Gly Lys Asn Leu Ala Ser Lys Tyr Leu Val Trp Glu Cys Ser Pro Pro
 545 550 555 560
 Trp Leu Cys Ile Lys Lys Val Leu Gln Thr Ile Met Thr Asp Pro Phe
 565 570 575
 Thr Glu Leu Ala Ile Thr Ile Cys Ile Ile Val Asn Thr Val Phe Leu
 580 585 590
 Ala Met Glu His His Asn Met Asp Asn Ser Leu Lys Asp Ile Leu Lys
 595 600 605
 Ile Gly Asn Trp Val Phe Thr Gly Ile Phe Ile Ala Glu Met Cys Leu
 610 615 620
 Lys Ile Ile Ala Leu Asp Pro Tyr His Tyr Phe Arg His Gly Trp Asn
 625 630 635 640
 Ile Phe Asp Ser Ile Val Ala Leu Val Ser Leu Ala Asp Val Leu Phe
 645 650 655
 His Lys Leu Ser Lys Asn Leu Ser Phe Leu Ala Ser Leu Arg Val Leu
 660 665 670
 Arg Val Phe Lys Leu Ala Lys Ser Trp Pro Thr Leu Asn Thr Leu Ile
 675 680 685
 Lys Ile Ile Gly His Ser Val Gly Ala Leu Gly Asn Leu Thr Val Val
 690 695 700
 Leu Thr Ile Val Val Phe Ile Phe Ser Val Val Gly Met Arg Leu Phe
 705 710 715 720

Gly Ala Lys Phe Asn Lys Thr Cys Ser Thr Ser Pro Glu Ser Leu Arg
 725 730 735
 Arg Trp His Met Gly Asp Phe Tyr His Ser Phe Leu Val Val Phe Arg
 740 745 750
 Ile Leu Cys Gly Glu Trp Ile Glu Asn Met Trp Glu Cys Met Gln Glu
 755 760 765
 Met Glu Gly Ser Pro Leu Cys Val Ile Val Phe Val Leu Ile Met Val
 770 775 780
 Val Gly Lys Leu Val Val Leu Asn Leu Phe Ile Ala Leu Leu Leu Asn
 785 790 795 800
 Ser Phe Ser Asn Glu Glu Lys Asp Gly Asn Pro Glu Gly Glu Thr Arg
 805 810 815
 Lys Thr Lys Val Gln Leu Ala Leu Asp Arg Phe Ser Arg Ala Phe Tyr
 820 825 830
 Phe Met Ala Arg Ala Leu Gln Asn Phe Cys Cys Lys Arg Cys Arg Arg
 835 840 845
 Gln Asn Ser Pro Lys Pro Asn Glu Ala Thr Glu Ser Phe Ala Gly Glu
 850 855 860
 Ser Arg Asp Thr Ala Thr Leu Asp Thr Arg Ser Trp Lys Glu Tyr Asp
 865 870 875 880
 Ser Glu Met Thr Leu Tyr Thr Gly Gln Ala Gly Ala Pro Leu Ala Pro
 885 890 895
 Leu Ala Lys Glu Glu Asp Asp Met Glu Cys Cys Gly Glu Cys Asp Ala
 900 905 910
 Ser Pro Thr Ser Gln Pro Ser Glu Glu Ala Gln Ala Cys Asp Leu Pro
 915 920 925
 Leu Lys Thr Lys Arg Leu Pro Ser Pro Asp Asp His Gly Val Glu Met
 930 935 940
 Glu Val Phe Ser Glu Glu Asp Pro Asn Leu Thr Ile Gln Ser Ala Arg
 945 950 955 960
 Lys Lys Ser Asp Ala Ala Ser Met Leu Ser Glu Cys Ser Thr Ile Asp
 965 970 975
 Leu Asn Asp Ile Phe Arg Asn Leu Gln Lys Thr Val Ser Pro Gln Lys
 980 985 990
 Gln Pro Asp Arg Cys Phe Pro Lys Gly Leu Ser Cys Ile Phe Leu Cys
 995 1000 1005
 Cys Lys Thr Ile Lys Lys Lys Ser Pro Trp Val Leu Trp Trp Asn Leu
 1010 1015 1020

Arg Lys Thr Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe
 1025 1030 1035 1040
 Ile Ile Phe Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp
 1045 1050 1055
 Val Asn Leu Pro Ser Arg Pro Gln Val Glu Lys Leu Leu Lys Cys Thr
 1060 1065 1070
 Asp Asn Ile Phe Thr Phe Ile Phe Leu Leu Glu Met Ile Leu Lys Trp
 1075 1080 1085
 Val Ala Phe Gly Phe Arg Lys Tyr Phe Thr Ser Ala Trp Cys Trp Leu
 1090 1095 1100
 Asp Phe Leu Ile Val Val Val Ser Val Leu Ser Leu Thr Asn Leu Pro
 1105 1110 1115 1120
 Asn Leu Lys Ser Phe Arg Asn Leu Arg Ala Leu Arg Pro Leu Arg Ala
 1125 1130 1135
 Leu Ser Gln Phe Glu Gly Met Lys Val Val Val Asn Ala Leu Met Ser
 1140 1145 1150
 Ala Ile Pro Ala Ile Leu Asn Val Leu Leu Val Cys Leu Ile Phe Trp
 1155 1160 1165
 Leu Ile Phe Cys Ile Leu Gly Val Asn Phe Phe Ser Gly Lys Phe Gly
 1170 1175 1180
 Arg Cys Ile Asn Gly Thr Asp Ile Asn Lys Tyr Phe Asn Ala Ser Asn
 1185 1190 1195 1200
 Val Pro Asn Gln Ser Gln Cys Leu Val Ser Asn Tyr Thr Trp Lys Val
 1205 1210 1215
 Pro Asn Val Asn Phe Asp Asn Val Gly Asn Ala Tyr Leu Ala Leu Leu
 1220 1225 1230
 Gln Val Ala Thr Tyr Lys Gly Trp Leu Asp Ile Met Asn Ala Ala Val
 1235 1240 1245
 Asp Ser Arg Gly Lys Asp Glu Gln Pro Ala Phe Glu Ala Asn Leu Tyr
 1250 1255 1260
 Ala Tyr Leu Tyr Phe Val Val Phe Ile Ile Phe Gly Ser Phe Phe Thr
 1265 1270 1275 1280
 Leu Asn Leu Phe Ile Gly Val Ile Ile Asp Asn Phe Asn Gln Gln Gln
 1285 1290 1295
 Lys Lys Leu Gly Gly Gln Asp Ile Phe Met Thr Glu Glu Gln Lys Lys
 1300 1305 1310
 Tyr Tyr Asn Ala Met Lys Lys Leu Gly Thr Lys Lys Pro Gln Lys Pro
 1315 1320 1325

Ile Pro Arg Pro Leu Asn Lys Cys Gln Ala Phe Val Phe Asp Leu Val
 1330 1335 1340
 Thr Ser Gln Val Phe Asp Val Ile Ile Leu Gly Leu Ile Val Thr Asn
 1345 1350 1355 1360
 Met Ile Ile Met Met Ala Glu Ser Glu Gly Gln Pro Asn Glu Val Lys
 1365 1370 1375
 Lys Ile Phe Asp Ile Leu Asn Ile Val Phe Val Val Ile Phe Thr Val
 1380 1385 1390
 Glu Cys Leu Ile Lys Val Phe Ala Leu Arg Gln His Tyr Phe Thr Asn
 1395 1400 1405
 Gly Trp Asn Leu Phe Asp Cys Val Val Val Val Leu Ser Ile Ile Ser
 1410 1415 1420
 Thr Leu Val Ser Gly Leu Glu Asn Ser Asn Val Phe Pro Pro Thr Leu
 1425 1430 1435 1440
 Phe Arg Ile Val Arg Leu Ala Arg Ile Gly Arg Ile Leu Arg Leu Val
 1445 1450 1455
 Arg Ala Ala Arg Gly Ile Arg Thr Leu Leu Phe Ala Leu Met Met Ser
 1460 1465 1470
 Leu Pro Ser Leu Phe Asn Ile Gly Leu Leu Leu Phe Leu Val Met Phe
 1475 1480 1485
 Ile Tyr Ala Ile Phe Gly Met Asn Trp Phe Ser Lys Val Lys Arg Gly
 1490 1495 1500
 Ser Gly Ile Asp Asp Ile Phe Asn Phe Asp Thr Phe Ser Gly Ser Met
 1505 1510 1515 1520
 Leu Cys Leu Phe Gln Ile Thr Thr Ser Ala Gly Trp Asp Ala Leu Leu
 1525 1530 1535
 Asn Pro Met Leu Glu Ser Lys Ala Ser Cys Asn Ser Ser Ser Gln Glu
 1540 1545 1550
 Ser Cys Gln Gln Pro Gln Ile Ala Ile Val Tyr Phe Val Ser Tyr Ile
 1555 1560 1565
 Ile Ile Ser Phe Leu Ile Val Val Asn Met Tyr Ile Ala Val Ile Leu
 1570 1575 1580
 Glu Asn Phe Asn Thr Ala Thr Glu Glu Ser Glu Asp Pro Leu Gly Glu
 1585 1590 1595 1600
 Asp Asp Phe Glu Ile Phe Tyr Glu Ile Trp Glu Lys Phe Asp Pro Glu
 1605 1610 1615
 Ala Thr Gln Phe Ile Gln Tyr Ser Ser Leu Ser Asp Phe Ala Asp Ala
 1620 1625 1630

Leu Pro Glu Pro Leu Arg Val Ala Lys Pro Asn Arg Phe Gln Phe Leu
 1635 1640 1645

Met Met Asp Leu Pro Met Val Met Gly Asp Arg Leu His Cys Met Asp
 1650 1655 1660

Val Leu Phe Ala Phe Thr Thr Arg Val Leu Gly Asn Ser Ser Gly Leu
 1665 1670 1675 1680

Asp Thr Met Lys Ala Met Met Glu Glu Lys Phe Met Glu Ala Asn Pro
 1685 1690 1695

Phe Lys Lys Leu Tyr Glu Pro Ile Val Thr Thr Thr Lys Arg Lys Glu
 1700 1705 1710

Glu Glu Glu Cys Ala Ala Val Ile Gln Arg Ala Tyr Arg Arg His Met
 1715 1720 1725

Glu Lys Met Ile Lys Leu Lys Leu Lys Gly Arg Ser Ser Ser Ser Leu
 1730 1735 1740

Gln Val Phe Cys Asn Gly Asp Leu Ser Ser Leu Asp Val Pro Lys Ile
 1745 1750 1755 1760

Lys Val His Cys Asp
 1765

<210> 6

<211> 3701

<212> DNA

<213> Homo sapiens

<220>

<221> CDS

<222> (1)..(3699)

<223> partial human NaN cDNA sequence

<220>

<221> unsure

<222> (922)

<223> y = c or t

<400> 6

tcc att gtc att gga ata gcg att gtg tca tat att cca gga atc acc 48
 Ser Ile Val Ile Gly Ile Ala Ile Val Ser Tyr Ile Pro Gly Ile Thr
 1 5 10 15

atc aaa cta ttg ccc ctg cgt acc ttc cgt gtg ttc aga gct ttg aaa 96
 Ile Lys Leu Leu Pro Leu Arg Thr Phe Arg Val Phe Arg Ala Leu Lys
 20 25 30

gca att tca gta gtt tca cgt ctg aag gtc atc gtg ggg gcc ttg cta 144
 Ala Ile Ser Val Val Ser Arg Leu Lys Val Ile Val Gly Ala Leu Leu
 35 40 45

cgc tct gtg aag aag ctg gtc aac gtg att atc ctc acc ttc ttt tgc	192
Arg Ser Val Lys Lys Leu Val Asn Val Ile Ile Leu Thr Phe Phe Cys	
50 55 60	
ctc agc atc ttt gcc ctg gta ggt cag cag ctc ttc atg gga agt ctg	240
Leu Ser Ile Phe Ala Leu Val Gly Gln Gln Leu Phe Met Gly Ser Leu	
65 70 75 80	
aac ctg aaa tgc atc tcg agg gac tgt aaa aat atc agt aac ccg gaa	288
Asn Leu Lys Cys Ile Ser Arg Asp Cys Lys Asn Ile Ser Asn Pro Glu	
85 90 95	
gct tat gac cat tgc ttt gaa aag aaa gaa aat tca cct gaa ttc aaa	336
Ala Tyr Asp His Cys Phe Glu Lys Lys Glu Asn Ser Pro Glu Phe Lys	
100 105 110	
atg tgt ggc atc tgg atg ggt aac agt gcc tgt tcc ata caa tat gaa	384
Met Cys Gly Ile Trp Met Gly Asn Ser Ala Cys Ser Ile Gln Tyr Glu	
115 120 125	
tgt aag cac acc aaa att aat cct gac tat aat tat acg aat ttt gac	432
Cys Lys His Thr Lys Ile Asn Pro Asp Tyr Asn Tyr Thr Asn Phe Asp	
130 135 140	
aac ttt ggc tgg tct ttt ctt gcc atg ttc cgg ctg atg acc caa gat	480
Asn Phe Gly Trp Ser Phe Leu Ala Met Phe Arg Leu Met Thr Gln Asp	
145 150 155 160	
tcc tgg gag aag ctt tat caa cag acc ctg cgt act act ggg ctc tac	528
Ser Trp Glu Lys Leu Tyr Gln Gln Thr Leu Arg Thr Thr Gly Leu Tyr	
165 170 175	
tca gtc ttc ttc ttc att gtg gtc att ttc ctg ggc tcc ttc tac ctg	576
Ser Val Phe Phe Phe Ile Val Val Ile Phe Leu Gly Ser Phe Tyr Leu	
180 185 190	
att aac tta acc ctg gct gtt gtt acc atg gca tat gag gag cag aac	624
Ile Asn Leu Thr Leu Ala Val Val Thr Met Ala Tyr Glu Glu Gln Asn	
195 200 205	
aag aat gta gct gca gag ata gag gcc aag gaa aag atg ttt cag gaa	672
Lys Asn Val Ala Ala Glu Ile Glu Ala Lys Glu Lys Met Phe Gln Glu	
210 215 220	
gcc cag cag ctg tta aag gag gaa aag gag gct ctg gtt gcc atg gga	720
Ala Gln Gln Leu Leu Lys Glu Glu Lys Glu Ala Leu Val Ala Met Gly	
225 230 235 240	
att gac aga agt tca ctt act tcc ctt gaa aca tca tat ttt acc cca	768
Ile Asp Arg Ser Ser Leu Thr Ser Leu Glu Thr Ser Tyr Phe Thr Pro	
245 250 255	
aaa aag aga aag ctc ttt ggt aat aag aaa agg aag tcc ttc ttt ttg	816
Lys Lys Arg Lys Leu Phe Gly Asn Lys Lys Arg Lys Ser Phe Phe Leu	
260 265 270	

aga gag tct ggg aaa gac cag cct cct ggg tca gat tct gat gaa gat	864
Arg Glu Ser Gly Lys Asp Gln Pro Pro Gly Ser Asp Ser Asp Glu Asp	
275 280 285	
tgc caa aaa aag cca cag ctc cta gag caa acc aaa cga ctg tcc cag	912
Cys Gln Lys Lys Pro Gln Leu Leu Glu Gln Thr Lys Arg Leu Ser Gln	
290 295 300	
aat cta tca ytg gac cac ttt gat gag cat gga gat cct ctc caa agg	960
Asn Leu Ser Xaa Asp His Phe Asp Glu His Gly Asp Pro Leu Gln Arg	
305 310 315 320	
cag aga gca ctg agt gct gtc agc atc ctc acc atc acc atg aag gaa	1008
Gln Arg Ala Leu Ser Ala Val Ser Ile Leu Thr Ile Thr Met Lys Glu	
325 330 335	
caa gaa aaa tca caa gag cct tgt ctc cct tgt gga gaa aac ctg gca	1056
Gln Glu Lys Ser Gln Glu Pro Cys Leu Pro Cys Gly Glu Asn Leu Ala	
340 345 350	
tcc aag tac ctc gtg tgg aac tgt tgc ccc cag tgg ctg tgc gtt aag	1104
Ser Lys Tyr Leu Val Trp Asn Cys Cys Pro Gln Trp Leu Cys Val Lys	
355 360 365	
aag gtc ctg aga act gtg atg act gac ccg ttt act gag ctg gcc atc	1152
Lys Val Leu Arg Thr Val Met Thr Asp Pro Phe Thr Glu Leu Ala Ile	
370 375 380	
acc atc tgc atc atc atc aac act gtc ttc ttg gcc atg gag cat cac	1200
Thr Ile Cys Ile Ile Ile Asn Thr Val Phe Leu Ala Met Glu His His	
385 390 395 400	
aag atg gag gcc agt ttt gag aag atg ttg aat ata ggg aat ttg gtt	1248
Lys Met Glu Ala Ser Phe Glu Lys Met Leu Asn Ile Gly Asn Leu Val	
405 410 415	
ttc act agc att ttt ata gca gaa atg tgc cta aaa atc att gcg ctc	1296
Phe Thr Ser Ile Phe Ile Ala Glu Met Cys Leu Lys Ile Ile Ala Leu	
420 425 430	
gat ccc tac cac tac ttt cgc cga ggc tgg aac att ttt gac agc att	1344
Asp Pro Tyr His Tyr Phe Arg Arg Gly Trp Asn Ile Phe Asp Ser Ile	
435 440 445	
gtt gct ctt ctg agt ttt gca gat gta atg aac tgt gta ctt caa aag	1392
Val Ala Leu Leu Ser Phe Ala Asp Val Met Asn Cys Val Leu Gln Lys	
450 455 460	
aga agc tgg cca ttc ttg cgt tcc ttc aga gtg ctc agg gtc ttc aag	1440
Arg Ser Trp Pro Phe Leu Arg Ser Phe Arg Val Leu Arg Val Phe Lys	
465 470 475 480	
tta gcc aaa tcc tgg cca act ttg aac aca cta att aag ata atc ggc	1488
Leu Ala Lys Ser Trp Pro Thr Leu Asn Thr Leu Ile Lys Ile Ile Gly	
485 490 495	

aac tct gtc gga gcc ctt gga agc ctg act gtg gtc ctg gtc att gtg	1536
Asn Ser Val Gly Ala Leu Gly Ser Leu Thr Val Val Leu Val Ile Val	
500 505 510	
atc ttt att ttc tca gta gtt ggc atg cag ctt ttt ggc cgt agc ttc	1584
Ile Phe Ile Phe Ser Val Val Gly Met Gln Leu Phe Gly Arg Ser Phe	
515 520 525	
aat tcc caa aag agt cca aaa ctc tgt aac ccg aca ggc ccg aca gtc	1632
Asn Ser Gln Lys Ser Pro Lys Leu Cys Asn Pro Thr Gly Pro Thr Val	
530 535 540	
tca tgt tta cgg cac tgg cac atg ggg gat ttc tgg cac tcc ttc cta	1680
Ser Cys Leu Arg His Trp His Met Gly Asp Phe Trp His Ser Phe Leu	
545 550 555 560	
gtg gta ttc cgc atc ctc tgc ggg gaa tgg atc gaa aat atg tgg gaa	1728
Val Val Phe Arg Ile Leu Cys Gly Glu Trp Ile Glu Asn Met Trp Glu	
565 570 575	
tgt atg caa gaa gcg aat gca tca tca tca ttg tgt gtt att gtc ttc	1776
Cys Met Gln Glu Ala Asn Ala Ser Ser Ser Leu Cys Val Ile Val Phe	
580 585 590	
ata ttg atc acg gtg ata gga aaa ctt gtg gtg ctc aac ctc ttc att	1824
Ile Leu Ile Thr Val Ile Gly Lys Leu Val Val Leu Asn Leu Phe Ile	
595 600 605	
gcc tta ctg ctc aat tcc ttt agc aat gag gaa aga aat gga aac tta	1872
Ala Leu Leu Leu Asn Ser Phe Ser Asn Glu Glu Arg Asn Gly Asn Leu	
610 615 620	
gaa gga gag gcc agg aaa act aaa gtc cag tta gca ctg gat cga ttc	1920
Glu Gly Glu Ala Arg Lys Thr Lys Val Gln Leu Ala Leu Asp Arg Phe	
625 630 635 640	
cgc cgg gct ttt tgt ttt gtg aga cac act ctt gag cat ttc tgt cac	1968
Arg Arg Ala Phe Cys Phe Val Arg His Thr Leu Glu His Phe Cys His	
645 650 655	
aag tgg tgc agg aag caa aac tta cca cag caa aaa gag gtg gca gga	2016
Lys Trp Cys Arg Lys Gln Asn Leu Pro Gln Gln Lys Glu Val Ala Gly	
660 665 670	
ggc tgt gct gca caa agc aaa gac atc att ccc ctg gtc atg gag atg	2064
Gly Cys Ala Ala Gln Ser Lys Asp Ile Ile Pro Leu Val Met Glu Met	
675 680 685	
aaa agg ggc tca gag acc cag gag gag ctt ggt ata cta acc tct gta	2112
Lys Arg Gly Ser Glu Thr Gln Glu Glu Leu Gly Ile Leu Thr Ser Val	
690 695 700	
cca aag acc ctg ggc gtc agg cat gat tgg act tgg ttg gca cca ctt	2160
Pro Lys Thr Leu Gly Val Arg His Asp Trp Thr Trp Leu Ala Pro Leu	
705 710 715 720	

gcg gag gag gaa gat gac gtt gaa ttt tct ggt gaa gat aat gca cag	2208
Ala Glu Glu Glu Asp Asp Val Glu Phe Ser Gly Glu Asp Asn Ala Gln	
725 730 735	
cgc atc aca caa cct gag cct gaa caa cag gcc tat gag ctc cat cag	2256
Arg Ile Thr Gln Pro Glu Pro Glu Gln Gln Ala Tyr Glu Leu His Gln	
740 745 750	
gag aac aag aag ccc acg agc cag aga gtt caa agt gtg gaa att gac	2304
Glu Asn Lys Lys Pro Thr Ser Gln Arg Val Gln Ser Val Glu Ile Asp	
755 760 765	
atg ttc tct gaa gat gag cct cat ctg acc ata cag gat ccc cga aag	2352
Met Phe Ser Glu Asp Glu Pro His Leu Thr Ile Gln Asp Pro Arg Lys	
770 775 780	
aag tct gat gtt acc agt ata cta tca gaa tgt agc acc att gat ctt	2400
Lys Ser Asp Val Thr Ser Ile Leu Ser Glu Cys Ser Thr Ile Asp Leu	
785 790 795 800	
cag gat ggc ttt gga tgg tta cct gag atg gtt ccc aaa aag caa cca	2448
Gln Asp Gly Phe Gly Trp Leu Pro Glu Met Val Pro Lys Lys Gln Pro	
805 810 815	
gag aga tgt ttg ccc aaa ggc ttt ggt tgc tgc ttt cca tgc tgt agc	2496
Glu Arg Cys Leu Pro Lys Gly Phe Gly Cys Cys Phe Pro Cys Cys Ser	
820 825 830	
gtg gac aag aga aag cct ccc tgg gtc att tgg tgg aac ctg cgg aaa	2544
Val Asp Lys Arg Lys Pro Pro Trp Val Ile Trp Trp Asn Leu Arg Lys	
835 840 845	
acc tgc tac caa ata gtg aaa cac agc tgg ttt gag agc ttt att atc	2592
Thr Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile Ile	
850 855 860	
ttt gtg att ctg ctg agc agt ggg gca ctg ata ttt gaa gat gtt cac	2640
Phe Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val His	
865 870 875 880	
ctt gag aac caa ccc aaa atc caa gaa tta cta aat tgt act gac att	2688
Leu Glu Asn Gln Pro Lys Ile Gln Glu Leu Leu Asn Cys Thr Asp Ile	
885 890 895	
att ttt aca cat att ttt atc ctg gag atg gta cta aaa tgg gta gcc	2736
Ile Phe Thr His Ile Phe Ile Leu Glu Met Val Leu Lys Trp Val Ala	
900 905 910	
ttc gga ttt gga aag tat ttc acc agt gcc tgg tgc tgc ctt gat ttc	2784
Phe Gly Phe Gly Lys Tyr Phe Thr Ser Ala Trp Cys Cys Leu Asp Phe	
915 920 925	
atc att gtg att gtc tct gtg acc acc ctc att aac tta atg gaa ttg	2832
Ile Ile Val Ile Val Ser Val Thr Thr Leu Ile Asn Leu Met Glu Leu	
930 935 940	

41

ttt gac atc atc atc ata agt ctc att atc cta aac atg att agc atg 3552
 Phe Asp Ile Ile Il Ile Ser Leu Ile Ile Leu Asn Met Ile Ser Met
 1170 1175 1180

atg gct gaa tca tac aac caa ccc aaa gcc atg aaa tcc atc ctt gac 3600
 Met Ala Glu Ser Tyr Asn Gln Pro Lys Ala Met Lys Ser Ile Leu Asp
 1185 1190 1195 1200

cat ctc aac tgg gtc ttt gtg gtc atc ttt acg tta gaa tgt ctc atc 3648
 His Leu Asn Trp Val Phe Val Val Ile Phe Thr Leu Glu Cys Leu Ile
 1205 1210 1215

aaa atc ttt gct ttg agg caa tac tac ttc acc aat ggc tgg aat tta 3696
 Lys Ile Phe Ala Leu Arg Gln Tyr Phe Thr Asn Gly Trp Asn Leu
 1220 1225 1230

ttt ga 3701
 Phe

<210> 7

<211> 1233

<212> PRT

<213> Homo sapiens

<400> 7

Ser Ile Val Ile Gly Ile Ala Ile Val Ser Tyr Ile Pro Gly Ile Thr
 1 5 10 15

Ile Lys Leu Leu Pro Leu Arg Thr Phe Arg Val Phe Arg Ala Leu Lys
 20 25 30

Ala Ile Ser Val Val Ser Arg Leu Lys Val Ile Val Gly Ala Leu Leu
 35 40 45

Arg Ser Val Lys Lys Leu Val Asn Val Ile Ile Leu Thr Phe Phe Cys
 50 55 60

Leu Ser Ile Phe Ala Leu Val Gly Gln Gln Leu Phe Met Gly Ser Leu
 65 70 75 80

Asn Leu Lys Cys Ile Ser Arg Asp Cys Lys Asn Ile Ser Asn Pro Glu
 85 90 95

Ala Tyr Asp His Cys Phe Glu Lys Lys Glu Asn Ser Pro Glu Phe Lys
 100 105 110

Met Cys Gly Ile Trp Met Gly Asn Ser Ala Cys Ser Ile Gln Tyr Glu
 115 120 125

Cys Lys His Thr Lys Ile Asn Pro Asp Tyr Asn Tyr Thr Asn Phe Asp
 130 135 140

Asn Phe Gly Trp Ser Phe Leu Ala Met Phe Arg Leu Met Thr Gln Asp
 145 150 155 160

Ser Trp Glu Lys Leu Tyr Gln Gln Thr Leu Arg Thr Thr Gly Leu Tyr
 165 170 175
 Ser Val Phe Phe Phe Ile Val Val Ile Phe Leu Gly Ser Phe Tyr Leu
 180 185 190
 Ile Asn Leu Thr Leu Ala Val Val Thr Met Ala Tyr Glu Glu Gln Asn
 195 200 205
 Lys Asn Val Ala Ala Glu Ile Glu Ala Lys Glu Lys Met Phe Gln Glu
 210 215 220
 Ala Gln Gln Leu Leu Lys Glu Glu Lys Glu Ala Leu Val Ala Met Gly
 225 230 235 240
 Ile Asp Arg Ser Ser Leu Thr Ser Leu Glu Thr Ser Tyr Phe Thr Pro
 245 250 255
 Lys Lys Arg Lys Leu Phe Gly Asn Lys Lys Arg Lys Ser Phe Phe Leu
 260 265 270
 Arg Glu Ser Gly Lys Asp Gln Pro Pro Gly Ser Asp Ser Asp Glu Asp
 275 280 285
 Cys Gln Lys Lys Pro Gln Leu Leu Glu Gln Thr Lys Arg Leu Ser Gln
 290 295 300
 Asn Leu Ser Xaa Asp His Phe Asp Glu His Gly Asp Pro Leu Gln Arg
 305 310 315 320
 Gln Arg Ala Leu Ser Ala Val Ser Ile Leu Thr Ile Thr Met Lys Glu
 325 330 335
 Gln Glu Lys Ser Gln Glu Pro Cys Leu Pro Cys Gly Glu Asn Leu Ala
 340 345 350
 Ser Lys Tyr Leu Val Trp Asn Cys Cys Pro Gln Trp Leu Cys Val Lys
 355 360 365
 Lys Val Leu Arg Thr Val Met Thr Asp Pro Phe Thr Glu Leu Ala Ile
 370 375 380
 Thr Ile Cys Ile Ile Ile Asn Thr Val Phe Leu Ala Met Glu His His
 385 390 395 400
 Lys Met Glu Ala Ser Phe Glu Lys Met Leu Asn Ile Gly Asn Leu Val
 405 410 415
 Phe Thr Ser Ile Phe Ile Ala Glu Met Cys Leu Lys Ile Ile Ala Leu
 420 425 430
 Asp Pro Tyr His Tyr Phe Arg Arg Gly Trp Asn Ile Phe Asp Ser Ile
 435 440 445
 Val Ala Leu Leu Ser Phe Ala Asp Val Met Asn Cys Val Leu Gln Lys
 450 455 460

Arg Ser Trp Pro Phe Leu Arg Ser Phe Arg Val Leu Arg Val Phe Lys
 465 470 475 480
 Leu Ala Lys Ser Trp Pro Thr Leu Asn Thr Leu Ile Lys Ile Ile Gly
 485 490 495
 Asn Ser Val Gly Ala Leu Gly Ser Leu Thr Val Val Leu Val Ile Val
 500 505 510
 Ile Phe Ile Phe Ser Val Val Gly Met Gln Leu Phe Gly Arg Ser Phe
 515 520 525
 Asn Ser Gln Lys Ser Pro Lys Leu Cys Asn Pro Thr Gly Pro Thr Val
 530 535 540
 Ser Cys Leu Arg His Trp His Met Gly Asp Phe Trp His Ser Phe Leu
 545 550 555 560
 Val Val Phe Arg Ile Leu Cys Gly Glu Trp Ile Glu Asn Met Trp Glu
 565 570 575
 Cys Met Gln Glu Ala Asn Ala Ser Ser Ser Leu Cys Val Ile Val Phe
 580 585 590
 Ile Leu Ile Thr Val Ile Gly Lys Leu Val Val Leu Asn Leu Phe Ile
 595 600 605
 Ala Leu Leu Leu Asn Ser Phe Ser Asn Glu Glu Arg Asn Gly Asn Leu
 610 615 620
 Glu Gly Glu Ala Arg Lys Thr Lys Val Gln Leu Ala Leu Asp Arg Phe
 625 630 635 640
 Arg Arg Ala Phe Cys Phe Val Arg His Thr Leu Glu His Phe Cys His
 645 650 655
 Lys Trp Cys Arg Lys Gln Asn Leu Pro Gln Gln Lys Glu Val Ala Gly
 660 665 670
 Gly Cys Ala Ala Gln Ser Lys Asp Ile Ile Pro Leu Val Met Glu Met
 675 680 685
 Lys Arg Gly Ser Glu Thr Gln Glu Glu Leu Gly Ile Leu Thr Ser Val
 690 695 700
 Pro Lys Thr Leu Gly Val Arg His Asp Trp Thr Trp Leu Ala Pro Leu
 705 710 715 720
 Ala Glu Glu Glu Asp Asp Val Glu Phe Ser Gly Glu Asp Asn Ala Gln
 725 730 735
 Arg Ile Thr Gln Pro Glu Pro Glu Gln Gln Ala Tyr Glu Leu His Gln
 740 745 750
 Glu Asn Lys Lys Pro Thr Ser Gln Arg Val Gln Ser Val Glu Ile Asp
 755 760 765

Met Phe Ser Glu Asp Glu Pro His Leu Thr Ile Gln Asp Pro Arg Lys
 770 775 780
 Lys Ser Asp Val Thr Ser Ile Leu Ser Glu Cys Ser Thr Ile Asp Leu
 785 790 795 800
 Gln Asp Gly Phe Gly Trp Leu Pro Glu Met Val Pro Lys Lys Gln Pro
 805 810 815
 Glu Arg Cys Leu Pro Lys Gly Phe Gly Cys Cys Phe Pro Cys Cys Ser
 820 825 830
 Val Asp Lys Arg Lys Pro Pro Trp Val Ile Trp Trp Asn Leu Arg Lys
 835 840 845
 Thr Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile Ile
 850 855 860
 Phe Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val His
 865 870 875 880
 Leu Glu Asn Gln Pro Lys Ile Gln Glu Leu Leu Asn Cys Thr Asp Ile
 885 890 895
 Ile Phe Thr His Ile Phe Ile Leu Glu Met Val Leu Lys Trp Val Ala
 900 905 910
 Phe Gly Phe Gly Lys Tyr Phe Thr Ser Ala Trp Cys Cys Leu Asp Phe
 915 920 925
 Ile Ile Val Ile Val Ser Val Thr Thr Leu Ile Asn Leu Met Glu Leu
 930 935 940
 Lys Ser Phe Arg Thr Leu Arg Ala Leu Arg Pro Leu Arg Ala Leu Ser
 945 950 955 960
 Gln Phe Glu Gly Met Lys Val Val Val Asn Ala Leu Ile Gly Ala Ile
 965 970 975
 Pro Ala Ile Leu Asn Val Leu Leu Val Cys Leu Ile Phe Trp Leu Val
 980 985 990
 Phe Cys Ile Leu Gly Val Tyr Phe Phe Ser Gly Lys Phe Gly Lys Cys
 995 1000 1005
 Ile Asn Gly Thr Asp Ser Val Ile Asn Tyr Thr Ile Ile Thr Asn Lys
 1010 1015 1020
 Ser Gln Cys Glu Ser Gly Asn Phe Ser Trp Ile Asn Gln Lys Val Asn
 1025 1030 1035 1040
 Phe Asp Asn Val Gly Asn Ala Tyr Leu Ala Leu Leu Gln Val Ala Thr
 1045 1050 1055
 Phe Lys Gly Trp Met Asp Ile Ile Tyr Ala Ala Val Asp Ser Thr Glu
 1060 1065 1070

Lys Glu Gln Gln Pro Glu Phe Glu Ser Asn Ser Leu Gly Tyr Ile Tyr
 1075 1080 1085

 Phe Val Val Phe Ile Ile Phe Gly Ser Phe Phe Thr Leu Asn Leu Phe
 1090 1095 1100

 Ile Gly Val Ile Ile Asp Asn Phe Asn Gln Gln Gln Lys Lys Leu Gly
 1105 1110 1115 1120

 Gly Gln Asp Ile Phe Met Thr Glu Glu Gln Lys Lys Tyr Tyr Asn Ala
 1125 1130 1135

 Met Lys Lys Leu Gly Ser Lys Lys Pro Gln Lys Pro Ile Pro Arg Pro
 1140 1145 1150

 Leu Asn Lys Cys Gln Gly Leu Val Phe Asp Ile Val Thr Ser Gln Ile
 1155 1160 1165

 Phe Asp Ile Ile Ile Ile Ser Leu Ile Ile Leu Asn Met Ile Ser Met
 1170 1175 1180

 Met Ala Glu Ser Tyr Asn Gln Pro Lys Ala Met Lys Ser Ile Leu Asp
 1185 1190 1195 1200

 His Leu Asn Trp Val Phe Val Val Ile Phe Thr Leu Glu Cys Leu Ile
 1205 1210 1215

 Lys Ile Phe Ala Leu Arg Gln Tyr Tyr Phe Thr Asn Gly Trp Asn Leu
 1220 1225 1230

 Phe

<210> 8

<211> 1243

<212> PRT

<213> Homo sapiens

<220>

<223> partial human NaN amino acid seq.

<400> 8

Ser Ile Val Ile Gly Ile Ala Ile Val Ser Tyr Ile Pro Gly Ile Thr
 1 5 10 15

 Ile Lys Leu Leu Pro Leu Arg Thr Phe Arg Val Phe Arg Ala Leu Lys
 20 25 30

 Ala Ile Ser Val Val Ser Arg Leu Lys Val Ile Val Gly Ala Leu Leu
 35 40 45

 Arg Ser Val Lys Lys Leu Val Asn Val Ile Ile Leu Thr Phe Phe Cys
 50 55 60

Leu Ser Ile Phe Ala Leu Val Gly Gln Gln Leu Phe Met Gly Ser Leu
 65 70 75 80
 Asn Leu Lys Cys Ile Ser Arg Asp Cys Lys Asn Ile Ser Asn Pro Glu
 85 90 95
 Ala Tyr Asp His Cys Phe Glu Lys Lys Glu Asn Ser Pro Glu Phe Lys
 100 105 110
 Met Cys Gly Ile Trp Met Gly Asn Ser Ala Cys Ser Ile Gln Tyr Glu
 115 120 125
 Cys Lys His Thr Lys Ile Asn Pro Asp Tyr Asn Tyr Thr Asn Phe Asp
 130 135 140
 Asn Phe Gly Trp Ser Phe Leu Ala Met Phe Arg Leu Met Thr Gln Asp
 145 150 155 160
 Ser Trp Glu Lys Leu Tyr Gln Gln Thr Leu Arg Thr Thr Gly Leu Tyr
 165 170 175
 Ser Val Phe Phe Phe Ile Val Val Ile Phe Leu Gly Ser Phe Tyr Leu
 180 185 190
 Ile Asn Leu Thr Leu Ala Val Val Thr Met Ala Tyr Glu Glu Gln Asn
 195 200 205
 Lys Asn Val Ala Ala Glu Ile Glu Ala Lys Glu Lys Met Phe Gln Glu
 210 215 220
 Ala Gln Gln Leu Leu Lys Glu Glu Lys Glu Ala Leu Val Ala Met Gly
 225 230 235 240
 Ile Asp Arg Ser Ser Leu Thr Ser Leu Glu Thr Ser Tyr Phe Thr Pro
 245 250 255
 Lys Lys Arg Lys Leu Phe Gly Asn Lys Lys Arg Lys Ser Phe Phe Leu
 260 265 270
 Arg Glu Ser Gly Lys Asp Gln Pro Pro Gly Ser Asp Ser Asp Glu Asp
 275 280 285
 Cys Gln Lys Lys Pro Gln Leu Leu Glu Gln Thr Lys Arg Leu Ser Gln
 290 295 300
 Asn Leu Ser Leu Asp His Phe Asp Glu His Gly Asp Pro Leu Gln Arg
 305 310 315 320
 Gln Arg Ala Leu Ser Ala Val Ser Ile Leu Thr Ile Thr Met Lys Glu
 325 330 335
 Gln Glu Lys Ser Gln Glu Pro Cys Leu Pro Cys Gly Glu Asn Leu Ala
 340 345 350
 Ser Lys Tyr Leu Val Trp Asn Cys Cys Pro Gln Trp Leu Cys Val Lys
 355 360 365

Lys Val Leu Arg Thr Val Met Thr Asp Pro Phe Thr Glu Leu Ala Ile
 370 375 380
 Thr Ile Cys Ile Ile Ile Asn Thr Val Phe Leu Ala Met Glu His His
 385 390 395 400
 Lys Met Glu Ala Ser Phe Glu Lys Met Leu Asn Ile Gly Asn Leu Val
 405 410 415
 Phe Thr Ser Ile Phe Ile Ala Glu Met Cys Leu Lys Ile Ile Ala Leu
 420 425 430
 Asp Pro Tyr His Tyr Phe Arg Arg Gly Trp Asn Ile Phe Asp Ser Ile
 435 440 445
 Val Ala Leu Leu Ser Phe Ala Asp Val Met Asn Cys Val Leu Gln Lys
 450 455 460
 Arg Ser Trp Pro Phe Leu Arg Ser Phe Arg Val Leu Arg Val Phe Lys
 465 470 475 480
 Leu Ala Lys Ser Trp Pro Thr Leu Asn Thr Leu Ile Lys Ile Ile Gly
 485 490 495
 Asn Ser Val Gly Ala Leu Gly Ser Leu Thr Val Val Leu Val Ile Val
 500 505 510
 Ile Phe Ile Phe Ser Val Val Gly Met Gln Leu Phe Gly Arg Ser Phe
 515 520 525
 Asn Ser Gln Lys Ser Pro Lys Leu Cys Asn Pro Thr Gly Pro Thr Val
 530 535 540
 Ser Cys Leu Arg His Trp His Met Gly Asp Phe Trp His Ser Phe Leu
 545 550 555 560
 Val Val Phe Arg Ile Leu Cys Gly Glu Trp Ile Glu Asn Met Trp Glu
 565 570 575
 Cys Met Gln Glu Ala Asn Ala Ser Ser Ser Leu Cys Val Ile Val Phe
 580 585 590
 Ile Leu Ile Thr Val Ile Gly Lys Leu Val Val Leu Asn Leu Phe Ile
 595 600 605
 Ala Leu Leu Leu Asn Ser Phe Ser Asn Glu Glu Arg Asn Gly Asn Leu
 610 615 620
 Glu Gly Glu Ala Arg Lys Thr Lys Val Gln Leu Ala Leu Asp Arg Phe
 625 630 635 640
 Arg Arg Ala Phe Cys Phe Val Arg His Thr Leu Glu His Phe Cys His
 645 650 655
 Lys Trp Cys Arg Lys Gln Asn Leu Pro Gln Gln Lys Glu Val Ala Gly
 660 665 670

Gly Cys Ala Ala Gln Ser Lys Asp Ile Ile Pro Leu Val Met Glu Met
 675 680 685
 Lys Arg Gly Ser Glu Thr Gln Glu Glu Leu Gly Ile Leu Thr Ser Val
 690 695 700
 Pro Lys Thr Leu Gly Val Arg His Asp Trp Thr Trp Leu Ala Pro Leu
 705 710 715 720
 Ala Glu Glu Glu Asp Asp Val Glu Phe Ser Gly Glu Asp Asn Ala Gln
 725 730 735
 Arg Ile Thr Gln Pro Glu Pro Glu Gln Gln Ala Tyr Glu Leu His Gln
 740 745 750
 Glu Asn Lys Lys Pro Thr Ser Gln Arg Val Gln Ser Val Glu Ile Asp
 755 760 765
 Met Phe Ser Glu Asp Glu Pro His Leu Thr Ile Gln Asp Pro Arg Lys
 770 775 780
 Lys Ser Asp Val Thr Ser Ile Leu Ser Glu Cys Ser Thr Ile Asp Leu
 785 790 795 800
 Gln Asp Gly Phe Gly Trp Leu Pro Glu Met Val Pro Lys Lys Gln Pro
 805 810 815
 Glu Arg Cys Leu Pro Lys Gly Phe Gly Cys Cys Phe Pro Cys Cys Ser
 820 825 830
 Val Asp Lys Arg Lys Pro Pro Trp Val Ile Trp Trp Asn Leu Arg Lys
 835 840 845
 Thr Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile Ile
 850 855 860
 Phe Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val His
 865 870 875 880
 Leu Glu Asn Gln Pro Lys Ile Gln Glu Leu Leu Asn Cys Thr Asp Ile
 885 890 895
 Ile Phe Thr His Ile Phe Ile Leu Glu Met Val Leu Lys Trp Val Ala
 900 905 910
 Phe Gly Phe Gly Lys Tyr Phe Thr Ser Ala Trp Cys Cys Leu Asp Phe
 915 920 925
 Ile Ile Val Ile Val Ser Val Thr Thr Leu Ile Asn Leu Met Glu Leu
 930 935 940
 Lys Ser Phe Arg Thr Leu Arg Ala Leu Arg Pro Leu Arg Ala Leu Ser
 945 950 955 960
 Gln Phe Glu Gly Met Lys Val Val Val Asn Ala Leu Ile Gly Ala Ile
 965 970 975

Pro Ala Ile Leu Asn Val Leu Leu Val Cys Leu Ile Phe Trp Leu Val
 980 985 990
 Phe Cys Ile Leu Gly Val Tyr Phe Phe Ser Gly Lys Phe Gly Lys Cys
 995 1000 1005
 Ile Asn Gly Thr Asp Ser Val Ile Asn Tyr Thr Ile Ile Thr Asn Lys
 1010 1015 1020
 Ser Gln Cys Glu Ser Gly Asn Phe Ser Trp Ile Asn Gln Lys Val Asn
 1025 1030 1035 1040
 Phe Asp Asn Val Gly Asn Ala Tyr Leu Ala Leu Leu Gln Val Ala Thr
 1045 1050 1055
 Phe Lys Gly Trp Met Asp Ile Ile Tyr Ala Ala Val Asp Ser Thr Glu
 1060 1065 1070
 Lys Glu Gln Gln Pro Glu Phe Glu Ser Asn Ser Leu Gly Tyr Ile Tyr
 1075 1080 1085
 Phe Val Val Phe Ile Ile Phe Gly Ser Phe Phe Thr Leu Asn Leu Phe
 1090 1095 1100
 Ile Gly Val Ile Ile Asp Asn Phe Asn Gln Gln Gln Lys Lys Leu Gly
 1105 1110 1115 1120
 Gly Gln Asp Ile Phe Met Thr Glu Glu Gln Lys Lys Tyr Tyr Asn Ala
 1125 1130 1135
 Met Lys Lys Leu Gly Ser Lys Lys Pro Gln Lys Pro Ile Pro Arg Pro
 1140 1145 1150
 Leu Asn Lys Cys Gln Gly Leu Val Phe Asp Ile Val Thr Ser Gln Ile
 1155 1160 1165
 Phe Asp Ile Ile Ile Ile Ser Leu Ile Ile Leu Asn Met Ile Ser Met
 1170 1175 1180
 Met Ala Glu Ser Tyr Asn Gln Pro Lys Ala Met Lys Ser Ile Leu Asp
 1185 1190 1195 1200
 His Leu Asn Trp Val Phe Val Val Ile Phe Thr Leu Glu Cys Leu Ile
 1205 1210 1215
 Lys Ile Phe Ala Leu Arg Gln Tyr Tyr Phe Thr Asn Gly Trp Asn Leu
 1220 1225 1230
 Phe Asp Cys Val Val Val Leu Leu Ser Ile Val
 1235 1240

<210> 9

<211> 20

<212> DNA

<213> Artificial Sequence

<220>
<221> variation
<222> (6)
<223> r = a or g

<220>
<223> Description of Artificial Sequence: rat NaN
forward primer no. 1

<400> 9
gaccertgga attggttgga 20

<210> 10
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: rat NaN
forward primer no. 2

<400> 10
aatccctgga attggttgga 20

<210> 11
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: rat NaN
forward primer no. 3

<400> 11
gaccctgga actggttaga 20

<210> 12
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: rat NaN
forward primer no. 4

<400> 12
gatctttgga actggcttga 20

<210> 13
<211> 21
<212> DNA
<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: rat Nan
forward primer no. 5

<400> 13

aacatagtgc tggagttcag g

21

<210> 14

<211> 21

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: rat NaN
forward primer no. 6

<400> 14

gtggcctttg gattccggag g

21

<210> 15

<211> 24

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: rat NaN
reverse primer no. 1

<400> 15

caagaaggcc cagctgaagg tgtc

24

<210> 16

<211> 24

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: rat NaN
reverse primer no. 2

<400> 16

gaggaatgcc. cacgcaaagg aatc

24

<210> 17

<211> 24

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: rat NaN
reverse primer no. 3

<400> 17
aagaaggac cagccaaagt tgtc

24

<210> 18
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: rat NaN
reverse primer no. 4, y = c or t, r = a or g, n =
a or c or g or t, w = a or t

<400> 18
acytccatrc anwcccat

20

<210> 19
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: rat NaN
reverse primer no. 5, r = a or g

<400> 19
agraartcna gccarcacca

20

<210> 20
<211> 19
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: rat NaN
reverse primer no. 6

<400> 20
tctgctgccg agccaggta

19

<210> 21
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: rat NaN
reverse primer no. 7

<400> 21
ctgagataac tgaaatcgcc

20

<210> 22
<211> 27
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: primer,
Marathon AP-1

<400> 22
ccatcctaatacgcactcactatagggc 27

<210> 23
<211> 23
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: primer,
Marathon AP-2

<400> 23
actcactatagggctcgagcggc 23

<210> 24
<211> 23
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: mouse NaN
forward primer

<400> 24
ccctgctgcgctcgggtgaagaag 23

<210> 25
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: mouse NaN
reverse primer

<400> 25
gacaaagtagatcccagagg 20

<210> 26
<211> 17
<212> DNA
<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: human NaN
forward primer

<400> 26

ctcagtagtt ggcattgc

17

<210> 27

<211> 24

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: human NaN
reverse primer

<400> 27

ggaaagaagc acgaccacac agtc

24

<210> 28

<211> 94

<212> PRT

<213> Rattus norvegicus

<220>

<223> C-terminal truncated rat NaN

<400> 28

Ala Ala Gly Gln Ala Met Arg Lys Gln Gly Asp Ile Leu Gly Pro Asn
1 5 10 15Ile His Gln Phe Ser Gln Ser Ser Glu Thr Pro Phe Leu Gly Cys Pro
20 25 30Gln Gln Arg Thr Cys Val Ser Phe Val Arg Pro Gln Arg Val Leu Arg
35 40 45Val Pro Trp Phe Pro Ala Trp Arg Thr Val Thr Phe Leu Ser Arg Pro
50 55 60Arg Ser Ser Glu Ser Ser Ala Trp Leu Gly Leu Val Glu Ser Ser Gly
65 70 75 80Trp Ser Gly Leu Pro Gly Glu Ser Gly Pro Ser Ser Leu Leu
85 90

<210> 29

<211> 211

<212> DNA

<213> Mus musculus

<400> 29

agttaaatgt tgagtgaatt gtggtggtga tttccactt gaggcctttg tgttaaagcc 60


```

caatgtgtgt gtgtgtgtgt gtgtgtgtgt gtgtgtgtgt gtgtgtgtgt gtgtgtgggt 120
gggggggtgt ggcagagtct ggtattggtt aggtgagagc aatcccagaa cgtccacctg 180
ctcttccatt ttattaatca ggcaggcctc t 211

```

```

<210> 30
<211> 242
<212> DNA
<213> Mus musculus

```

```

<400> 30
gtaagccact ggctcttaac taaaatgctc gttggcatta gaacatttct gagctggggg 60
ggtgggtgtg gtggtggtgg tgggtggtgg ggtgggtggg gtggtggtgg tgatgggtgg 120
ggtggaggtg gnggtggagg tgggtggctg ggtgggtggg gtggtggtgg tgggtggangt 180
ggangtgggt gcgtgggtgg gnggtggtg gtggaggtgg tggctgtggt ggtngtgggt 240
gc 242

```

```

<210> 31
<211> 200
<212> DNA
<213> Mus musculus

```

```

<400> 31
tgtgcatgct tgattcccag ctcttatggt ctgattactc ggtccttagg agcaaggcca 60
gactgtccac cctgacacac acacacacac acacacacac acacacacac acacacacac 120
acagtgtaga gaattacctc attcttggag tttctctgga aaaggaatgt ctcaaagcca 180
agttcacaga gcaacagctg 200

```

```

<210> 32
<211> 181
<212> DNA
<213> Mus musculus

```

```

<400> 32
tgttagaaac tctaagacaa tgaagcacca tgctggaaat aagagcacia actcttttctt 60
catgcattac ccactgcttg tgccttcacc ttagtgctcg tgctctctct ttctctctct 120
ctctctctct ctctctctct ctctctctct ctctctctct ctctctctct tttttttttt 180
t 181

```

```

<210> 33
<211> 128
<212> DNA
<213> Mus musculus

```

```

<400> 33
cacacacaca cacacacaca cacacacaca cacacacaca gagaaacact gtgcagtgca 60
tacatatata gataaatata tcttaaaaaa agaaccatgt gattgagtta taaaaatctc 128
caacttat

```

```

<210> 34
<211> 200
<212> DNA

```

<213> Mus musculus

<400> 34

```
agggtcatttc ctctgcagtg tgcttggcag gaaaaacttc ctggctattc aagtcagtgc 60
cctgcttgat catccatgta tcacacacac acaaaacaaa caaacaaca aacaaaaccc 120
tggggaagaa ggaagaggtt aagcacatag gcagagagca gccaggctga ctcagagcaa 180
acacctgac attcttccat                                     200
```

<210> 35

<211> 158

<212> DNA

<213> Mus musculus

<400> 35

```
gtgctgggat caaaggcgtg cgccgccacc acgcccggcc cctttttatg tttcaaattt 60
acttttatca tgtgcaegtg tgtgggtgctg tgcattgtgtg tgcgtgcgtg tgcgtgtgng 120
tgtgngtgtg tgtgtgtgtg tgtgtgtgtg tgtgtgtg                                     158
```

<210> 36

<211> 113

<212> DNA

<213> Mus musculus

<400> 36

```
cacacacaca cacacacaca cacacacaca cacacacaca cacacacaca cacacacttg 60
catctttgag ttaattggat aggctgagtc ttacaccgga atcatactgt tgc                                     113
```

<210> 37

<211> 200

<212> DNA

<213> Mus musculus

<400> 37

```
ccaatgagag actcttgtct caaaaaagcc atgggtgtcca gatcctgagg aataaacacct 60
aagaatgtgc tctggcctga aaacacacac acacacacac acacacacac acacacacac 120
agttttatatt atttatttaa aaaaatatgt ctctaggcat tgctgaaatg tctcctacag 180
gattaagtca accagagcca                                     200
```

<210> 38

<211> 7

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: protein seq.
basis for rat NaN reverse primers

<220>

<221> VARIANT

<222> (3)

<223> Xaa = Val or Asp

1/28
FIG. 1A

Nucleotide sequence of rat NaN. Translation initiation begins
at position 41 (ATG). Reading frame ends at position 5336 (TGA).

```

1  ACGGTGCCCT GATCCTCTGT ACCAGGAAGA CAGGGTGAAG ATGGAGGAGA
51  GGTACTACCC GGTGATCTTC CCGGACGAGC GGAATTTCCG CCCCTTCACT
101 TCCGACTCTC TGGCTGCCAT AGAGAAGCGG ATTGCTATCC AAAAGGAGAG
151 GAAGAAGTCC AAAGACAAGG CGGCAGCTGA GCCCCAGCCT CGGCCTCAGC
201 TTGACCTAAA GGCCTCCAGG AAGTTACCTA AGCTTTATGG TGACATTCCC
251 CCTGAGCTTG TAGCGAAGCC TCTGGAAGAC CTGGACCCAT TCTACAAAGA
301 CCATAAGACA TTCATGGTGT TGAACAAGAA GAGAACAATT TATCGCTTCA
351 GCGCCAAGCG GGCCTTGTTT ATTCTGGGGC CTTTTAATCC CCTCAGAAGC
401 TTAATGATTC GTATCTCTGT CCATTCAGTC TTTAGCATGT TCATCATCTG
451 CACGGTGATC ATCAACTGTA TGTTCATGGC GAATTCTATG GAGAGAAGTT
501 TCGACAACGA CATTCCCGAA TACGTCTTCA TTGGGATTTA TATTTTAGAA
551 GCTGTGATTA AAATATTGGC AAGAGGCTTC ATTGTGGATG AGTTTTCTTT
601 CCTCCGAGAT CCGTGGAAC TGGCTGGACTT CATTTGTCATT GGAACAGCGA
651 TCGCAACTTG TTTTCCGGGC AGCCAAGTCA ATCTTTCAGC TCTTCGTACC
701 TTCCGAGTGT TCAGAGCTCT GAAGGCGATT TCAGTTATCT CAGGTCTGAA
751 GGTCATCGTA GGTGCCCTGC TGCCTCGGT GAAGAAGCTG GTAGACGTGA
801 TGGTCCTCAC TCTCTTCTGC CTCAGCATCT TTGCCCTGGT CGGTCAGCAG
851 CTGTTTCATG GAATTCTGAA CCAGAAGTGT ATTAAGCACA ACTGTGGCCC
901 CAACCTGCA TCCAACAAGG ATTGTTTTGA AAAGGAAAAA GATAGCGAAG
951 ACTTCATAAT GTGTGGTACC TGGCTCGGCA GCAGACCCTG TCCCAATGGT
1001 TCTACGTGCG ATAAAACCAC ATTGAACCCA GACAATAATT ATACAAAGTT
1051 TGACAACTTT GGCTGGTCCT TTCTCGCCAT GTTCCGGGTT ATGACTCAAG
1101 ACTCCTGGGA GAGGCTTTAC CGACAGATCC TGCAGGACCTC TGGGATCTAC
1151 TTTGTCTTCT TCTTCGTGGT GGTTCATCTT CTGGGCTCCT TCTACCTGCT
1201 TAACCTAACC CTGGCTGTTG TCACCATGGC TTATGAAGAA CAGAACAGAA
1251 ATGTAGCTGC TGAGACAGAG GCCAAGGAGA AAATGTTTCA GGAAGCCCAG
1301 CAGCTGTAA GGGAGGAGAA GGAGGCTCTG GTTGCCATGG GAATTGACAG
1351 AAGTTCCCTT AATTCCCTTC AAGCTTCATC CTTTTCCTCCG AAGAAGAGGA
1401 AGTTTTTCGG TAGTAAGACA AGAAAGTCCT TCTTTATGAG AGGGTCCAAG
1451 ACGGCCCAAG CCTCAGCGTC TGATTCAGAG GACGATGCCT CTAAAAATCC
1501 ACAGCTCCTT GAGCAGACCA AACGACTGTC CCAGAACTTG CCAGTGGATC
1551 TCTTTGATGA GCACGTGGAC CCCCTCCACA GGCAGAGAGC GCTGAGCGCT
1601 GTCAGTATCT TAACCATCAC CATGCAGGAA CAAGAAAAAT TCCAGGAGCC
1651 TTGTTTCCCA TGTGGGAAAA ATTTGGCCTC TAAGTACCTG GTGTGGGACT

```

FIG. 1B

2/28

1701 GTAGCCCTCA ATGGCTGTGC ATAAAGAAGG TCCTGCGGAC CATCATGACG
 1751 GATCCCTTTA CTGAGCTGGC CATCACCATC TGCATCATCA TCAATACCGT
 1801 TTTCTTAGCC GTGGAGCACC ACAACATGGA TGACAACCTTA AAGACCATAC
 1851 TGAAAATAGG AAACCTGGGT TTCACGGGAA TTTTCATAGC GGAAATGTGT
 1901 CTCAAGATCA TCGCGCTCGA CCCTTACCAC TACTTCCGGC ACGGCTGGAA
 1951 TGTTTTTGAC AGCATCGTGG CCCTCCTGAG TCTCGCTGAT GTGCTNTACA
 2001 ACACACTGTC TGATAACAAT AGGTCTTTCT TGGCTTCCCT CAGAGTGCTG
 2051 AGGGTCTTCA AGTTAGCCAA ATCCTGGCCC ACGTTAAACA CTCTCATTA
 2101 GATCATCGGC CACTCCGTGG GCGCGCTTGG AAACCTGACT GTGGTCTTGA
 2151 CTATCGTGGT CTTTCATCTTT TCTGTGGTGG GCATGCGGCT CTTGCGCACC
 2201 AAGTTTAACA AGACCGCCTA CGCCACCCAG GAGCGGCCCA GGCGGCGCTG
 2251 GCACATGGAT AATTTCTACC ACTCCTTCCT GGTGGTGTTC CGCATCCTCT
 2301 GTGGGGAATG GATCGAGAAC ATGTGGGGCT GCATGCAGGA TATGGACGGC
 2351 TCCCCGTGTG GCATCATTTG CTTTGTCTTG ATAATGGTGA TCGGGAAGCT
 2401 TGTGGTGTCT AACCTCTTCA TTGCCTTGCT GCTCAATTCC TTCAGCAATG
 2451 AGGAGAAGGA TGGGAGCCTG GAAGGAGAGA CCAGGAAAC CAAAGTGCAG
 2501 CTAGCCCTGG ATCGGTTCCT CCGGGCCTTC TCCTTCATGC TGCACGCTCT
 2551 TCAGAGTTTT TGTGCAAGA AATGCAGGAG GAAAACTCG CCAAAGCCAA
 2601 AAGAGACAAC AGAAAGCTTT GCTGGTGAGA ATAAAGACTC AATCCTCCCG
 2651 GATGCGAGGC CCTGGAAGGA GTATGATACA GACATGGCTT TGTACACTGG
 2701 ACAGGCCGGG GCTCCGCTGG CCCCCTCGC AGAGGTAGAG GACGATGTGG
 2751 AATATGTGGG TGAAGGCGGT GCCCTACCCA CCTCACAACA TAGTGCTGGA
 2801 GTTCAGGCCG GTGACCTCCC TCCAGAGACC AAGCAGCTCA CTAGCCCGGA
 2851 TGACCAAGGG GTTGAAATGG AAGTATTTTC TGAAGAAGAT CTGCATTTAA
 2901 GCATACAGAG TCCTCGAAAG AAGTCTGACG CAGTGAGCAT GCTCTCGGAA
 2951 TGCAGCACAA TTGACCTGAA TGATATCTTT AGAAATTTAC AGAAAACAGT
 3001 TTCCCCCAA AAGCAGCCAG ATAGATGCTT TCCCAAGGGC CTTAGTTGTC
 3051 ACTTTCATG CCACAAAACA GACAAGAGAA AGTCCCCCTG GGTCTGTGG
 3101 TGGAACATTC GGAAAACCTG CTACCAAATC GTGAAGCACA GCTGGTTTGA
 3151 GAGTTTCATA ATCTTTGTTA TTCTGCTGAG CAGTGGAGCG CTGATATTTG
 3201 AAGATGTCAA TCTCCCCAGC CGGCCCCAAG TTGAGAAATT ACTAAGGTGT
 3251 ACCGATAATA TTTTCACATT TATTTTCCTC CTGGAATGA TCCTGAAGTG
 3301 GGTGGCCTTT GGATTCGGGA GGTATTTTAC CAGTGCCTGG TGCTGGCTTG
 3351 ATTTCCTCAT TGTGGTGGTG TCTGTGCTCA GTCTCATGAA TCTACCAAGC
 3401 TTGAAGTCTT TCCGGACTCT GCGGGCCCTG AGACCTCTGC GGGCGCTGTC
 3451 CCAGTTTGAA GGAATGAAGG TTGTCGTCTA CGCCCTGATC AGCGCCATAC
 3501 CTGCCATTC CAATGTCTTG CTGGTCTGCC TCATTTTCTG GCTCGTATTT
 3551 TGTATCTTGG GAGTAAATTT ATTTTCTGGG AAGTTTGGA GGTGCATTAA
 3601 CGGGACAGAC ATAAATATGT ATTTGGATT TACCGAAGTT CCGAACCAG

FIG. 1C

3/28

3651 GCCAATGTAA CATTAGTAAT TACTCGTGGA AGGTCCCGCA GGTCAACTTT
 3701 GACAACGTGG GGAATGCCTA TCTCGCCCTG CTGCAAGTGG CAACCTATAA
 3751 GGGCTGGCTG GAAATCATGA ATGCTGCTGT CGATTCCAGA GAGAAAGACG
 3801 AGCAGCCGGA CTTTGAGGCG AACCTCTACG CGTATCTCTA CTTTGTTGGT
 3851 TTTATCATCT TCGGCTCCTT CTATTACCCTG AACCTCTTTA TCGGTGTTAT
 3901 TATTGACAAC TTCAATCAGC AGCAGAAAAA GTTAGGTGGC CAAGACATTT
 3951 TTATGACAGA AGAACAGAAG AAATATTACA ATGCAATGAA AAAGTTAGGA
 4001 ACCAAGAAAC CTCAAAAGCC CATCCCAAGG CCCCTGAACA ANTGTCAAGC
 4051 CTTTGTGTTC GACCTGGTCA CAAGCCATGT CTTTGACGTC ATCATCTCTG
 4101 GTCTTATTGT CTAAATATG ATTATCATGA TGGCTGAATC TGCCGACCAG
 4151 CCCAAAGATG TGAAGAAAAC CTTTGATATC CTCAACATAG CCTTCGTGGT
 4201 CATCTTTACC ATAGAGTGTC TCATCAAAGT CTTTGCTTTG AGGCAACACT
 4251 ACTTCACCAA TGGCTGGAAC TTATTTGATT GTGTGGTCGT GGTCTTTCT
 4301 ATCATTAGTA CCCTGGTTTC CCGCTTGGAG GACAGTGACA TTTCTTTCCC
 4351 GCCCACGCTC TTCAGAGTCG TCCGCTTGGC TCGGATTGGT CGAATCCTCA
 4401 GGCTGGTCCG GGCTGCCCCG GGAATCAGGA CCCTCCTCTT TGCTTTGATG
 4451 ATGTCTCTCC CCTCTCTCTT CAACATCGGT CTGCTGCTCT TCCTGGTGAT
 4501 GTTCATTTAC GCCATCTTTG GGATGAGCTG GTTTTCCAAA GTGAAGAAGG
 4551 GCTCCGGGAT CGACGACATC TTCAACTTCG AGACCTTTAC GGGCAGCATG
 4601 CTGTGCCTCT TCCAGATAAC CACTTCGGCT GGCTGGGATA CCCTCCTCAA
 4651 CCCCATGCTG GAGGCAAAAG AACACTGCAA CTCCTCCTCC CAAGACAGCT
 4701 GTCAGCAGCC GCAGATAGCC GTCGCTACT TCGTCAGTTA CATCATCATC
 4751 TCCTTCCTCA TCGTGGTCAA CATGTACATC GCTGTGATCC TCGAGAACTT
 4801 CAACACAGCC ACGGAGGAGA GCGAGGACCC TCTGGGAGAG GACGACTTTG
 4851 AAATCTTCTA TGAGGTCTGG GAGAAGTTTG ACCCCGAGGC GTCGCAGTTC
 4901 ATCCAGTATT CGGCCCTCTC TGACTTTGCG GACGCCCTGC CGGAGCCGTT
 4951 GCGTGTGGCC AAGCCGAATA AGTTTCAGTT TCTAGTGATG GACTTGCCCA
 5001 TGGTGATGGG CGACCGCCTC CATTCGATGG ATGTTCTCTT TGCTTTCACT
 5051 ACCAGGGTCC TCGGGGACTC CAGCGGCTTG GATACCATGA AAACCATGAT
 5101 GGAGGAGAAG TTTATGGAGG CCAACCTTTT TAAGAAGCTC TACGAGCCCA
 5151 TAGTCACCAC CACCAAGAGG AAGGAGGAGG AGCAAGGCGC CGCCGTCATC
 5201 CAGAGGGCCT ACCGGAACA CATGGAGAAG ATGGTCAAAC TGAGGCTGAA
 5251 GGACAGGTCA AGTTCATCGC ACCAGGTGTT TTGCAATGGA GACTTGTTCA
 5301 GCTTGATGTG GGCCAAGGTC AAGGTTTACA ATGACTGAAC CCTCATCTCC
 5351 ACCCCTACCT CACTGCCTCA CAGCTTAGCC TCCAGCCTCT GCGGAGCAGG
 5401 CGGCAGACTC ACTGAACACA GGCCGTTCGA TCTGTGTTTT TGCTGTAACG
 5451 AGGTGACAGG TTGGCGTCCA TTTTAAATG ACTCTTGGA AGATTTTCATG
 5501 TAGAGAGATG TTAGAAGGGA CTGCAAAGGA CACCGACCAT AACGGAAGGC
 5551 CTGGAGGACA GTCCAACTTA CATAAAGATG AGAAACAAGA AGGAAAGATC

FIG. 1D

5601 CCAGGAAAAC TTCAGATTGT GTTCTCAGTA CATCCCCAA TGTGTCTGTT
5651 CGGTGTTTTG AGTATGTGAC CTGCCACATG TAGCTCTTTT TTGCATGTAC
5701 GTCAAAACCC TGCAGTAAGT TAATAGCTTG CTACGGGTGT TCCTACCAGC
5751 ATCACAGAAT TGGGTGTATG ACTCAAACCT AAAAGCATGA CTCTGACTTG
5801 TCAGTCAGCA CCCCRACTTT CAGACGCTCC AATCTCTGTC CCAGGTGTCT
5851 AACGAATAAA TAGGTAAAAG AAAAA

FIG. 2A

5/28

Predicted amino acid sequence of rat NaN (1765 a.a).

1 MEERYYPVIF PDERNFRPFT SDSLAAIEKR IAIQKERKKS KDKAAAEPQP

51 RPQLDLKASR KLPKLYGDIP PELVAKPLED LDPFYKDHTK FMVLNKKRTI
DI-S1

101 YRFSAKRALF ILGPFNPLRS LMIRISVHSV FSMFIICTVI INCMFMANS
DI-S2 DI-S3

151 ERSFDNDIPE YVFIGIYILE AVIKILARGF IVDEFSFLRD PWNWLD FIVI
DI-S4

201 GTAIATCFPG SQVNLSALRT FRVFRALKAI SVISGLKVIV GALLRSVKKL
DI-S5

251 VDVMVLTLEFC LSIFALVGQO LFMGILNQKC IKHNCGPNPA SNKDCFEKEK
DI-SS1

301 DSEDFIMCGT WLGSRPCPNG STCDKTTLNP DNNYTKFDNF GWSFLAMFRV
DI-SS2 DI-S6

351 MTODSWERLY RQILRTSGIY FVFFFVVVIF LGSFYLLNLT LAVVTMAYEE

401 QNRNVAAETE AKEKMFQEAQ QLLREEKEAL VAMGIDRSSL NSLQASSFSP

451 KKRKFFGSKT RKSFFMRGSK TAQASASDSE DDASKNPQLL EQTKRLSQNL

501 PVDLFDEHVD PLHRQRALSA VSILTITMQE QEKFQEPCCP CGKNLASKYL
DII-S1

551 VWDCSPQWLC IKKVLRTIMT DPFTELAITI CIIINTVFLA VEHNMDDNL
DII-S2 DII-S3

601 KTILKIGNWV FTGIFIAEMC LKIIALDPYH YFRHGWNVFD SIVALLSLAD
DII-S4

651 VLNTLSDNN RSFLASLRVL RVFKIAKSWP TLNTLIKIIG HSVGALGNLT
DII-S5 DII-SS1

701 VVLTVVFIF SVVGMRLFGT KFNKTAYATQ ERPRRRWHMD NEYHSFLVVF
DII-SS2 DII-S6

751 RILCGEWIEN MWGCMQDMDG SPLCIIVFVL IMVIGKLVVL NLFIALLLNS

FIG. 2B

6/28

801 FSNEEKDGSL EGETRKTQVQ LALDRFRRAF SFMLHALQSF CCKKCRRKNS
851 PKPKETTESF AGENKDSILP DARPWKEYDT DMALYTGQAG APLAPLAEVE

901 DDVEYCGEGG ALPTSQHSAG VQAGDLPPET KQLTSPDDQG VEMEVFSEED

951 LHLSIQSPRK KSDAVSMLSE CSTIDLNDIF RNLQKTVSPK KQPDRCFPKG
DIII-S1
1001 LSCHFLCHKT DKRKSPWVLW WNIRKTCYQI VKHSWFESFI IFVILLSSGA
DIII-S2
1051 LIFEDVNLPS RPQVEKLLRC TDNIFTFIFL LEMILKWVAF GERRYFTSAW
DIII-S3 DIII-S4
1101 CWLDFLIVVV SVLSLMNLPS LKSFRTLRL RPLRALSOFE GMKVVVYALI
DIII-S5
1151 SAIPAILNVL LVCLIFWLVE CILGVNLESG KFGRCINGTD INMYLDFTEV
DIII-SS1 DIII-SS2
1201 PNRSQCNISN YSWKVPQVNF DNVGNAYLAL LOVATYKGWL EIMNAAVDSR
DIII-S6
1251 EKDEQPDFEA NLYAYLYFVV FIIFGSFETL NLEFIGVIIDN FNQQQKKLGG
DIV-S1
1301 QDIFMTEEQK KYYNAMKKLG TKKPQKPIPR PLNRCQAFVF DLVTSHVFDV

1351 IILGLIVLNM IIMMAESADQ PKDVKKTFDI LNIAFVVIFT IECLIKVFAL
DIV-S4
1401 RQHYFTNGWN LFDCVVVVLS IISTLVSRL E DSDISFPPTL FRVRLARIG
DIV-S5
1451 RILRLVRAAR GIRTLFALM MSLPSLFNIG LLLFLVMFIY AIFGMSWFSK
DIV-SS1 DIV-SS2
1501 VKKGSGIDDI FNFTFTGSM LCLFOITSA GWDTLNPM L EAKEHCNSSS
DIV-S6
1551 QDSCQQPOIA VVYFVSYIII SPLIVNMYI AVILENFNTA TEESEDPLGE

1601 DDFEIFYEVW EKFDPEASQF IQYSALSDFA DALPEPLRVA KPNKFQFLVM

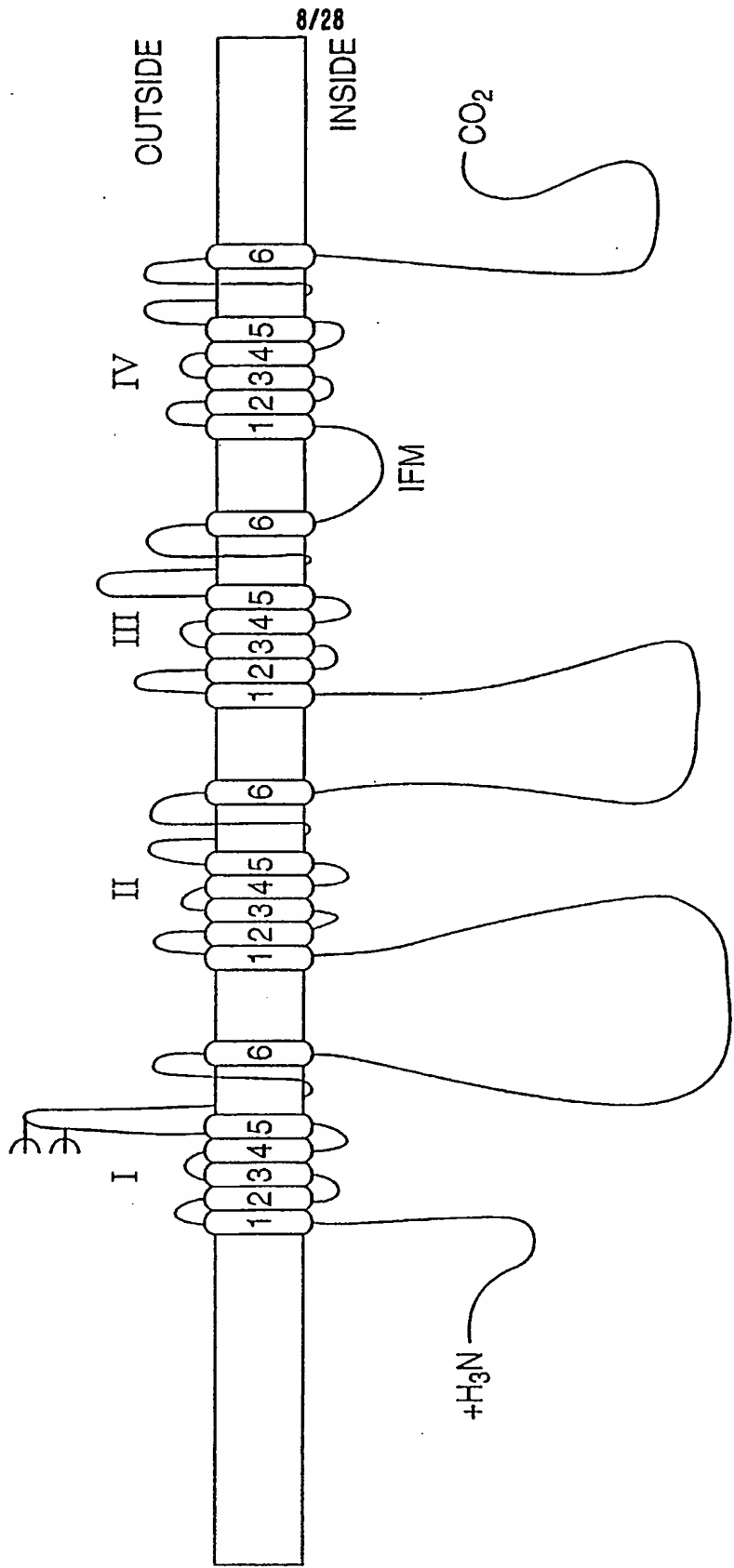
1651 DLPMVMGDRL HCMDVLFAFT TRVLGDSSGL DTMKTMMEEK FMEANPFKKL

FIG. 2C

1701 YEPIVTTTKR KEEEQGAIVI QRAYRKHMEK MVKLRLKDRS SSSHQVFCNG

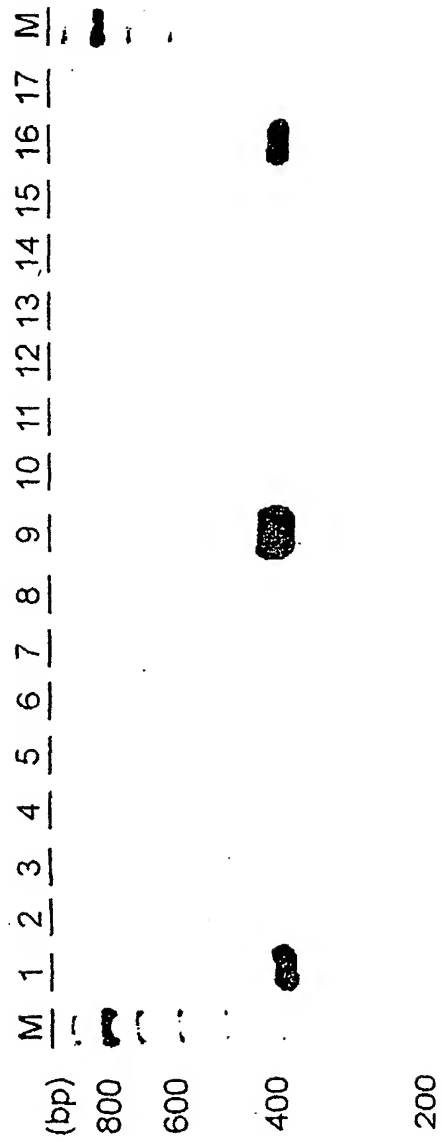
1751 DLSSLDVAKV KVHND*

FIG. 3
VOLTAGE-GATED SODIUM CHANNEL α SUBUNIT



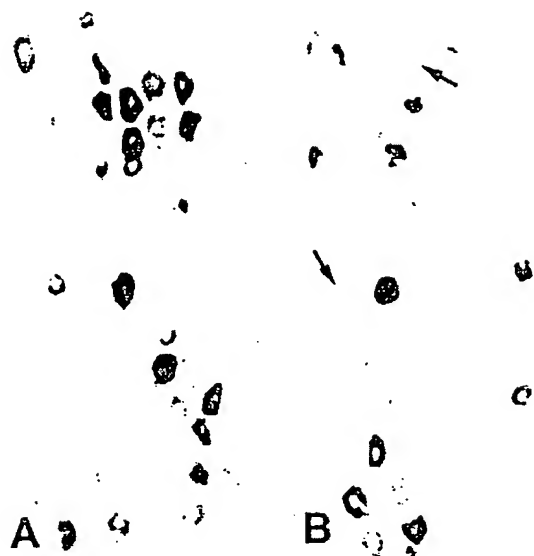
9/28

FIG. 4



10/28

FIG. 5



C — D

E F —

II/28

FIG. 6

RESTRICTION ENZYME ANALYSIS OF α -SUBUNIT PCR PRODUCTS FROM DOMAIN I USING THE FOLLOWING PRIMERS: NACHD1A.1-4 (FORWARD PRIMERS) AND NAAGEN.REV1-3 (REVERSE PRIMERS).

Generic Primer pair	F1 R1	F2 R1	F1 R1	F1/F3 R1	F1 R1	F1/F3 R1	F1 R1	F1/F3 R1	F1 R1	F2 R2	F4 R3	F2 R3
	α I 558 bp	α II 561 bp	α III 561 bp	α VI 507 bp	α PN1 501 bp	α RH1 518 bp	α u1 602 bp	α NS 479 bp	α NaG 501 bp	α NaN 468 bp		
EcoR V	+	-	-	-	-	-	-	-	-	-	-	-
EcoNI	-	+	-	-	-	-	-	-	-	-	-	-
Ava I	-	-	+	-	-	-	-	-	-	-	-	-
Sph I	-	-	-	+	-	-	-	-	-	-	-	-
Bam H I	-	-	-	-	+	-	-	+	-	-	-	-
Acc I	-	-	-	-	-	+	-	-	-	-	+	185, 283
Ngo M I	-	-	-	-	-	-	+	-	+	-	+	-
Afl II	-	-	-	-	-	-	-	+	-	+	-	-
Xba I	-	-	-	-	-	-	-	-	+	-	+	-
EcoR I	-	-	-	-	-	-	-	-	-	-	-	+

12/28
FIG. 7A-1

Sequence of th mouse NaN cDNA.

```

1      TCTGAGCCAA GGGTGAAGAT GGAGGAGAGG TACTATCCAG TGATCTTCCC AGACGAGAGG
61     AATTTCCGCC CCTTCACTTT CGACTCTTTG GCTGCAATAG AGAAGCGGAT CACCATCCAA
121    AAGGAGAAGA AGAAATCCAA AGACAAGGCA GCAACTGAGC CCCAGCCTCG GCCTCAGCTC
181    GACCTAAAGG CCTCCAGGAA GTTACCTAAG CTCTATGGCG ACGTTCCCCC TGACCTTATA
241    GCGAAGCCCC TGGAAGATCT GGACCCATTT TACAAAGACC ATAAGACATT CATGGTATTG
301    AACAGAAGA GAACAATCTA TCGCTTCAGC GCCAAGAGGG CCTTGTTTCAT TCTGGGGCCT
361    TTTAATCCCA TCAGAAGCTT CATGATTCGC ATCTCTGTCC ATTCAGTCTT CAGCATGTTC
421    ATTATCTGCA CAGTGATCAT CAACTGTATG TTCATGGCTA ATAATTCTTC TGTGGACAGT
481    CGTCCTAGCA GTAACATTCC CGAATACGTC TTCATTGGGA TTTATGTTTT AGAAGCTGTG
541    ATTAAAATAT TGGCAAGAGG CTTCAATTGT GATGAGTTTT CCTACCTCCG AGATCCTTGG
601    AACTGGCTGG ACTTCATTGT CATCGGAACA GCGATAGCGC CTGTGTTTTCT CGGTAACAAA
661    GTCAATAATC TTTCCACTCT ACGTACCTTC CGAGTGTGGA GAGCTCTGAA AGCCATTTCT
721    GTAATCTCAG GTCTGAAGGT CATCGTGGGT GCCCTGCTGC GCTCCGTGAA GAAGCTAGTG
781    GACGTGATGG TCCTCACTCT CTTTTGCCTC AGCATCTTTG CCCTGGTTGG TCAGCAGCTC
841    TTCATGGGAA TTCTGAGCCA GAAATGTATT AAGGACGACT GTGGCCCTAA CGCTTTTTCC
901    AACAGGATT GCTTTGTAAA AGAAAATGAT AGCGAGGACT TCATAATGTG TGGCAACTGG
961    CTCGGCAGAA GATCCTGCCC CGATGGTTCC ACGTGCAATA AAACCACATT TAACCCAGAT
1021   TATAATTATA CAACTTTGA CAGCTTTGGC TGGTCTTTTC TCGCCATGTT CCGGGTTATG
1081   ACTCAAGACT CCTGGGAGAA GCTTTATCGA CAGATCCTTC GCACCTCCGG GATCTACTTT
1141   GTCTTCTTCT TCGTGGTCGT CATCTTCCTG GGCTCTTTCT ACCTGCTTAA CTTAACCCGTG
1201   GCTGTGCTCA CCATGGCTTA CGAGGAACAG AACAGAAATG TCGCTGCCGA GACAGAGGCC
1261   AAGGAGAAGA TGTTTCAGGA AGCCCAGCAG CTGTTGAGGG AGGAAAAGGA GGCTCTGGTT
1321   GCCATGGGAA TTGACAGAAC TTCCCTTAAT TCCCTCCAAG CTTGTCCTT TTCCCCAAAG
1381   AAGAGGAAGT TTTTGGCAG TAAGACAAGA AAGTCCTTCT TTATGAGAGG GTCCAAGACA
1441   GCGCGAGCCT CAGCGTCCGA TTCAGAGGAC GATGCCTCTA AAAACCCACA ACTCCTTGAG
1501   CAAACAAAAC GACTATCCCA GAACTTGCCC GTAGAACTCT TTGATGAGCA CGTGGACCCC
1561   CTCCATAGGC AGAGAGCGCT GAGTGCCGTC AGTATCTTAA CCATCACCAT GCAGGAACAA
1621   GAAAAATCCC AGGAGCCTTG TTTCCCGTGT GGGAAAAACT TGGCATCCAA GTACCTGGTG
1681   TGGGAATGTA GCCCTCCGTG GCTGTGCATA AAGAAGGTCC TGCAGACTAT CATGACAGAC
1741   CCCTTCACTG AGCTGGCCAT CACCATCTGC ATCATCGTCA AACTGTCTT CTTGGCCATG
1801   GAACACCACA ATATGGATAA CTCTTTAAAA GACATACTGA AAATAGGAAA CTGGGTTTTTC
1861   ACTGGAATTT TCATAGCGGA AATGTGTCTC AAGATCATTG CGCTAGACCC TTACCACTAC
1921   TTCCGGCACG GCTGGAACAT CTTTGACAGC ATTGTGGCCC TTGTGAGTCT CGCTGACGTG

```

13/28
FIG. 7A-2

1981 CTCTTCCACA AACTGTCTAA AAACCTCTCC TTCTTGGCTT CCCTCAGAGT GCTGAGGGTC
 2041 TTCAAGTTAG CCAAATCCTG GCCCACATTA AACACTCTCA TTAAGATCAT CGGCCACTCC
 2101 GTGGGTGCGC TCGGAAACCT GACTGTGGTC CTAACGATCG TGGTCTTCAT CTTTCCGTG
 2161 GTTGGCATGC GGCTCTTTGG TGCCAAGTTT AACAAGACTT GCTCCACCTC TCCGGAGTCC
 2221 CTCCGGCGCT GGCACATGGG TGATTTCTAC CATTCTTCC TGGTGGTGTT CCGCATCCTC
 2281 TGTGGGGAGT GGATCGAGAA CATGTGGGAA TGCATGCAGG AGATGGAAGG CTCCCCGTG
 2341 TGTGTCATCG TCTTTGTGCT GATCATGGTG GTCGGGAAGC TCGTGGTGCT TAACCTCTTC
 2401 ATTGCCTTGC TGCTCAATTC CTTCAGCAAT GAGGAAAAGG ATGGGAACCC AGAAGGAGAG
 2461 ACCAGGAAAA CCAAAGTGCA GCTAGCCCTG GATCGGTTCA GCCGAGCGTT CTACTTCATG
 2521 GCGCGCGCTC TTCAGAATTT CTGTTGCAAG AGATGCAGGA GGCAAACTC GCCAAAGCCA
 2581 AATGAGGCAA CAGAAAGCTT TGCTGGTGAG AGTAGAGACA CAGCCACCCT GGATACAAGG
 2641 TCCTGGAAGG AGTATGATTC AGAAATGACT CTGTACACTG GGCAGGCCGG GGCTCCACTG
 2701 GCCCCACTGG CAAAAGAAGA GGACGATATG GAATGTTGTG GTGAATGTGA TGCCTCACCT
 2761 ACCTCACAGC CTAGTGAGGA AGCTCAGGCC TGTGACCTCC CTCTGAAGAC CAAGCGGCTC
 2821 CCCAGCCCAQ ATGACCACGG GGTGAAATG GAAGTGTGTTT CCGAAGAAGA TCCGAATTTA
 2881 ACCATACAGA GTGCTCGAAA GAAGTCTGAT GCGGCAAGCA TGCTCTCAGA ATGCAGCACA
 2941 ATAGACCTGA ATGATATCTT TAGAAATTTA CAGAAAACAG TTTCCCCCA AAAGCAACCA
 3001 GATCGATGCT TTCCCAAGGG CCTCAGTTGT ATCTTTCTAT GTTGCAAAAC AATCAAAAAA
 3061 AAGTCCCCCT GGTCTCTGTG GTGGAATCTT CGGAAAACCT GCTACCAAT CGTGAAGCAT
 3121 AGCTGOTTG AGAGCTTCAT AATTTTGTG ATCCTGTGTA GCAGCGGAGC ACTGATATTC
 3181 GAAGATGTCA ATCTTCCCAQ CCGGCCCAA GTTGA AAAAT TACTGAAGTG TACCGATAAT
 3241 ATTTTCACAT TTATTTTCT CCTGGAATG ATTTTGAAGT GGGTGGCCTT TGGATTCCGG
 3301 AAGTATTTCA CCAGTGCTG GTGCTGGCTC GATTTCTCA TTGTGGTGGT GTCTGTGCTC
 3361 AGCCTCACGA ACTTACCAA CTTGAAGTCC TTCCGGAATC TGCGAGCGCT GAGACCTCTG
 3421 CGGGCACTGT CTCAGTTTGA AGGAATGAAG GTTGTGTGTA ATGCCCTCAT GAGTGCCATA
 3481 CCTGCCATCC TCAATGTCTT GCTGGTCTGC CTCATTTTCT GGCTCATATT TTGTATCCTG
 3541 GGAGTAAATT TTTTCTCTG GAAGTTTGA AGATGCATTA ATGGAACAGA CATAAATAAA
 3601 TATTTCAACG CTCCAATGT TCCAAACCAA AGCCAATGTT TAGTTAGTAA TTACACGTGG
 3661 AAAGTCCCGA ATGTCAACTT TGACAACGTG GGGAAATGCCT ACCTTGCCCT GCTGCAAGTG
 3721 GCGACCTATA AGGGCTGGCT GGACATTATG AATGCAGCTG TTGATTCCAG AGGGAAAGAT
 3781 GAGCAGCCGG CCTTTGAGGC GAATCTATAC GCATACCTTT ACTTCGTGGT TTTATCATC
 3841 TTCGGCTCAT TCTTTACCCT GAACCTCTT ATCGGTGTTA TTATTGACAA CTTCAATCAG
 3901 CAGCAGAAAA AGTTAGGTGG CCAAGACATT TTTATGACAG AAGAACAGAA GAAATATTAC
 3961 AATGCAATGA AAAAGTTAGG AACCAAGAAG CCTCAAAAGC CCATCCCAAG GCCCCTGAAC
 4021 AAATGTCAAG CCTTCGTGTT CGATTTGGTC ACAAGCCAGG TCTTTGACGT CATCATCTG
 4081 GGTCTTATTG TCACAAACAT GATTATCATG ATGGCTGAAT CTGAAGGCCA GCCCAACGAA
 4141 GTGAAGAAAA TCTTTGATAT TCTCAACATA GTCTTCGTG TCATCTTTAC CGTAGAGTGT
 4201 CTCATCAAAG TCTTTGCTTT GAGGCAACAC TACTTCACCA ATGGCTGGAA CTTATTGAT

14/28

FIG. 7A-3

4261 TGTGTGGTCG TGGTTCTTTC CATCATTAGT ACCTTGGTTT CTGGCTTGGG GAACAGCAAC
4321 GTCTTCCCGC CCACACTCTT CAGGATTGTC CGCTTGGCTC GGATCGGTCG AATCCTCAGA
4381 CTGGTCCGGG CGGCTCGAGG AATCAGGACA CTCCTTTTCG CGTTGATGAT GTCTCTCCCC
4441 TCTCTCTTCA ACATTGGTCT GCTTCTCTTT CTGGTGATGT TCATTTATGC CATCTTTGGG
4501 ATGAACTGGT TTTCCAAAGT GAAGAGAGGC TCTGGGATTG ATGACATCTT CAACTTTGAC
4561 ACTTTCTCGG GCAGCATGCT CTGCCTCTTC CAGATAACCA CTTCAGCCCG CTGGGATGCT
4621 CTCCTCAACC CCATGCTGGA ATCAAAAGCC TCTTGCAATT CCTCCTCCCA AGAGAGCTGT
4681 CAGCAGCCGC AGATAGCCAT AGTCTACTTC GTCAGCTACA TCATCATCTC CTTTCTCATT
4741 GTGGTTAACA TGTACATAGC TGTGATTCTA GAGAACTTCA ACACAGCCAC AGAGGAGAGC
4801 GAGGACCCCC TGGGCGAAGA CGACTTTGAG ATCTTCTATG AGATCTGGGA GAAAGTTGAC
4861 CCCGAAGCAA CACAGTTCAT CCAGTACTCA TCCCTCTCTG ACTTCGCCGA CGCCCTGCCC
4921 GAGCCGTTGC GTGTGGCCAA GCCCAACAGG TTTCAGTTTC TCATGATGGA CTGCCCCATG
4981 GTGATGGGTG ATCGCCTCCA TTGCATGAT GTTCTCTTTG CTTTCACCAC CAGGGTCCTC
5041 GGGAACTCCA GCGGCTTGGG TACCATGAAA GCCATGATGG AGGAGAAGTT CATGGAGGCC
5101 AATCCTTTCA AGAAGTTGTA CGAGCCCAT TGCACCACCA CAAAGAGGAA GGAGGAGGAG
5161 GAATGTGCCG CTGTATCCA GAGGCGCTAC CGGAGACACA TGGAGAAGAT GATCAAGCTG
5221 AAGCTGAAAG GCAGGTCAAG TTCATCGCTC CAGGTGTTTT GCAATGGAGA CTGTCTAGC
5281 TTGGATGTGC CCAAGATCAA GGTTCATTGT GACTGAAACC CCCACCTGCA CGCCTACCTC
5341 ACAGCCTCAC AGCTCAGCCC CCAGCCTCTG GCGAACCAAGC GGCGGACTCA CCGAACAGGC
5401 CGTTCAACTT GTTTTTTTGG GTGAAAGAGG TGATAGGTTG GTGTCCATTT TTAATGATT
5461 CTTGGAAAGA TTGAACGTCG GAACATGTTA GAAAGGACTG CCAAGGACAT CCACAGTAAC
5521 GGAAGGCCTG AAGGACAGTT CAAATTATGT AAAGAAACGA GAAGGAAAGG TCACATGTCT
5581 GTTCAGTTTT AAGTATGTGA CTTGCCACAT GTAGCTCCTT TGCATGTTAA GTGAGAAGTC
5641 AAAACCCTGC CATAAGTAAA TAGCTTTGTT GCAGGTGTTT CTACCAGTGC TCCGGATTTG
5701 GGTGTATGGC TCAAACCTGA AAGCATGACT CTGACTTGTC AGCACCCCAA CTTTCAGAAG
5761 CTCTGATCTC TGTCCTAGGT GTTTGACAAA TAAATACATA AAANAAAAAA AAAAAAAAAA
5821 AA

15/28
FIG. 7B-1

Protein sequence of mNaN

Molecular Weight 201451.00 Daltons

1765 Amino Acids

198 Strongly Basic(+) Amino Acids (K,R)

177 Strongly Acidic(-) Amino Acids (D,E)

712 Hydrophobic Amino Acids (A,I,L,F,W,V)

453 Polar Amino Acids (N,C,Q,S,T,Y)

8.260 Isoelectric Point

22.540 Charge at PH 7.0

```

1  MEERYYPVIF PDERNFRPPT FDSLAAIEKR ITIQEKKKKS KDKAATEPQP RPQLDLKASR
21  KLPKLYGDVP PDLIAKPLED LDPFYKDHKT FMVLNKKRTI YRFSAKRALF ILQPFNPIRS
121 FMIRISVHSV PSMFIICTVI INCMEMANNS SVDSRPSSNI PRYVFIGIYV LEAVIKILAR
      DI-S1                               DI-S2
181 GFIVDEFSYL RDPHNWLDEF VIGTAIAPCF LGNKVMNLST LRTERVLRAL KAISVISGLK
      DI-S3                               DI-S4
241 VIVGALLRSV KKLVDVMVLT LECLSIKALV GOOLEFMGILS QKCINDDCGP NAFSNKDCPV
      DI-S5
301 KENDSEDFIM CGNWLGRRSC PDGSTCNKTT FNPDYNYTNF DSEGHSPILAM FRVMTQDSWE
      DI-SS1 DI-SS2
361 KLYRQILRTS GIYFVEFFVY VIPLGSEYLL NLTAVVTMA YEEQNRNVAA ETEAKEKMFQ
      DI-S6
421 EAQQLLREEK EALVAMQIDR TSLNSLQASS FSPKKRKFFG SKTRKSFFMR GSKTARASAS
481 DSEDDASKNP QLLBQTKRLS QNLPVELFDE HVDPLHRQRA LSAVSILTIT MQEQEKSQEP
541 CFFCGKNLAS KYLVWECSPF WLCIKKVLQT IMTDPTELA ITICIIVNTV FLAMEHHNMD
      DII-S1
601 NSLKDILKIG NHVETGIFIA EMCLKIIALD PYHYFRHGWN IFDSIVALVS LADVLEHKLS
      DII-S2                               DII-S3
661 KNLSPLASLR VLRVEKLAKS WPTLNTLIKI IGHSVGLGN LTVVLTIVVF IFSVVGMRLE
      DII-S4                               DII-S5
721 GAKFNKTCST SPESLRRWHM GDEYHSFLVV FRILCGEWIE NMWECMQEME GSPLCVIVEF
      DII-SS1 DII-SS2

```

FIG. 7B-2

SUBSTITUTE SHEET (RULE 26)

FIG. 8A-1 Partial Human NaN Nucleotide Sequence

17/28

TCCATTGTCATTGGAATAGCGATTGTGTCATATATCCAGGAATCACCATCAAACTATTGCCCC
TGCGTACCTTCCGTGTGTTGAGAGCTTTGAAAGCAATTCAGTAGTTTCACGTCTGAAGGTCAT
CGTGGGGGCCTTGCTACGCTCTGTGAAGAAGCTGGTCAACGTGATTATCCTCACCTTCTTTTGC
CTCAGCATCTTTGCCCTGGTAGGTGAGCAGCTCTTCATGGGAAGTCTGAACCTGAAATGCATCT
CGAGGGACTGTAAAAATATCAGTAACCCGGAAGCTTATGACCATTGCTTTGAAAAGAAAAGAA
ATTACCTGAATTCAAAATGTGTGGCATCTGGATGGGTAACAGTGCCTGTTCCATACAATATGA
ATGTAAGCACACCAAAATTAATCCTGACTATAATTATACGAATTTTGACAACCTTTGGCTGGTCT
TTTCTTGCCATGTTCCGGCTGATGACCCAAGATTCTGGGAGAAGCTTTATCAACAGACCCTGC
GTACTACTGGGCTCTACTCAGTCTTCTTCTTCATTGTGGTCATTTTCTGGGCTCCTTCTACCTGA
TTAACTTAACCTGGCTGTTGTTACCATGGCATATGAGGAGCAGAACAAGAATGTAGCTGCAG
AGATAGAGGCCAAGGAAAAAGATGTTTCAGGAAGCCCAGCAGCTGTTAAAGGAGGAAAAAGGAG
GCTCTGGTTGCCATGGGAATTGACAGAAGTTCACTTACTTCCCTTGAAAACATCATATTTTACCC
CAAAAAAGAGAAAAGCTCTTTGGTAATAAGAAAAGGAAGTCCTTCTTTTGAGAGAGTCTGGGA
AAGACCAGCCTCCTGGGTCAGATTCTGATGAAGATTGCCAAAAAAAGCCACAGCTCCTAGAGC
AAACCAAAACGACTGTCCAGAACTCTATCAVTGGACCACTTTGATGAGCATGGAGATCCTCTCCA
AAGGCAGAGAGCACTGAGTGCTGTCAGCATCCTCACCATCACCATGAAGGAACAAGAAAAATC
ACAAGAGCCTTGCTCTCCCTGTGGGAGAAAACCTGGCATCCAAGTACCTCGTGTGGAACTGTTGC
CCCCAGTGGCTGTGCGTTAAGAAGGTCCTGAGAACTGTGATGACTGACCCGTTTACTGAGCTGG
CCATCACCATCTGCATCATCATCAACACTGTCTTCTTGGCCATGGAGCATCACAAGATGGAGGC
CAGTTTTGAGAAGATGTTGAATATAGGGAAATTTGGTTTTCACTAGCATTTTTATAGCAGAAATG
TGCCTAAAAATCATTGCGCTCGATCCCTACCACTACTTTCGCCGAGGCTGGAACATTTTTGACA
GCATTGTTGCTCTTCTGAGTTTTGCAGATGTAATGAACTGTGTACTTCAAAAGAGAAGCTGGCC
ATTCTTGCGTTCCCTCAGAGTGCTCAGGGTCTTCAAGTTAGCCAAATCCTGGCCAACTTTGAAC
ACACTAATTAAGATAATCGGCAACTCTGTGCGAGCCCTTGGAAAGCCTGACTGTGGTCTGGTCA
TTGTGATCTTTATTTTCTCAGTAGTTGGCATGCAGCTTTTGGCCGTAGCTTCAATTCCCAAAAG
AGTCCAAAACCTCTGTAACCCGACAGGCCCGACAGTCTCATGTTTACGGCACTGGCAGATGGGG
GATTTCTGGCACTCCTTCTAGTGGTATTCCGCATCCTCTGCGGGGAATGGATCGAAAAATATGT
GGGAATGTATGCAAGAAGCGAATGCATCATCATATTGTGTGTTATTGTCTTCATATTGATCAC
GGTGATAGGAAAACTTGTGGTGCTCAACCTCTTCATTGCCTTACTGCTCAATTCCTTTAGCAAT
GAGGAAAAGAAATGGAACTTAGAAGGAGAGGCCAGGAAAACATAAGTCCAGTTAGCACTGGA
TCGATTCCGCCGGGCTTTTGTGTTTGTGAGACACACTCTTGAGCATTTCTGTACAAAGTGGTGCA
GGAAGCAAACTTACCACAGCAAAAAAGAGGTGGCAGGAGGCTGTGCTGCACAAAGCAAAAGAC
ATCATTCCCCTGGTCATGGAGATGAAAAGGGGCTCAGAGACCCAGGAGGCTTGGTATACTA
ACCTCTGTACCAAAGACCCTGGGCGTCAGGCATGATTGGACTTGGTTGGCACCACTTGCAGGAG
GAGGAAGATGACGTTGAATTTCTGGTGAAGATAATGCACAGCGCATCACACAACCTGAGCCT
GAACAACAGGCCTATGAGCTCCATCAGGAGAACAAGAAGCCCAAGAGCCAGAGAGTTCAAAAG
TGTGGAAATTGACATGTTCTCTGAAGATGAGCCTCATCTGACCATACAGGATCCCCGAAAGAA
GTCTGATGTTACCAGTATACTATCAGAAATGTAGCACCATTGATCTTCAGGATGGCTTTGGATGG
TTACCTGAGATGGTTCCCAAAAGCAACCAGAGAGATGTTTGCCCAAGGCTTTGGTTGCTGCT
TTCCATGCTGTAGCGTGGACAAGAGAAAGCCTCCCTGGGTCAATTGGTGGAACCTGCGGAAAA

18/28

FIG. 8A-2

CCTGCTACCAAATAGTGAAACACAGCTGGTTTGAGAGCTTTATTATCTTTGTGATTCTGCTGAG
CAGTGGGGCACTGATATTTGAAGATGTTACCTTGAGAACCAACCCAAAATCCAAGAATTACT
AAATTGTA CTGACATTATTTTACACATATTTTATCCTGGAGATGGTACTAAAATGGGTAGCC
TTCGGATTTGAAAGTATTTACCAAGTGCCTGGTGCTGCCTTGATTTTCATCATTGTGATTGTCTC
TGTGACCAACCTCATTAACCTAATGGAATTGAAGTCCTTCCGGACTCTACGAGCACTGAGGCCT
CTTCGTGCGCTGTCCCAGTTTGAAGGAATGAAGGTGGTGGTCAATGCTCTCATAGGTGCCATAC
CTGCCATTCTGAATGTTTTGCTTGTCTGCCTCATTTTCTGGCTCGTATTTTGTATTCTGGGAGTAT
ACTTCTTTTCTGGAAAATTTGGGAAATGCATTAATGGAACAGACTCAGTTATAAATTATACCAT
CATTACAAATAAAAAGTCAATGTGAAAGTGGCAATTTCTCTTGATCAACCAGAAAGTCAACTTT
GACAATGTGGGAAATGCTTACCTCGCTCTGCTGCAAGTGGCAACATTTAAGGGCTGGATGGAT
ATTATATATGCAGCTGTTGATTCCACAGAGAAA GAACAACAGCCAGAGTTTGAGAGCAATTCA
CTCGGTTACATTTACTTCGTAGTCTTTATCATCTTTGGCTCATTCTTCACTCTGAATCTCTTCATT
GGCGTTATCATTGACAACCTCAACCAACAGCAGAAAAAGTTAGGTGGCCAAGACATTTTTATG
ACAGAAGAACAGAAGAAATACTATAATGCAATGAAAAAATTAGGATCCAAAAAACCTCAAAA
ACCCATTCCACGGCCTCTGAACAAATGTCAAGGTCTCGTGTTTCGACATAGTCACAAGCCAGATC
TTTGACATCATCATCATAAGTCTCATTATCCTAAACATGATTAGCATGATGGCTGAATCATACA
ACCAACCCAAAGCCATGAAATCCATCCTTGACCATCTCAACTGGGTCTTTGTGGTCATCTTTAC
GTTAGAATGTCTCATCAAAATCTTTGCTTTGAGGCAATACTACTTCACCAATGGCTGGAATTTA
TTTGA

19/28

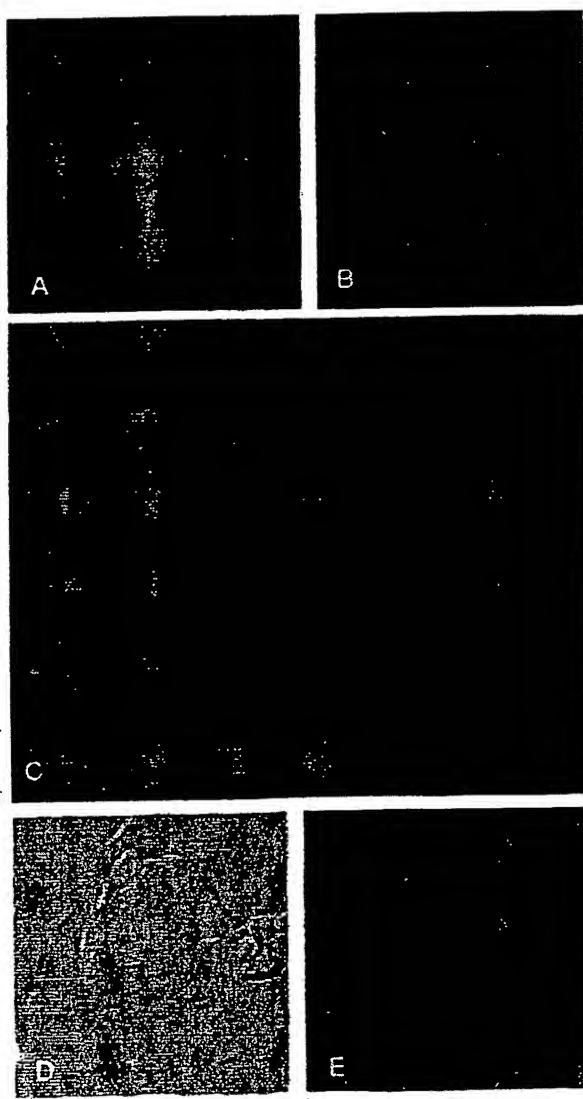
FIG. 8B

Partial Human NaNAmino Acid Sequence

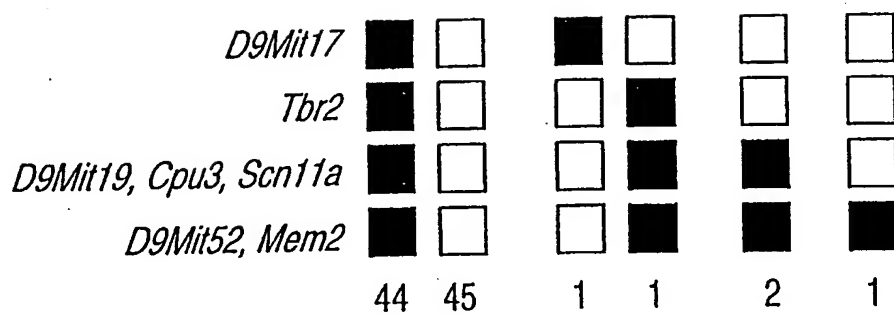
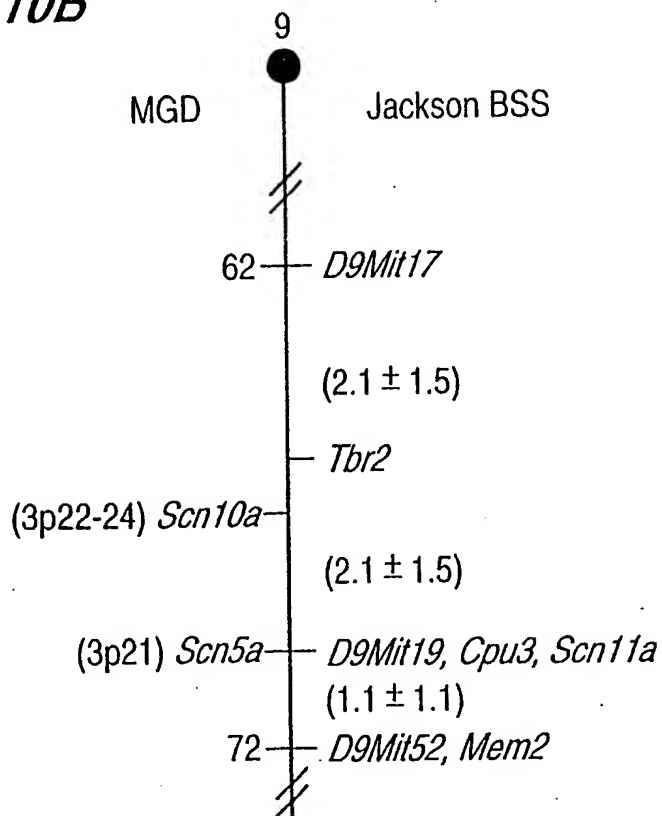
SI VIGIAIVSYI PGITIKLLPL RTFRVFRALK AISVVSRLKV IVGALLRSVK KLVNVITLTF
FCLSIFALVG QQLFMGSLNL KCISRDCKNI SNPEAYDHCF EKKENSPEFK MCGIWMGNSA
CSIQYECKHT KINPDYNYTN FDNFGWSFLA MFRLMTQDSW EKLYQQTLRT TGLYSVFFFI
VVIFLGSFYI INLTAVVTM AYEEQNKNVA AEIEAKEKMF QEAQQLKEE KEALVAMGID
RSSLTSLSTS YFTPKRKLK GNKKRKSFFL RESGKDQPPG SDSDEDCQKK PQLLEQTKRL
SQNLSLDHFD EHGDPQRQR ALSAVSILTI TMKEQEKSE PCLPCGENLA SKYLVWNCCP
QWLCVKKVLR TVMTDPFTEL AITICMINT VFLAMEHHKM EASF EKMLNI GNLVFTSIFI
AEMCLKIAL DPYHYFRGW NIFDSIVALL SFADVMNCVL QKRSWPFLRS
FRVLRVFKLAKSWPTLNTLI KIIGNSVGAL GSLTVVLVTV IFISVVGMMQ LFGRSFNSQK
SPKLCNPTGP TVSCLRHWHM GDFWHSFLVV FRILCGEWIE NMWECMQEAN ASSSLCVTVF
ILITVIGKLV VLNLFIALLL NSFSNEERNG NLEGEARKTK VQLALDRFRR AFCFVRHTLE
HFCHKWCRKQ NLPQQKEVAG GCAAQSKDII PLVMEMKRGS ETQEELGILT SVPKTLGVRH
DWTWLAPLAE EEDDVEFSGE DNAQRITQPE PEQQA YELHQ ENKKPTSQRVQSVEIDMFSE
DEPHLTIQDP RKKSDVTSIL SECSTIDLQD GFGWLPEMVP KKQPERCLPK GFGCCFPCCS
VDKRKPPWVI WWNLRKTCYQ IVKHSWFESF IIFVILLSSG ALIFEDVHLE NQPKIQELLN
CTDIIFTHIF ILEMVLKWVA FGFGKYFTSA WCCLDFIIVI VSVTTLINLM ELKSFRTLRA
LRPLRALSQF EGMKV'VVNAL IGAIPAILNV LLVCLIFWLV FCILGVYFFS GKFGKCTINGT
DSVINYTIIT NKSQCESGNF SWINQKVNFD NVGNAYLALL QVATFKGWMD IYAAVDSTE
KEQQPEFESN SLGYTYFVVF IIFGSFFTLN LFIGVIIDNF NQQQKKLGGQ DIFMTEEQKK
YYNAMKKLGS KKPQKPIPRP LNKCCQLVFD IVTSQIFDII IISLIILNMI SMMAESYNQP
KAMKSILDHL NWVFVVIIFL ECLIKIFALR QYFTNGWNL FDCVVVLLSIV

20/28

FIG. 9
NaN immunostaining in DRG neurons



21/28

FIG. 10A**FIG. 10B**

22/28

FIG. 11A

Sequence of human NaN cDNA. Open reading frame (cdc)
Extends from position 31 (ATG) to the termination codon TGA at
Position 5400.

```
1  ATCTGCTCAA GCCAGGAATC TCGGGTGAAG ATGGATGACA GATGCTACCC
51  AGTAATCTTT CCAGATGAGC GGAATTTCCG CCCCTTCACT TCCGACTCTC
101 TGGCTGCAAT TGAGAAGCGG ATTGCCATCC AAAAGGAGAA AAAGAAGTCT
151 AAAGACCAGA CAGGAGAAGT ACCCCAGCCT CGGCCTCAGC TTGACCTAAA
201 GGCCTCCAGG AAGTTGCCCC AGCTCTATGG CGACATTCCT CGTGAGCTCA
251 TAGGAAAGCC TCTGGAAGAC TTGGACCCAT TCTACCGAAA TCATAAGACA
301 TTTATGGTGT TAAACAGAAA GAGGACAATC TACCGCTTCA GTGCCAAGCA
351 TGCCTTGTTT ATTTTGGGC CTTTCAATTC AATCAGAAGT TTAGCCATTA
401 GAGTCTCAGT CCATTCATTG TTCAGCATGT TCATTATCGG CACCGTTATC
451 ATCAACTGCG TGTTTCATGGC TACAGGGCCT GCTAAAAACA GCAACAGTAA
501 CAATACTGAC ATTGCAGAGT GTGTCTTCAC TGGGATTTAT ATTTTGAAG
551 CTTTGATTAA AATATTGGCA AGAGGTTTCA TTCTGGATGA GTTTTCTTTC
601 CTTGAGATC CATGGAAGT GCTGGACTCC ATTGTCATTG GAATAGCGAT
651 TGTGTCATAT ATTCCAGGAA TCACCATCAA ACTATTGCCC CTGCGTACCT
701 TCCGTGTGTT CAGAGCTTTG AAAGCAATTT CAGTAGTTTC ACGTCTGAAG
751 GTCATCGTGG GGGCCTTGCT ACGCTCTGTG AAGAAGCTGG TCAACGTGAT
801 TATCCTCACC TTCTTTTGCC TCAGCATCTT TGCCCTGGTA GGTCAGCAGC
851 TCTTCATGGG AAGTCTGAAC CTGAAATGCA TCTCGAGGGA CTGTAAAAAT
901 ATCAGTAACC CGGAAGCTTA TGACCATTGC TTTGAAAAGA AAGAAAATTC
951 ACCTGAATTC AAAATGTGTG GCATCTGGAT GGGTAACAGT GCCTGTTCCA
1001 TACAATATGA ATGTAAGCAC ACCAAAATTA ATCCTGACTA TAATTATACG
1051 AATTTTGACA ACTTTGGCTG GTCTTTTCTT GCCATGTTCC GGCTGATGAC
1101 CCAAGATTCC TGGGAGAAGC TTTATCAACA GACCCTGCGT ACTACTGGGC
1151 TCTACTCAGT CTTCTTCTTC ATTGTGGTCA TTTTCTGGG CTCCTTCTAC
1201 CTGATTAAC TAACCCTGGC TGTTGTTACC ATGGCATATG AGGAGCAGAA
```


FIG. 11A-2 23/28

1251 CAAGAATGTA GCTGCAGAGA TAGAGGCCAA GGAAAAGATG TTTCAGGAAG
1301 CCCAGCAGCT GTTAAAGGAG GAAAAGGAGG CTCTGGTTGC CATGGGAATT
1351 GACAGAAGTT CACTTACTTC CCTTGAAACA TCATATTTTA CCCCCAAAAA
1401 GAGAAAGCTC TTTGGTAATA AGAAAAGGAA GTCCTTCTTT TTGAGAGAGT
1451 CTGGGAAAGA CCAGCCTCCT GGGTCAGATT CTGATGAAGA TTGCCAAAAA
1501 AAGCCACAGC TCCTAGAGCA AACCAAACGA CTGTCCCAGA ATCTATCACT
1551 GGACCACTTT GATGAGCATG GAGATCCTCT CCAAAGGCAG AGAGCACTGA
1601 GTGCTGTCAG CATCCTCACC ATCACCATGA AGGAACAAGA AAAATCACAA
1651 GAGCCTTGTC TCCCTTGTGG AGAAAACCTG GCATCCAAGT ACCTCGTGTG
1701 GAACTGTTGC CCCAGTGGC TGTGCGTTAA GAAGGTCCTG AGAACTGTGA
1751 TGA CTGACCC GTTTACTGAG CTGGCCATCA CCATCTGCAT CATCATCAAC
1801 ACTGTCTTCT TGGCCATGGA GCATGACAAG ATGGAGGCCA GTTTTGAGAA
1851 GATGTTGAAT ATAGGGAATT TGGTTTTTAC TAGCATTTTT ATAGCAGAAA
1901 TGTGCCTAAA AATCATTGCG CTCGATCCCT ACCACTACTT TCGCCGAGGC
1951 TGGAACATTT TTGACAGCAT TGTGCTCTT CTGAGTTTTG CAGATGTAAT
2001 GAACTGTGTA CTTCAAAAGA GAAGCTGGCC ATTCTTGCGT TCCTTCAGAG
2051 TGCTCAGGGT CTTCAAGTTA GCCAAATCCT GGCCAACTTT GAACACACTA
2101 ATTAAGATAA TCGGCAACTC TGTCGGAGCC CTTGGAAGCC TGACTGTGGT
2151 CCTGGTCATT GTGATCTTTA TTTTCTCAGT AGTTGGCATG CAGCTTTTTG
2201 GCCGTAGCTT CAATTCCCAA AAGAGTCCAA AACTCTGTAA CCCGACAGGC
2251 CCGACAGTCT CATGTTTACG GCACTGGCAC ATGGGGGATT TCTGGCACTC
2301 CTTCTAGTG GTATTCCGCA TCCTCTGCGG GGAATGGATC GAAAATATGT
2351 GGAATGTAT GCAAGAAGCG AATGCATCAT CATCATTGTG TGTTATTGTC
2401 TTCATATTGA TCACGGTGAT AGGAAAACCT GTGGTGCTCA ACCTCTTCAT
2451 TGCCTTACTG CTCAATTCCT TTAGCAATGA GGAAAGAAAT GGAAACTTAG
2501 AAGGAGAGGC CAGGAAAACCT AAAGTCCAGT TAGCACTGGA TCGATTCCGC
2551 CGGGCTTTTT GTTTGTGAG ACACACTCTT GAGCATTTCT GTCACAAGTG

24/28

FIG. 11A-3

2601 GTGCAGGAAG CAAAACTTAC CACAGCAAAA AGAGGTGGCA GGAGGCTGTG
2651 CTGCACAAAG CAAAGACATC ATTCCCCTGG TCATGGAGAT GAAAAGGGGC
2701 TCAGAGACCC AGGAGGAGCT TGGTATACTA ACCTCTGTAC CAAAGACCCT
2751 GGGCGTCAGG CATGATTGGA CTTGGTTGGC ACCACTTGCG GAGGAGGAAG
2801 ATGACGTTGA ATTTTCTGGT GAAGATAATG CACAGCGCAT CACACAACCT
2851 GAGCCTGAAC AACAGGCCTA TGAGCTCCAT CAGGAGAACA AGAAGCCCAC
2901 GAGCCAGAGA GTTCAAAGTG TGGAAATTGA CATGTTCTCT GAAGATGAGC
2951 CTCATCTGAC CATACAGGAT CCCCAGAAAG AGTCTGATGT TACCAGTATA
3001 CTATCAGAAT GTAGCACCAT TGATCTTCAG GATGGCTTTG GATGGTTACC
3051 TGAGATGGTT CCCAAAAAGC AACCAGAGAG ATGTTTGCCC AAAGGCTTTG
3101 GTTGCTGCTT TCCATGCTGT AGCGTGGACA AGAGAAAGCC TCCCTGGGTC
3151 ATTTGGTGGA ACCTGCGGAA AACCTGCTAC CAAATAGTGA AACACAGCTG
3201 GTTTGAGAGC TTTATTATCT TTGTGATTCT GCTGAGCAGT GGGGCACTGA
3251 TATTTGAAGA TGTTACCTT GAGAACCAAC CCAAAATCCA AGAATTACTA
3301 AATTGTACTG ACATTATTTT TACACATATT TTTATCCTGG AGATGGTACT
3351 AAAATGGGTA GCCTTCGGAT TTGGAAAGTA TTTACCAGT GCCTGGTGCT
3401 GCCTTGATTT CATCATTGTG ATTGTCTCTG TGACCACCCT CATTAACTTA
3451 ATGGAATTGA AGTCCTCCG GACTCTACGA GCACTGAGGC CTCTTCGTGC
3501 GCTGTCCCAG TTTGAAGGAA TGAAGGTGGT GGTCAATGCT CTCATAGGTG
3551 CCATACCTGC CATTCTGAAT GTTTTGCTTG TCTGCCTCAT TTTCTGGCTC
3601 GTATTTTGTA TTCTGGGAGT ATACTTCTTT TCTGGAAAAT TTGGGAAATG
3651 CATTAAATGA ACAGACTCAG TTATAAATTA TACCATCATT ACAAATAAAA
3701 GTCAATGTGA AAGTGGCAAT TTCTCTTGA TCAACCAGAA AGTCAACTTT
3751 GACAATGTGG GAAATGCTTA CCTCGCTCTG CTGCAAGTGG CAACATTTAA
3801 GGGCTGGATG GATATTATAT ATGCAGCTGT TGATTCCACA GAGAAAGAAC
3851 AACAGCCAGA GTTTGAGAGC AATTCACCTG GTTACATTTA CTTCGTAGTC
3901 TTTATCATCT TTGGCTCATT CTTCACTCTG AATCTCTTCA TTGGCGTTAT
3951 CATTGACAAC TTCAACCAAC AGCAGAAAAA GTTAGGTGGC CAAGACATTT

25/28
FIG. 11A-4

4001 TTATGACAGA AGAACAGAAG AAATACTATA ATGCAATGAA AAAATTAGGA
4051 TCCAAAAAAC CTCAAAAACC CATTCCACGG CCTCTGAACA AATGTCAAGG
4101 TCTCGTGTTT GACATAGTCA CAAGCCAGAT CTTTGACATC ATCATCATAA
4151 GTCTCATTAT CCTAAACATG ATTAGCATGA TGGCTGAATC ATACAACCAA
4201 CCCAAAGCCA TGAAATCCAT CCTTGACCAT CTCAACTGGG TCTTTGTGGT
4251 CATCTTTACG TTAGAATGTC TCATCAAAAT CTTTGCTTTG AGGCAATACT
4301 ACTTCACCAA TGGCTGGAAT TTATTTGACT GTGTGGTCGT GCTTCTTTCC
4351 ATTGTTAGTA CAATGATTTC TACCTTGGAA AATCAGGAGC ACATTCTTTT
4401 CCCTCCGACG CTCTTCAGAA TTGTCCGCTT GGCTCGGATT GGCCGAATCC
4451 TGAGGCTTGT CCGGGCTGCA CGAGGAATCA GGACTCTCCT CTTTGCTCTG
4501 ATGATGTCGC TTCCTTCTCT GTTCAACATT GGTCTTCTAC TCTTTCTGAT
4551 TATGTTTATC TATGCCATTG TGGGTATGAA CTGGTTTTCC AAAGTGAATC
4601 CAGAGTCTGG AATCGATGAC ATATTCAACT TCAAGACTTT TGCCAGCAGC
4651 ATGCTCTGTC TCTTCCAGAT AAGCACATCA GCAGGTTGGG ATTCCCTGCT
4701 CAGCCCCATG CTGCGATCAA AAGAATCATG TAACTCTTCC TCAGAAAACCT
4751 GCCACCTCCC TGGCATAGCC ACATCCTACT TTGTCAGTTA CATTATCATC
4801 TCCTTTCTCA TTGTTGTCAA CATGTACATT GCTGTGATTT TAGAGAACTT
4851 CAATACAGCC ACTGAAGAAA GTGAGGACCC TTTGGGTGAA GATGACTTTG
4901 ACATATTTTA TGAAGTGTGG GAAAAGTTTG ACCCAGAAGC AACACAATTT
4951 ATCAAATATT CTGCCCTTTC TGACTTTGCT GATGCCTTGC CTGAGCCTTT
5001 GCGTGTCGCA AAGCCAAATA AATATCAATT TCTAGTAATG GACTTGCCCA
5051 TGGTGAGTGA AGATCGCCTC CACTGCATGG ATATTCTTTT CGCCTTCACC
5101 GCTAGGGTAC TCGGTGGCTC TGATGGCCTA GATAGTATGA AAGCAATGAT
5151 GGAAGAGAAG TTCATGGAAG CCAATCCTCT CAAGAAGTTG TATGAACCCA
5201 TAGTCACCAC CACCAAGAGA AAGGAAGAGG AAAGAGGTGC TGCTATTATT
5251 CAAAAGGCCT TTCGAAAGTA CATGATGAAG GTGACCAAGG GTGACCAAGG
5301 TGACCAAAAT GACTTGGAAG ACGGGCCTCA TTCACCACTC CAGACTCTTT

FIG. 11A-5

5351 GCAATGGAGA CTTGTCTAGC TTTGGGGTGG CCAAGGGCAA GGTCCACTGT
5401 GACTGAGCCC TCACCTCCAC GCCTACCTCA TAGCTTCACA GCCTTGCCTT
5451 CAGCCTCTGA GCTCCAGGGG TCAGCAGCTT AGTGTATCAA CAGGGAGTGG
5501 ATTCACCAAA TTAGCCATTC CATTTTCTTT TCTGGCTAAA ATAAATGATA
5551 TTTCAATTTC ATTTTAAATG ATACTTACAG AGATATAAGA TAAGGCTACT
5601 TGACAACCAG TGGTACTATT ATAATAAGGA AGAAGACACC AGGAAGGACT
5651 GTAAAAGGAC ATACCAATTT TAGGATTGAA ATAGTTCAGG CCGGGCGCAG
5701 TGGCTCATGC CTGTAATCCC AGCACTTTGA GAGGCCAAGG CAGGTGGATC
5751 ACGAGGTCAA GAGATCGAGA CCATCCTGGC CAACATGATG AAACTCCGTC
5801 TCTCTAAAAA TACAAAAATT AGCTGGGCAT GGTGGCGTGC GCCTGTAGTC
5851 CCACTACTTG

27/28

FIG. 11B

Sequence of human NaN.

1 MDDRCYPVIF PDERNFRPFT SDSLAAIEKR IAIQKEKKKS KDQTGEVPQP
51 RPQLDLKASR KLPKLYGDIP RELIGKPLED LDPFYRNHKT FMVLNRKRTI
101 YRFSAKHALF IFGPFNSIRS LAIRSVHSL FSMFIIGTVI INCVFMTATGP
151 AKSNSNNNTD IAECVFTGIY IFEALIKILA RGFILDEFSS LRDPWNWLDS
201 IVIGIAIVSY IPGITIKLLP LRTFRVFRAL KAISVVSRLK VIVGALLRSV
251 KKLNVNVIILT FFCLSIFALV GQQLFMGSLN LKCISRDCKN ISNPEAYDHC
301 FEKKENSPEF KMCGIWMGNS ACSIQYECKH TKINPDYNYT NFDNFGWSFL
351 AMFRLMTQDS WEKLYQOTLR TTGLYSVFFF IVVIFLGSFY LINLTLAVVT
401 MAYEEQNKNV AAEIEAKEKM FQEAQQLKE EKEALVAMGI DRSSLTSLET
451 SYFTPKKRKL FGNKKRKSFF LRESGKDQPP GSDSDEDCQK KPQLLEQTKR
501 LSQNLSLDHF DEHGDPLQRQ RALSAVSILT ITMKEQESQ EPCLPCGENL
551 ASKYLWNCC PQWLCVKKVL RTVMTDPFTE LAITICIIIN TVFLAMEHHK
601 MEASF EKMLN IGNLVFTSIF IAEMCLKIIA LDPYHYFRRG WNIFDSIVAL
651 LSFADVMNCV LQKRSWPFLR SFRVLRVFKL AKSWPTLNTL IKIIGNSVGA
701 LGSLTVVLVI VIFIFSVVGM QLFGRSFNSQ KSPKLCNPTG PTVSCLRHHW
751 MGDFWHSFLV VFRILCGEWI ENMWEQMSEA NASSSLCVIV FILITVIGKL
801 VVLNLFIALL LNSFSNEERN GNLEGEARKT KVQLALDRFR RAFCFVRHTL
851 EHFCHKWCRK QNLPQQKEVA GGCAAQSKDI IPLVMEMKRG SETQEELGIL
901 TSVPKTLGVR HDWTWLAPLA EEEDDVEFSG EDNAQRITQP EPEQQAYELH
951 QENKKPTSQR VQSVEIDMFS EDEPHLTIQD PRKKS DVTSI LSECSTIDLQ
1001 DGFGWLPEMV PKKQPERCLP KGFGCCFPCC SVDKRKPPWV IWWNLRKTCY
1051 QIVKHSWFES FIIFVILLSS GALIFEDVHL ENQPKIQELL NCTDIIIFTHI
1101 FILEMVLKWW AFGFGKYFTS AWCLDFIIV IVSVTTLINL MELKSFRTLR
1151 ALRPLRALSQ FEGMKVVVNA LIGAIPAILN VLLVCLIFWL VFCILGVYFF

FIG. 11B-2

1201 SGKFGKACING TDSVINYTII TNKSQCESGN FSWINQKVN F DNVGNAYLAL
1251 LQVATFKGWM DIIYAAVDST EKEQQPEFES NSLGYIYFVV FIIFGSFFTL
1301 NLFIGVIIDN FNQQQKKLGG QDIFMTEEQK KYYNAMKKLG SKKPQKPIPR
1351 PLNKCQGLVF DIVTSQIFDI IIISLIILNM ISMMAESYNQ PKAMKSILDH
1401 LNWVFWVIFT LECLIKIFAL RQYYFTNGWN LFDCVVVLLS IVSTMISTLE
1451 NQEHIPFPPT LFRIVRLARI GRILRLVRAA RGIRTLLFAL MMSLPSLFNI
1501 GLLLFLIMFI YAILGMNWFS KVPESGIDD IFNFKTFASS MLCLFQISTS
1551 AGWDSLSPM LRSKESCNSS SENCHLPGIA TSYFVSYIII SFLIVVMYI
1601 AVILENFNTA TEESEDPLGE DDFDIFYEVW EKFDPEATQF IKYSALSDFA
1651 DALPEPLRVA KPNKYQFLVM DLPMVSEDRL HCM DILFAFT ARVLGGS DGL
1701 DSMKAMMEK FMEANPLKKL YEPIVTTTKR KEEERGAAII QKA FRKYMMK
1751 VTKGDQGDQN DLENGPHSPL QTL CNGDLSS FGVAKGVHC D.

<400> 38
Met Trp Xaa Cys Met Glu Val
1 5

<210> 39
<211> 22
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: rat NaN
forward primer

<400> 39
ccctgctgcg ctcggtgaag aa 22

<210> 40
<211> 20
<212> PRT
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: a.a. seq.
used to derive epitope for polyclonal antibody

<400> 40
Cys Gly Pro Asn Pro Ala Ser Asn Lys Asp Cys Phe Glu Lys Glu Lys
1 5 10 15

Asp Ser Glu Asp
20

<210> 41
<211> 5860
<212> DNA
<213> Homo sapiens

<220>
<221> CDS
<222> (31)..(5403)
<223> full length cDNA sequence for human NaN

<400> 41
atctgctcaa gccaggaatc tcgggtgaag atg gat gac aga tgc tac cca gta 54
Met Asp Asp Arg Cys Tyr Pro Val
1 5

atc ttt cca gat gag cgg aat ttc cgc ccc ttc act tcc gac tct ctg 102
Ile Phe Pro Asp Glu Arg Asn Phe Arg Pro Phe Thr Ser Asp Ser Leu
10 15 20

gct gca att gag aag cgg att gcc atc caa aag gag aaa aag aag tct 150
Ala Ala Ile Glu Lys Arg Ile Ala Ile Gln Lys Glu Lys Lys Lys Ser

25	30	35	40	
aaa gac cag aca gga gaa gta ccc cag cct cgg cct cag ctt gac cta				198
Lys Asp Gln Thr Gly Glu Val Pro Gln Pro Arg Pro Gln Leu Asp Leu	45	50	55	
aag gcc tcc agg aag ttg ccc aag ctc tat ggc gac att cct cgt gag				246
Lys Ala Ser Arg Lys Leu Pro Lys Leu Tyr Gly Asp Ile Pro Arg Glu	60	65	70	
ctc ata gga aag cct ctg gaa gac ttg gac cca ttc tac cga aat cat				294
Leu Ile Gly Lys Pro Leu Glu Asp Leu Asp Pro Phe Tyr Arg Asn His	75	80	85	
aag aca ttt atg gtg tta aac aga aag agg aca atc tac cgc ttc agt				342
Lys Thr Phe Met Val Leu Asn Arg Lys Arg Thr Ile Tyr Arg Phe Ser	90	95	100	
gcc aag cat gcc ttg ttc att ttt ggg cct ttc aat tca atc aga agt				390
Ala Lys His Ala Leu Phe Ile Phe Gly Pro Phe Asn Ser Ile Arg Ser	105	110	115	120
tta gcc att aga gtc tca gtc cat tca ttg ttc agc atg ttc att atc				438
Leu Ala Ile Arg Val Ser Val His Ser Leu Phe Ser Met Phe Ile Ile	125	130	135	
ggc acc gtt atc atc aac tgc gtg ttc atg gct aca ggg cct gct aaa				486
Gly Thr Val Ile Ile Asn Cys Val Phe Met Ala Thr Gly Pro Ala Lys	140	145	150	
aac agc aac agt aac aat act gac att gca gag tgt gtc ttc act ggg				534
Asn Ser Asn Ser Asn Asn Thr Asp Ile Ala Glu Cys Val Phe Thr Gly	155	160	165	
att tat att ttt gaa gct ttg att aaa ata ttg gca aga ggt ttc att				582
Ile Tyr Ile Phe Glu Ala Leu Ile Lys Ile Leu Ala Arg Gly Phe Ile	170	175	180	
ctg gat gag ttt tct ttc ctt cga gat cca tgg aac tgg ctg gac tcc				630
Leu Asp Glu Phe Ser Phe Leu Arg Asp Pro Trp Asn Trp Leu Asp Ser	185	190	195	200
att gtc att gga ata gcg att gtg tca tat att cca gga atc acc atc				678
Ile Val Ile Gly Ile Ala Ile Val Ser Tyr Ile Pro Gly Ile Thr Ile	205	210	215	
aaa cta ttg ccc ctg cgt acc ttc cgt gtg ttc aga gct ttg aaa gca				726
Lys Leu Leu Pro Leu Arg Thr Phe Arg Val Phe Arg Ala Leu Lys Ala	220	225	230	
att tca gta gtt tca cgt ctg aag gtc atc gtg ggg gcc ttg cta cgc				774
Ile Ser Val Val Ser Arg Leu Lys Val Ile Val Gly Ala Leu Leu Arg	235	240	245	
tct gtg aag aag ctg gtc aac gtg att atc ctc acc ttc ttt tgc ctc				822
Ser Val Lys Lys Leu Val Asn Val Ile Ile Leu Thr Phe Phe Cys Leu				

250	255	260	
agc atc ttt gcc ctg gta ggt cag cag ctc ttc atg gga agt ctg aac			870
Ser Ile Phe Ala Leu Val Gly Gln Gln Leu Phe Met Gly Ser Leu Asn			
265	270	275	280
ctg aaa tgc atc tcg agg gac tgt aaa aat atc agt aac ccg gaa gct			918
Leu Lys Cys Ile Ser Arg Asp Cys Lys Asn Ile Ser Asn Pro Glu Ala			
	285	290	295
tat gac cat tgc ttt gaa aag aaa gaa aat tca cct gaa ttc aaa atg			966
Tyr Asp His Cys Phe Glu Lys Lys Glu Asn Ser Pro Glu Phe Lys Met			
	300	305	310
tgt ggc atc tgg atg ggt aac agt gcc tgt tcc ata caa tat gaa tgt			1014
Cys Gly Ile Trp Met Gly Asn Ser Ala Cys Ser Ile Gln Tyr Glu Cys			
	315	320	325
aag cac acc aaa att aat cct gac tat aat tat acg aat ttt gac aac			1062
Lys His Thr Lys Ile Asn Pro Asp Tyr Asn Tyr Thr Asn Phe Asp Asn			
	330	335	340
ttt ggc tgg tct ttt ctt gcc atg ttc cgg ctg atg acc caa gat tcc			1110
Phe Gly Trp Ser Phe Leu Ala Met Phe Arg Leu Met Thr Gln Asp Ser			
	345	350	355
tgg gag aag ctt tat caa cag acc ctg cgt act act ggg ctc tac tca			1158
Trp Glu Lys Leu Tyr Gln Gln Thr Leu Arg Thr Thr Gly Leu Tyr Ser			
	365	370	375
gtc ttc ttc ttc att gtg gtc att ttc ctg ggc tcc ttc tac ctg att			1206
Val Phe Phe Phe Ile Val Val Ile Phe Leu Gly Ser Phe Tyr Leu Ile			
	380	385	390
aac tta acc ctg gct gtt gtt acc atg gca tat gag gag cag aac aag			1254
Asn Leu Thr Leu Ala Val Val Thr Met Ala Tyr Glu Glu Gln Asn Lys			
	395	400	405
aat gta gct gca gag ata gag gcc aag gaa aag atg ttt cag gaa gcc			1302
Asn Val Ala Ala Glu Ile Glu Ala Lys Glu Lys Met Phe Gln Glu Ala			
	410	415	420
cag cag ctg tta aag gag gaa aag gag gct ctg gtt gcc atg gga att			1350
Gln Gln Leu Leu Lys Glu Glu Lys Glu Ala Leu Val Ala Met Gly Ile			
	425	430	435
gac aga agt tca ctt act tcc ctt gaa aca tca tat ttt acc cca aaa			1398
Asp Arg Ser Ser Leu Thr Ser Leu Glu Thr Ser Tyr Phe Thr Pro Lys			
	445	450	455
aag aga aag ctc ttt ggt aat aag aaa agg aag tcc ttc ttt ttg aga			1446
Lys Arg Lys Leu Phe Gly Asn Lys Lys Arg Lys Ser Phe Phe Leu Arg			
	460	465	470
gag tct ggg aaa gac cag cct cct ggg tca gat tct gat gaa gat tgc			1494
Glu Ser Gly Lys Asp Gln Pro Pro Gly Ser Asp Ser Asp Glu Asp Cys			

475	480	485	
caa aaa aag cca cag ctc cta gag caa acc aaa cga ctg tcc cag aat			1542
Gln Lys Lys Pro Gln Leu Leu Glu Gln Thr Lys Arg Leu Ser Gln Asn			
490	495	500	
cta tca ctg gac cac ttt gat gag cat gga gat cct ctc caa agg cag			1590
Leu Ser Leu Asp His Phe Asp Glu His Gly Asp Pro Leu Gln Arg Gln			
505	510	515	520
aga gca ctg agt gct gtc agc atc ctc acc atc acc atg aag gaa caa			1638
Arg Ala Leu Ser Ala Val Ser Ile Leu Thr Ile Thr Met Lys Glu Gln			
	525	530	535
gaa aaa tca caa gag cct tgt ctc cct tgt gga gaa aac ctg gca tcc			1686
Glu Lys Ser Gln Glu Pro Cys Leu Pro Cys Gly Glu Asn Leu Ala Ser			
	540	545	550
aag tac ctc gtg tgg aac tgt tgc ccc cag tgg ctg tgc gtt aag aag			1734
Lys Tyr Leu Val Trp Asn Cys Cys Pro Gln Trp Leu Cys Val Lys Lys			
	555	560	565
gtc ctg aga act gtg atg act gac ccg ttt act gag ctg gcc atc acc			1782
Val Leu Arg Thr Val Met Thr Asp Pro Phe Thr Glu Leu Ala Ile Thr			
	570	575	580
atc tgc atc atc atc aac act gtc ttc ttg gcc atg gag cat cac aag			1830
Ile Cys Ile Ile Ile Asn Thr Val Phe Leu Ala Met Glu His His Lys			
	585	590	595
atg gag gcc agt ttt gag aag atg ttg aat ata ggg aat ttg gtt ttc			1878
Met Glu Ala Ser Phe Glu Lys Met Leu Asn Ile Gly Asn Leu Val Phe			
	605	610	615
act agc att ttt ata gca gaa atg tgc cta aaa atc att gcg ctc gat			1926
Thr Ser Ile Phe Ile Ala Glu Met Cys Leu Lys Ile Ile Ala Leu Asp			
	620	625	630
ccc tac cac tac ttt cgc cga ggc tgg aac att ttt gac agc att gtt			1974
Pro Tyr His Tyr Phe Arg Arg Gly Trp Asn Ile Phe Asp Ser Ile Val			
	635	640	645
gct ctt ctg agt ttt gca gat gta atg aac tgt gta ctt caa aag aga			2022
Ala Leu Leu Ser Phe Ala Asp Val Met Asn Cys Val Leu Gln Lys Arg			
	650	655	660
agc tgg cca ttc ttg cgt tcc ttc aga gtg ctc agg gtc ttc aag tta			2070
Ser Trp Pro Phe Leu Arg Ser Phe Arg Val Leu Arg Val Phe Lys Leu			
	665	670	675
gcc aaa tcc tgg cca act ttg aac aca cta att aag ata atc ggc aac			2118
Ala Lys Ser Trp Pro Thr Leu Asn Thr Leu Ile Lys Ile Ile Gly Asn			
	685	690	695
tct gtc gga gcc ctt gga agc ctg act gtg gtc ctg gtc att gtg atc			2166
Ser Val Gly Ala Leu Gly Ser Leu Thr Val Val Leu Val Ile Val Ile			

700	705	710	
ttt att ttc tca gta gtt ggc atg cag ctt ttt ggc cgt agc ttc aat			2214
Phe Ile Phe Ser Val Val Gly Met Gln Leu Phe Gly Arg Ser Phe Asn			
715	720	725	
tcc caa aag agt cca aaa ctc tgt aac ccg aca ggc ccg aca gtc tca			2262
Ser Gln Lys Ser Pro Lys Leu Cys Asn Pro Thr Gly Pro Thr Val Ser			
730	735	740	
tgt tta cgg cac tgg cac atg ggg gat ttc tgg cac tcc ttc cta gtg			2310
Cys Leu Arg His Trp His Met Gly Asp Phe Trp His Ser Phe Leu Val			
745	750	755	760
gta ttc cgc atc ctc tgc ggg gaa tgg atc gaa aat atg tgg gaa tgt			2358
Val Phe Arg Ile Leu Cys Gly Glu Trp Ile Glu Asn Met Trp Glu Cys			
765	770	775	
atg caa gaa gcg aat gca tca tca tca ttg tgt gtt att gtc ttc ata			2406
Met Gln Glu Ala Asn Ala Ser Ser Ser Leu Cys Val Ile Val Phe Ile			
780	785	790	
ttg atc acg gtg ata gga aaa ctt gtg gtg ctc aac ctc ttc att gcc			2454
Leu Ile Thr Val Ile Gly Lys Leu Val Val Leu Asn Leu Phe Ile Ala			
795	800	805	
tta ctg ctc aat tcc ttt agc aat gag gaa aga aat gga aac tta gaa			2502
Leu Leu Leu Asn Ser Phe Ser Asn Glu Glu Arg Asn Gly Asn Leu Glu			
810	815	820	
gga gag gcc agg aaa act aaa gtc cag tta gca ctg gat cga ttc cgc			2550
Gly Glu Ala Arg Lys Thr Lys Val Gln Leu Ala Leu Asp Arg Phe Arg			
825	830	835	840
cgg gct ttt tgt ttt gtg aga cac act ctt gag cat ttc tgt cac aag			2598
Arg Ala Phe Cys Phe Val Arg His Thr Leu Glu His Phe Cys His Lys			
845	850	855	
tgg tgc agg aag caa aac tta cca cag caa aaa gag gtg gca gga ggc			2646
Trp Cys Arg Lys Gln Asn Leu Pro Gln Gln Lys Glu Val Ala Gly Gly			
860	865	870	
tgt gct gca caa agc aaa gac atc att ccc ctg gtc atg gag atg aaa			2694
Cys Ala Ala Gln Ser Lys Asp Ile Ile Pro Leu Val Met Glu Met Lys			
875	880	885	
agg ggc tca gag acc cag gag gag ctt ggt ata cta acc tct gta cca			2742
Arg Gly Ser Glu Thr Gln Glu Glu Leu Gly Ile Leu Thr Ser Val Pro			
890	895	900	
aag acc ctg ggc gtc agg cat gat tgg act tgg ttg gca cca ctt gcg			2790
Lys Thr Leu Gly Val Arg His Asp Trp Thr Trp Leu Ala Pro Leu Ala			
905	910	915	920
gag gag gaa gat gac gtt gaa ttt tct ggt gaa gat aat gca cag cgc			2838
Glu Glu Glu Asp Asp Val Glu Phe Ser Gly Glu Asp Asn Ala Gln Arg			

	925	930	935	
atc aca caa cct gag cct gaa caa cag gcc tat gag ctc cat cag gag				2886
Ile Thr Gln Pro Glu Pro Glu Gln Gln Ala Tyr Glu Leu His Gln Glu				
	940	945	950	
aac aag aag ccc acg agc cag aga gtt caa agt gtg gaa att gac atg				2934
Asn Lys Lys Pro Thr Ser Gln Arg Val Gln Ser Val Glu Ile Asp Met				
	955	960	965	
ttc tct gaa gat gag cct cat ctg acc ata cag gat ccc cga aag aag				2982
Phe Ser Glu Asp Glu Pro His Leu Thr Ile Gln Asp Pro Arg Lys Lys				
	970	975	980	
tct gat gtt acc agt ata cta tca gaa tgt agc acc att gat ctt cag				3030
Ser Asp Val Thr Ser Ile Leu Ser Glu Cys Ser Thr Ile Asp Leu Gln				
	985	990	995	1000
gat ggc ttt gga tgg tta cct gag atg gtt ccc aaa aag caa cca gag				3078
Asp Gly Phe Gly Trp Leu Pro Glu Met Val Pro Lys Lys Gln Pro Glu				
	1005	1010	1015	
aga tgt ttg ccc aaa ggc ttt ggt tgc tgc ttt cca tgc tgt agc gtg				3126
Arg Cys Leu Pro Lys Gly Phe Gly Cys Cys Phe Pro Cys Cys Ser Val				
	1020	1025	1030	
gac aag aga aag cct ccc tgg gtc att tgg tgg aac ctg cgg aaa acc				3174
Asp Lys Arg Lys Pro Pro Trp Val Ile Trp Trp Asn Leu Arg Lys Thr				
	1035	1040	1045	
tgc tac caa ata gtg aaa cac agc tgg ttt gag agc ttt att atc ttt				3222
Cys Tyr Gln Ile Val Lys His Ser Trp Phe Glu Ser Phe Ile Ile Phe				
	1050	1055	1060	
gtg att ctg ctg agc agt ggg gca ctg ata ttt gaa gat gtt cac ctt				3270
Val Ile Leu Leu Ser Ser Gly Ala Leu Ile Phe Glu Asp Val His Leu				
	1065	1070	1075	1080
gag aac caa ccc aaa atc caa gaa tta cta aat tgt act gac att att				3318
Glu Asn Gln Pro Lys Ile Gln Glu Leu Leu Asn Cys Thr Asp Ile Ile				
	1085	1090	1095	
ttt aca cat att ttt atc ctg gag atg gta cta aaa tgg gta gcc ttc				3366
Phe Thr His Ile Phe Ile Leu Glu Met Val Leu Lys Trp Val Ala Phe				
	1100	1105	1110	
gga ttt gga aag tat ttc acc agt gcc tgg tgc tgc ctt gat ttc atc				3414
Gly Phe Gly Lys Tyr Phe Thr Ser Ala Trp Cys Cys Leu Asp Phe Ile				
	1115	1120	1125	
att gtg att gtc tct gtg acc acc ctc att aac tta atg gaa ttg aag				3462
Ile Val Ile Val Ser Val Thr Thr Leu Ile Asn Leu Met Glu Leu Lys				
	1130	1135	1140	
tcc ttc cgg act cta cga gca ctg agg cct ctt cgt gcg ctg tcc cag				3510
Ser Phe Arg Thr Leu Arg Ala Leu Arg Pro Leu Arg Ala Leu Ser Gln				

1145	1150	1155	1160	
ttt gaa gga atg aag gtg gtg gtc aat gct ctc ata ggt gcc ata cct				3558
Phe Glu Gly Met Lys Val Val Val Asn Ala Leu Ile Gly Ala Ile Pro				
	1165	1170	1175	
gcc att ctg aat gtt ttg ctt gtc tgc ctc att ttc tgg ctc gta ttt				3606
Ala Ile Leu Asn Val Leu Leu Val Cys Leu Ile Phe Trp Leu Val Phe				
	1180	1185	1190	
tgt att ctg gga gta tac ttc ttt tct gga aaa ttt ggg aaa tgc att				3654
Cys Ile Leu Gly Val Tyr Phe Phe Ser Gly Lys Phe Gly Lys Cys Ile				
	1195	1200	1205	
aat gga aca gac tca gtt ata aat tat acc atc att aca aat aaa agt				3702
Asn Gly Thr Asp Ser Val Ile Asn Tyr Thr Ile Ile Thr Asn Lys Ser				
	1210	1215	1220	
caa tgt gaa agt ggc aat ttc tct tgg atc aac cag aaa gtc aac ttt				3750
Gln Cys Glu Ser Gly Asn Phe Ser Trp Ile Asn Gln Lys Val Asn Phe				
	1225	1230	1235	1240
gac aat gtg gga aat gct tac ctc gct ctg ctg caa gtg gca aca ttt				3798
Asp Asn Val Gly Asn Ala Tyr Leu Ala Leu Leu Gln Val Ala Thr Phe				
	1245	1250	1255	
aag ggc tgg atg gat att ata tat gca gct gtt gat tcc aca gag aaa				3846
Lys Gly Trp Met Asp Ile Ile Tyr Ala Ala Val Asp Ser Thr Glu Lys				
	1260	1265	1270	
gaa caa cag cca gag ttt gag agc aat tca ctc ggt tac att tac ttc				3894
Glu Gln Gln Pro Glu Phe Glu Ser Asn Ser Leu Gly Tyr Ile Tyr Phe				
	1275	1280	1285	
gta gtc ttt atc atc ttt ggc tca ttc ttc act ctg aat ctc ttc att				3942
Val Val Phe Ile Ile Phe Gly Ser Phe Phe Thr Leu Asn Leu Phe Ile				
	1290	1295	1300	
ggc gtt atc att gac aac ttc aac caa cag cag aaa aag tta ggt ggc				3990
Gly Val Ile Ile Asp Asn Phe Asn Gln Gln Gln Lys Lys Leu Gly Gly				
	1305	1310	1315	1320
caa gac att ttt atg aca gaa gaa cag aag aaa tac tat aat gca atg				4038
Gln Asp Ile Phe Met Thr Glu Glu Gln Lys Lys Tyr Tyr Asn Ala Met				
	1325	1330	1335	
aaa aaa tta gga tcc aaa aaa cct caa aaa ccc att cca cgg cct ctg				4086
Lys Lys Leu Gly Ser Lys Lys Pro Gln Lys Pro Ile Pro Arg Pro Leu				
	1340	1345	1350	
aac aaa tgt caa ggt ctc gtg ttc gac ata gtc aca agc cag atc ttt				4134
Asn Lys Cys Gln Gly Leu Val Phe Asp Ile Val Thr Ser Gln Ile Phe				
	1355	1360	1365	
gac atc atc atc ata agt ctc att atc cta aac atg att agc atg atg				4182
Asp Ile Ile Ile Ile Ser Leu Ile Ile Leu Asn Met Ile Ser Met Met				

1370	1375	1380	
gct gaa tca tac aac caa ccc aaa gcc atg aaa tcc atc ctt gac cat			4230
Ala Glu Ser Tyr Asn Gln Pro Lys Ala Met Lys Ser Ile Leu Asp His			
1385	1390	1395	1400
ctc aac tgg gtc ttt gtg gtc atc ttt acg tta gaa tgt ctc atc aaa			4278
Leu Asn Trp Val Phe Val Val Ile Phe Thr Leu Glu Cys Leu Ile Lys			
	1405	1410	1415
atc ttt gct ttg agg caa tac tac ttc acc aat ggc tgg aat tta ttt			4326
Ile Phe Ala Leu Arg Gln Tyr Tyr Phe Thr Asn Gly Trp Asn Leu Phe			
	1420	1425	1430
gac tgt gtg gtc gtg ctt ctt tcc att gtt agt aca atg att tct acc			4374
Asp Cys Val Val Val Leu Leu Ser Ile Val Ser Thr Met Ile Ser Thr			
	1435	1440	1445
ttg gaa aat cag gag cac att cct ttc cct ccg acg ctc ttc aga att			4422
Leu Glu Asn Gln Glu His Ile Pro Phe Pro Pro Thr Leu Phe Arg Ile			
	1450	1455	1460
gtc cgc ttg gct cgg att ggc cga atc ctg agg ctt gtc cgg gct gca			4470
Val Arg Leu Ala Arg Ile Gly Arg Ile Leu Arg Leu Val Arg Ala Ala			
	1465	1470	1475
cga gga atc agg act ctc ctc ttt gct ctg atg atg tcg ctt cct tct			4518
Arg Gly Ile Arg Thr Leu Leu Phe Ala Leu Met Met Ser Leu Pro Ser			
	1485	1490	1495
ctg ttc aac att ggt ctt cta ctc ttt ctg att atg ttt atc tat gcc			4566
Leu Phe Asn Ile Gly Leu Leu Leu Phe Leu Ile Met Phe Ile Tyr Ala			
	1500	1505	1510
att ctg ggt atg aac tgg ttt tcc aaa gtg aat cca gag tct gga atc			4614
Ile Leu Gly Met Asn Trp Phe Ser Lys Val Asn Pro Glu Ser Gly Ile			
	1515	1520	1525
gat gac ata ttc aac ttc aag act ttt gcc agc agc atg ctc tgt ctc			4662
Asp Asp Ile Phe Asn Phe Lys Thr Phe Ala Ser Ser Met Leu Cys Leu			
	1530	1535	1540
ttc cag ata agc aca tca gca ggt tgg gat tcc ctg ctc agc ccc atg			4710
Phe Gln Ile Ser Thr Ser Ala Gly Trp Asp Ser Leu Leu Ser Pro Met			
	1545	1550	1555
ctg cga tca aaa gaa tca tgt aac tct tcc tca gaa aac tgc cac ctc			4758
Leu Arg Ser Lys Glu Ser Cys Asn Ser Ser Ser Glu Asn Cys His Leu			
	1565	1570	1575
cct ggc ata gcc aca tcc tac ttt gtc agt tac att atc atc tcc ttt			4806
Pro Gly Ile Ala Thr Ser Tyr Phe Val Ser Tyr Ile Ile Ile Ser Phe			
	1580	1585	1590
ctc att gtt gtc aac atg tac att gct gtg att tta gag aac ttc aat			4854
Leu Ile Val Val Asn Met Tyr Ile Ala Val Ile Leu Glu Asn Phe Asn			

1595	1600	1605	
aca gcc act gaa gaa agt gag gac cct ttg ggt gaa gat gac ttt gac			4902
Thr Ala Thr Glu Glu Ser Glu Asp Pro Leu Gly Glu Asp Asp Phe Asp			
1610	1615	1620	
ata ttt tat gaa gtg tgg gaa aag ttt gac cca gaa gca aca caa ttt			4950
Ile Phe Tyr Glu Val Trp Glu Lys Phe Asp Pro Glu Ala Thr Gln Phe			
1625	1630	1635	1640
atc aaa tat tct gcc ctt tct gac ttt gct gat gcc ttg cct gag cct			4998
Ile Lys Tyr Ser Ala Leu Ser Asp Phe Ala Asp Ala Leu Pro Glu Pro			
1645	1650	1655	
ttg cgt gtc gca aag cca aat aaa tat caa ttt cta gta atg gac ttg			5046
Leu Arg Val Ala Lys Pro Asn Lys Tyr Gln Phe Leu Val Met Asp Leu			
1660	1665	1670	
ccc atg gtg agt gaa gat cgc ctc cac tgc atg gat att ctt ttc gcc			5094
Pro Met Val Ser Glu Asp Arg Leu His Cys Met Asp Ile Leu Phe Ala			
1675	1680	1685	
ttc acc gct agg gta ctc ggt ggc tct gat ggc cta gat agt atg aaa			5142
Phe Thr Ala Arg Val Leu Gly Gly Ser Asp Gly Leu Asp Ser Met Lys			
1690	1695	1700	
gca atg atg gaa gag aag ttc atg gaa gcc aat cct ctc aag aag ttg			5190
Ala Met Met Glu Glu Lys Phe Met Glu Ala Asn Pro Leu Lys Lys Leu			
1705	1710	1715	1720
tat gaa ccc ata gtc acc acc acc aag aga aag gaa gag gaa aga ggt			5238
Tyr Glu Pro Ile Val Thr Thr Thr Lys Arg Lys Glu Glu Glu Arg Gly			
1725	1730	1735	
gct gct att att caa aag gcc ttt cga aag tac atg atg aag gtg acc			5286
Ala Ala Ile Ile Gln Lys Ala Phe Arg Lys Tyr Met Met Lys Val Thr			
1740	1745	1750	
aag ggt gac caa ggt gac caa aat gac ttg gaa aac ggg cct cat tca			5334
Lys Gly Asp Gln Gly Asp Gln Asn Asp Leu Glu Asn Gly Pro His Ser			
1755	1760	1765	
cca ctc cag act ctt tgc aat gga gac ttg tct agc ttt ggg gtg gcc			5382
Pro Leu Gln Thr Leu Cys Asn Gly Asp Leu Ser Ser Phe Gly Val Ala			
1770	1775	1780	
aag ggc aag gtc cac tgt gac tgagccctca cctccacgcc tacctcatag			5433
Lys Gly Lys Val His Cys Asp			
1785	1790		
cttcacagcc ttgccttcag cctctgagct ccaggggtca gcagcttagt gtatcaacag			5493
ggagtggatt caccaaatta gccattccat tttcttttct ggctaaaata aatgatattt			5553
caatttcatt ttaaatgata cttacagaga tataagataa ggctacttga caaccagtgg			5613

tactattata ataaggaaga agacaccagg aaggactgta aaaggacata ccaatttttag 5673
 gattgaaata gttcaggccg ggcgcagtgg ctcatgcctg taatcccagc actttgagag 5733
 gccaaaggcag gtggatcacg aggtcaagag atcgagacca tcctggccaa catgatgaaa 5793
 ctccgtctct ctaaaaatac aaaaattagc tgggcacgtg ggcgtgcgcc tgtagtccca 5853
 ctacttg 5860

<210> 42

<211> 1791

<212> PRT

<213> Homo sapiens

<400> 42

Met	Asp	Asp	Arg	Cys	Tyr	Pro	Val	Ile	Phe	Pro	Asp	Glu	Arg	Asn	Phe	1	5	10	15
Arg	Pro	Phe	Thr	Ser	Asp	Ser	Leu	Ala	Ala	Ile	Glu	Lys	Arg	Ile	Ala	20	25	30	
Ile	Gln	Lys	Glu	Lys	Lys	Lys	Ser	Lys	Asp	Gln	Thr	Gly	Glu	Val	Pro	35	40	45	
Gln	Pro	Arg	Pro	Gln	Leu	Asp	Leu	Lys	Ala	Ser	Arg	Lys	Leu	Pro	Lys	50	55	60	
Leu	Tyr	Gly	Asp	Ile	Pro	Arg	Glu	Leu	Ile	Gly	Lys	Pro	Leu	Glu	Asp	65	70	75	80
Leu	Asp	Pro	Phe	Tyr	Arg	Asn	His	Lys	Thr	Phe	Met	Val	Leu	Asn	Arg	85	90	95	
Lys	Arg	Thr	Ile	Tyr	Arg	Phe	Ser	Ala	Lys	His	Ala	Leu	Phe	Ile	Phe	100	105	110	
Gly	Pro	Phe	Asn	Ser	Ile	Arg	Ser	Leu	Ala	Ile	Arg	Val	Ser	Val	His	115	120	125	
Ser	Leu	Phe	Ser	Met	Phe	Ile	Ile	Gly	Thr	Val	Ile	Ile	Asn	Cys	Val	130	135	140	
Phe	Met	Ala	Thr	Gly	Pro	Ala	Lys	Asn	Ser	Asn	Ser	Asn	Asn	Thr	Asp	145	150	155	160
Ile	Ala	Glu	Cys	Val	Phe	Thr	Gly	Ile	Tyr	Ile	Phe	Glu	Ala	Leu	Ile	165	170	175	
Lys	Ile	Leu	Ala	Arg	Gly	Phe	Ile	Leu	Asp	Glu	Phe	Ser	Phe	Leu	Arg	180	185	190	
Asp	Pro	Trp	Asn	Trp	Leu	Asp	Ser	Ile	Val	Ile	Gly	Ile	Ala	Ile	Val	195	200	205	

Ser Tyr Ile Pro Gly Ile Thr Ile Lys Leu Leu Pro Leu Arg Thr Phe
 210 215 220
 Arg Val Phe Arg Ala Leu Lys Ala Ile Ser Val Val Ser Arg Leu Lys
 225 230 235 240
 Val Ile Val Gly Ala Leu Leu Arg Ser Val Lys Lys Leu Val Asn Val
 245 250 255
 Ile Ile Leu Thr Phe Phe Cys Leu Ser Ile Phe Ala Leu Val Gly Gln
 260 265 270
 Gln Leu Phe Met Gly Ser Leu Asn Leu Lys Cys Ile Ser Arg Asp Cys
 275 280 285
 Lys Asn Ile Ser Asn Pro Glu Ala Tyr Asp His Cys Phe Glu Lys Lys
 290 295 300
 Glu Asn Ser Pro Glu Phe Lys Met Cys Gly Ile Trp Met Gly Asn Ser
 305 310 315 320
 Ala Cys Ser Ile Gln Tyr Glu Cys Lys His Thr Lys Ile Asn Pro Asp
 325 330 335
 Tyr Asn Tyr Thr Asn Phe Asp Asn Phe Gly Trp Ser Phe Leu Ala Met
 340 345 350
 Phe Arg Leu Met Thr Gln Asp Ser Trp Glu Lys Leu Tyr Gln Gln Thr
 355 360 365
 Leu Arg Thr Thr Gly Leu Tyr Ser Val Phe Phe Phe Ile Val Val Ile
 370 375 380
 Phe Leu Gly Ser Phe Tyr Leu Ile Asn Leu Thr Leu Ala Val Val Thr
 385 390 395 400
 Met Ala Tyr Glu Glu Gln Asn Lys Asn Val Ala Ala Glu Ile Glu Ala
 405 410 415
 Lys Glu Lys Met Phe Gln Glu Ala Gln Gln Leu Leu Lys Glu Glu Lys
 420 425 430
 Glu Ala Leu Val Ala Met Gly Ile Asp Arg Ser Ser Leu Thr Ser Leu
 435 440 445
 Glu Thr Ser Tyr Phe Thr Pro Lys Lys Arg Lys Leu Phe Gly Asn Lys
 450 455 460
 Lys Arg Lys Ser Phe Phe Leu Arg Glu Ser Gly Lys Asp Gln Pro Pro
 465 470 475 480
 Gly Ser Asp Ser Asp Glu Asp Cys Gln Lys Lys Pro Gln Leu Leu Glu
 485 490 495
 Gln Thr Lys Arg Leu Ser Gln Asn Leu Ser Leu Asp His Phe Asp Glu
 500 505 510

His Gly Asp Pro Leu Gln Arg Gln Arg Ala Leu Ser Ala Val Ser Ile
 515 520 525
 Leu Thr Ile Thr Met Lys Glu Gln Glu Lys Ser Gln Glu Pro Cys Leu
 530 535 540
 Pro Cys Gly Glu Asn Leu Ala Ser Lys Tyr Leu Val Trp Asn Cys Cys
 545 550 555 560
 Pro Gln Trp Leu Cys Val Lys Lys Val Leu Arg Thr Val Met Thr Asp
 565 570 575
 Pro Phe Thr Glu Leu Ala Ile Thr Ile Cys Ile Ile Ile Asn Thr Val
 580 585 590
 Phe Leu Ala Met Glu His His Lys Met Glu Ala Ser Phe Glu Lys Met
 595 600 605
 Leu Asn Ile Gly Asn Leu Val Phe Thr Ser Ile Phe Ile Ala Glu Met
 610 615 620
 Cys Leu Lys Ile Ile Ala Leu Asp Pro Tyr His Tyr Phe Arg Arg Gly
 625 630 635 640
 Trp Asn Ile Phe Asp Ser Ile Val Ala Leu Leu Ser Phe Ala Asp Val
 645 650 655
 Met Asn Cys Val Leu Gln Lys Arg Ser Trp Pro Phe Leu Arg Ser Phe
 660 665 670
 Arg Val Leu Arg Val Phe Lys Leu Ala Lys Ser Trp Pro Thr Leu Asn
 675 680 685
 Thr Leu Ile Lys Ile Ile Gly Asn Ser Val Gly Ala Leu Gly Ser Leu
 690 695 700
 Thr Val Val Leu Val Ile Val Ile Phe Ile Phe Ser Val Val Gly Met
 705 710 715 720
 Gln Leu Phe Gly Arg Ser Phe Asn Ser Gln Lys Ser Pro Lys Leu Cys
 725 730 735
 Asn Pro Thr Gly Pro Thr Val Ser Cys Leu Arg His Trp His Met Gly
 740 745 750
 Asp Phe Trp His Ser Phe Leu Val Val Phe Arg Ile Leu Cys Gly Glu
 755 760 765
 Trp Ile Glu Asn Met Trp Glu Cys Met Gln Glu Ala Asn Ala Ser Ser
 770 775 780
 Ser Leu Cys Val Ile Val Phe Ile Leu Ile Thr Val Ile Gly Lys Leu
 785 790 795 800
 Val Val Leu Asn Leu Phe Ile Ala Leu Leu Leu Asn Ser Phe Ser Asn
 805 810 815

Glu Glu Arg Asn Gly Asn Leu Glu Gly Glu Ala Arg Lys Thr Lys Val
 820 825 830
 Gln Leu Ala Leu Asp Arg Phe Arg Arg Ala Phe Cys Phe Val Arg His
 835 840 845
 Thr Leu Glu His Phe Cys His Lys Trp Cys Arg Lys Gln Asn Leu Pro
 850 855 860
 Gln Gln Lys Glu Val Ala Gly Gly Cys Ala Ala Gln Ser Lys Asp Ile
 865 870 875 880
 Ile Pro Leu Val Met Glu Met Lys Arg Gly Ser Glu Thr Gln Glu Glu
 885 890 895
 Leu Gly Ile Leu Thr Ser Val Pro Lys Thr Leu Gly Val Arg His Asp
 900 905 910
 Trp Thr Trp Leu Ala Pro Leu Ala Glu Glu Glu Asp Asp Val Glu Phe
 915 920 925
 Ser Gly Glu Asp Asn Ala Gln Arg Ile Thr Gln Pro Glu Pro Glu Gln
 930 935 940
 Gln Ala Tyr Glu Leu His Gln Glu Asn Lys Lys Pro Thr Ser Gln Arg
 945 950 955 960
 Val Gln Ser Val Glu Ile Asp Met Phe Ser Glu Asp Glu Pro His Leu
 965 970 975
 Thr Ile Gln Asp Pro Arg Lys Lys Ser Asp Val Thr Ser Ile Leu Ser
 980 985 990
 Glu Cys Ser Thr Ile Asp Leu Gln Asp Gly Phe Gly Trp Leu Pro Glu
 995 1000 1005
 Met Val Pro Lys Lys Gln Pro Glu Arg Cys Leu Pro Lys Gly Phe Gly
 1010 1015 1020
 Cys Cys Phe Pro Cys Cys Ser Val Asp Lys Arg Lys Pro Pro Trp Val
 1025 1030 1035 1040
 Ile Trp Trp Asn Leu Arg Lys Thr Cys Tyr Gln Ile Val Lys His Ser
 1045 1050 1055
 Trp Phe Glu Ser Phe Ile Ile Phe Val Ile Leu Leu Ser Ser Gly Ala
 1060 1065 1070
 Leu Ile Phe Glu Asp Val His Leu Glu Asn Gln Pro Lys Ile Gln Glu
 1075 1080 1085
 Leu Leu Asn Cys Thr Asp Ile Ile Phe Thr His Ile Phe Ile Leu Glu
 1090 1095 1100
 Met Val Leu Lys Trp Val Ala Phe Gly Phe Gly Lys Tyr Phe Thr Ser
 1105 1110 1115 1120

Ala Trp Cys Cys Leu Asp Phe Ile Ile Val Ile Val Ser Val Thr Thr
 1125 1130 1135
 Leu Ile Asn Leu Met Glu Leu Lys Ser Phe Arg Thr Leu Arg Ala Leu
 1140 1145 1150
 Arg Pro Leu Arg Ala Leu Ser Gln Phe Glu Gly Met Lys Val Val Val
 1155 1160 1165
 Asn Ala Leu Ile Gly Ala Ile Pro Ala Ile Leu Asn Val Leu Leu Val
 1170 1175 1180
 Cys Leu Ile Phe Trp Leu Val Phe Cys Ile Leu Gly Val Tyr Phe Phe
 1185 1190 1195 1200
 Ser Gly Lys Phe Gly Lys Cys Ile Asn Gly Thr Asp Ser Val Ile Asn
 1205 1210 1215
 Tyr Thr Ile Ile Thr Asn Lys Ser Gln Cys Glu Ser Gly Asn Phe Ser
 1220 1225 1230
 Trp Ile Asn Gln Lys Val Asn Phe Asp Asn Val Gly Asn Ala Tyr Leu
 1235 1240 1245
 Ala Leu Leu Gln Val Ala Thr Phe Lys Gly Trp Met Asp Ile Ile Tyr
 1250 1255 1260
 Ala Ala Val Asp Ser Thr Glu Lys Glu Gln Gln Pro Glu Phe Glu Ser
 1265 1270 1275 1280
 Asn Ser Leu Gly Tyr Ile Tyr Phe Val Val Phe Ile Ile Phe Gly Ser
 1285 1290 1295
 Phe Phe Thr Leu Asn Leu Phe Ile Gly Val Ile Ile Asp Asn Phe Asn
 1300 1305 1310
 Gln Gln Gln Lys Lys Leu Gly Gly Gln Asp Ile Phe Met Thr Glu Glu
 1315 1320 1325
 Gln Lys Lys Tyr Tyr Asn Ala Met Lys Lys Leu Gly Ser Lys Lys Pro
 1330 1335 1340
 Gln Lys Pro Ile Pro Arg Pro Leu Asn Lys Cys Gln Gly Leu Val Phe
 1345 1350 1355 1360
 Asp Ile Val Thr Ser Gln Ile Phe Asp Ile Ile Ile Ile Ser Leu Ile
 1365 1370 1375
 Ile Leu Asn Met Ile Ser Met Met Ala Glu Ser Tyr Asn Gln Pro Lys
 1380 1385 1390
 Ala Met Lys Ser Ile Leu Asp His Leu Asn Trp Val Phe Val Val Ile
 1395 1400 1405
 Phe Thr Leu Glu Cys Leu Ile Lys Ile Phe Ala Leu Arg Gln Tyr Tyr
 1410 1415 1420

Phe Thr Asn Gly Trp Asn Leu Phe Asp Cys Val Val Val Leu Leu Ser
 1425 1430 1435 1440
 Ile Val Ser Thr Met Ile Ser Thr Leu Glu Asn Gln Glu His Ile Pro
 1445 1450 1455
 Phe Pro Pro Thr Leu Phe Arg Ile Val Arg Leu Ala Arg Ile Gly Arg
 1460 1465 1470
 Ile Leu Arg Leu Val Arg Ala Ala Arg Gly Ile Arg Thr Leu Leu Phe
 1475 1480 1485
 Ala Leu Met Met Ser Leu Pro Ser Leu Phe Asn Ile Gly Leu Leu Leu
 1490 1495 1500
 Phe Leu Ile Met Phe Ile Tyr Ala Ile Leu Gly Met Asn Trp Phe Ser
 1505 1510 1515 1520
 Lys Val Asn Pro Glu Ser Gly Ile Asp Asp Ile Phe Asn Phe Lys Thr
 1525 1530 1535
 Phe Ala Ser Ser Met Leu Cys Leu Phe Gln Ile Ser Thr Ser Ala Gly
 1540 1545 1550
 Trp Asp Ser Leu Leu Ser Pro Met Leu Arg Ser Lys Glu Ser Cys Asn
 1555 1560 1565
 Ser Ser Ser Glu Asn Cys His Leu Pro Gly Ile Ala Thr Ser Tyr Phe
 1570 1575 1580
 Val Ser Tyr Ile Ile Ile Ser Phe Leu Ile Val Val Asn Met Tyr Ile
 1585 1590 1595 1600
 Ala Val Ile Leu Glu Asn Phe Asn Thr Ala Thr Glu Glu Ser Glu Asp
 1605 1610 1615
 Pro Leu Gly Glu Asp Asp Phe Asp Ile Phe Tyr Glu Val Trp Glu Lys
 1620 1625 1630
 Phe Asp Pro Glu Ala Thr Gln Phe Ile Lys Tyr Ser Ala Leu Ser Asp
 1635 1640 1645
 Phe Ala Asp Ala Leu Pro Glu Pro Leu Arg Val Ala Lys Pro Asn Lys
 1650 1655 1660
 Tyr Gln Phe Leu Val Met Asp Leu Pro Met Val Ser Glu Asp Arg Leu
 1665 1670 1675 1680
 His Cys Met Asp Ile Leu Phe Ala Phe Thr Ala Arg Val Leu Gly Gly
 1685 1690 1695
 Ser Asp Gly Leu Asp Ser Met Lys Ala Met Met Glu Glu Lys Phe Met
 1700 1705 1710
 Glu Ala Asn Pro Leu Lys Lys Leu Tyr Glu Pro Ile Val Thr Thr Thr
 1715 1720 1725

Lys Arg Lys Glu Glu Glu Arg Gly Ala Ala Ile Ile Gln Lys Ala Phe
 1730 1735 1740

Arg Lys Tyr Met Met Lys Val Thr Lys Gly Asp Gln Gly Asp Gln Asn
 1745 1750 1755 1760

Asp Leu Glu Asn Gly Pro His Ser Pro Leu Gln Thr Leu Cys Asn Gly
 1765 1770 1775

Asp Leu Ser Ser Phe Gly Val Ala Lys Gly Lys Val His Cys Asp
 1780 1785 1790

<210> 43

<211> 8

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: protein seq.
 basis for rat NaN reverse primer no. 5

<400> 43

Ala Trp Cys Trp Leu Asp Phe Leu
 1 5

<210> 44

<211> 23

<212> DNA

<213> Artificial Sequence

<220>

<223> human NaN reverse primer


<400> 44

gtgccgtaaa catgagactg tcg

23

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US00/19342

A. CLASSIFICATION OF SUBJECT MATTER IPC(7) :C07K 14/705; C12N 5/10, 15/12, 15/63 US CL :435/320.1, 325; 530/350; 536/23.5 According to International Patent Classification (IPC) or to both national classification and IPC		
B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) U.S. : 435/320.1, 325; 530/350; 536/23.5 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) BRS, STN, MEDLINE, GENESEQ, PIR, SWISS-PROT, STREMBL, GENEMBL search terms: sodium channel, tetrodotoxin		
C. DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X - A	WO 97/01577 A1 (UNIVERSITY COLLEGE LONDON) 16 January 1997, see especially pages 85-93.	1-6, 8-9, 22, 25 ----- 7, 33-35
<input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/> See patent family annex.		
* Special categories of cited documents:		
"A" document defining the general state of the art which is not considered to be of particular relevance	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention	
"B" earlier document published on or after the international filing date	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone	
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art	
"O" document referring to an oral disclosure, use, exhibition or other means	"G" document member of the same patent family	
"P" document published prior to the international filing date but later than the priority date claimed		
Date of the actual completion of the international search 10 SEPTEMBER 2000	Date of mailing of the international search report 02 NOV 2000	
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230	Authorized officer  MICHAEL PAK Telephone No. (703) 308-0196	

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US00/19342

Box I Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
1-9, 22, 25, 33-35

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US00/19342

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

Group I, claim(s) 1-9, 22, 25, 33-35, drawn to an isolated nucleic acid molecule, a method of producing a transformed cell, a Na⁺ channel and a method of producing recombinant NaN protein.

Group II, claim(s) 10-14, drawn to a method to identify an agent that modulates the activity of the Na⁺ channel.

Group III, claim(s) 15, drawn to a method to identify an agent that modulates the transcription or translation of mRNA.

Group IV, claim(s) 16, drawn to a method to treat pain by administering an agent that modulates the activity of the Na⁺ channel.

Group V, claim(s) 17, drawn to a method to treat pain by administering an agent that modulates the transcription or translation of mRNA.

Group VI, claim(s) 18, drawn to an isolated nucleic acid that is antisense.

Group VII, claim(s) 19, drawn to a scintigraphic method to image loci of pain generation.

Group VIII, claim(s) 20, drawn to a method to identify tissues by detecting NaN.

Group IX, claim(s) 21, drawn to a method to identify tissues by detecting mRNA.

Group X, claim(s) 23-24, drawn to an antibody.

Group XI, claim(s) 26-27 and 36-37, drawn to a therapeutic composition comprising an agent and a method of treatment using the agent of claim 26.

Group XII, claim(s) 28, drawn to a method to screen candidate compound for use in treating pain.

Group XIII, claim(s) 29-32, drawn to a chimeric NaN channel and the DNA encoding the chimeric NaN channel.

The inventions listed as Groups I-XIII do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features because the product of claim 1 is anticipated by UNIVERSITY COLLEGE LONDON (WO 97/01577A1 16 January 1997) and thus, does not share a special technical feature with any other group.

The products of Group VI, X-XI, and XIII does not share the same or corresponding special technical feature with Group I, because they are drawn to products having materially different structures and functions, and each defines a separate invention over the art.

The methods of Groups II-V, VII-IX, and XII do not share the same or corresponding special technical feature with Group I, because the methods have materially different process steps and are practiced for materially different purposes, and each defines a separate invention over the art.

Since Groups I-XIII do not share a special technical feature, unity of invention is lacking.